

1 **Chapter 12**

2 ***Phytoremediation using aquatic plants***

3 **12.1 Introduction**

4

5 Freshwaters are affected by a diverse range of pollutants which increases the demand for
6 effective remediation. Aquatic phytoremediation is a nature-based solution that has the potential to
7 provide efficient, spatially adaptable and multi-targeted treatment of polluted waters using the
8 ability of macrophytes to take-up, sequester and degrade pollutants. This chapter considers the
9 primary phytoremediation mechanisms that macrophytes employ to remove inorganic, organic and
10 biological waterborne pollutants before highlighting some of the common macrophyte accumulators
11 that have been studied. Three common macrophyte planting systems (i) constructed wetlands
12 (CWs), (ii) wild macrophyte planting/harvesting and (iii) floating treatment wetlands (FTWs), are
13 considered to understand how macrophytes are deployed for targeted aquatic phytoremediation.

14 Important practical considerations for implementing aquatic phytoremediation include the
15 use of invasive species, the optimal harvesting time and frequency for pollutant removal with
16 macrophyte biomass, and the full extent of the role that microbial biofilms play in phytoremediation.
17 In this chapter, these issues are unpacked and recommendations for future programmes of research
18 and development are made. Finally, the opportunities to generate 'added value' from expanding
19 aquatic phytoremediation in terms of the provision of ecosystem services and the potential for
20 resource recovery are outlined.

21 **12.2 Water contamination and water security**

22

23 Surface waters are vital for supporting people and ecosystems; however, freshwater
24 availability is under increasing pressure due to a growing human population requiring access to safe

25 water(Heathwaite, 2010). Global freshwater resources comprise 2.5% of the total global water
26 budget, although only 0.0072% (93,120km³) of the total global waters are available for drinking,
27 energy, food production and the industry sector(Lawford et al., 2013; Zimmerman et al.,
28 2008).Tilman *et al.*(2011) predicts that crop production will need to increase by 100-110% by 2050
29 to feed the growing population, leading to a global freshwater deficit of approximately 2,400km³ per
30 year (Rockström et al., 2014).

31 Many surface waters are currently of sub-optimal standards due to a range of stressors
32 impacting freshwaters such as point source and diffuse pollution, land-use change and climate
33 change, which further compounds the challenge of providing water security (Ormerod *et al.*, 2010;
34 Berger *et al.*, 2017).One of the major pressures on water quality in the United Kingdom is nutrient
35 enrichment from diffuse pollution (Ulénet al., 2007), whereas elsewhere in countries such as
36 China,additional issues of heavy metal pollution are also prominent (Cheng, 2003). Interactions
37 between different stressors in space and time can also lead to additive effects(Heathwaite, 2010),
38 for example, increased land-use change towards intensive agriculture and a potential increase in
39 storm frequency may increase the delivery of nitrogen (N) phosphorus (P) and fine sedimentto
40 receiving water(Dunn et al., 2012).

41 Table 12.1 summarises the surface water pollutants that are of concern and where
42 remediation solutions are being developed. Water pollutants can be broadly categorised as either:
43 organic, e.g. hydrocarbons, pesticides and algal toxins, orinorganic,e.g. metals or syntheticand
44 manure-based fertilisers containing excess amounts of N and P,or biological,e.g. pathogens and algal
45 toxins.The mobilisation and effects of different pollutants have been discussed extensively
46 elsewhere (Heisler et al., 2008; Ohe et al., 2004; Liess & Carsten Von Der Ohe, 2005; Edwards, 2015;
47 Lintelmann et al., 2003). However, different pollutants may have multiple sources, for example, N
48 and P can be released from agriculture, aquaculture and urban waste water streams.

49 Managing waterborne pollutants through *in-situ* best management practices (BMPs) that
50 target the source of pollution is the principal approach to improving water quality(Lam et al., 2011).
51 However, lag times associated with the improvement of water quality and subsequent ecological
52 recovery of receiving watersfollowing mitigation may range from 1 to >50 years(Meals et al., 2010).
53 The 'legacy effect' is one such component delaying water quality improvements in spite of BMPs
54 being in place(Haygarth et al., 2014). Water bodies, such as those with long residence times, may
55 become reservoirs for pollutants over time, meaning that although source management is in place,
56 the receiving waters remains high in pollutant levels for significant amounts of time(Meals et al.,
57 2010). Therefore, developing management systemsthat combine BMPs with other methods of
58 remediating waters with high levels of pollutants, both at source and throughout the catchment,is
59 needed to sustainablyimprove water quality.

60 The pollution of water with inorganic elements such as N, P and metals also provides an
61 opportunity to recover elements as part of a 'circular economy' approach(Masi et al., 2017; Quilliam
62 et al., 2015). Energy-intensive mining for macronutrients such as P and potassium (K) are exhausting
63 finite supplies of nutrientsfor the production of agricultural fertilisers(Jones et al., 2013), whilst
64 liquidfertilisers and nutrient-rich solid manures applied to agricultural land are readily transferred to
65 receiving waters.Coupling systems that remediate water pollution and enable the capture of these
66 resources may help close the loop on nutrient loss (Quilliam et al., 2015). Therefore, macrophyte
67 phytoremediation has the potential to be employed for both the sustainable remediation of surface
68 waters and as a management strategy for recovering nutrients.

69 12.3 Aquatic phytoremediation

70

71 Aquatic phytoremediation is a phytotechnology used for the removal of pollutants from
72 surface waters and the restoration of impacted water bodies (rivers, streams, lakes, ponds). Within
73 surface waters plants can be cultured to remove pollutants from both the water column and the
74 sediment (Newete & Byrne, 2016; Miretzky et al., 2004), and can be deployed at either the point
75 source, or within water bodies where diffuse pollution is problematic (Lu *et al.*, 2011). Aquatic
76 phytoremediation specifically uses macrophytes (i.e. freshwater adapted angiosperms, pteridophytes
77 and ferns) for removing and degrading pollutants within aquatic environments (Rai, 2009). This
78 definition does not include microalgae species. Macrophytes can be broadly classified into three
79 primary growth forms: floating, submerged and emergent (Figure 12.1). Floating macrophytes
80 occupy the water surface and include genera such as *Lemna* (duckweeds), *Hydrocharis* (frogbit) and
81 *Nymphaea* (water-lilies) which may be free-floating or rooted. Submerged macrophytes grow
82 primarily below the water surface and may be anchored to the substrate, although *Ceratophyllum*
83 (hornworts) are a widespread genus of unrooted submerged plants. Emergent macrophytes occupy
84 the margins of water bodies and are rooted into the substrate but have significant shoot growth
85 above the water level, e.g. *Typha* (reedmace) and *Phragmites* (common reed). These different growth
86 forms facilitate the removal of pollutants from both the water column and the sediment depending
87 on the way in which they are deployed (Newete and Byrne, 2016).

88

89 Macrophytes have significant capacity for uptake of nutrients and other substances from
90 their growth medium, and can thus lower the pollution concentration of a target water body (Dhote
91 and Dixit, 2009). Macrophytes can remove and degrade pollutants using the key mechanisms of
92 rhizo/phyto-filtration, phytoextraction, phytovolatilization and phytodegradation (Table 12.2).
93 Emergent and floating macrophytes primarily take up nutrients and other contaminants (whether
94 from the substrate or water column) through their roots, whereas stem tissue can also be an
95 important pathway for removal from the water column for submerged macrophytes (Denny, 1972;

96 Gabrielson, Perkins and Welch, 1984; Dhote and Dixit, 2009). Specific mechanisms for pollutant
97 removal and degradation by macrophytes depend primarily on the type of pollutant (nutrient, heavy
98 metals, organic pollutants, biological), and the location of the pollutant within the surface water
99 body (water column, lake or streambed sediment)(Miretzky, Saralegui and Cirelli, 2004;
100 Padmavathiamma and Li, 2007; Vymazal, 2011; Xing *et al.*, 2013; McAndrew, Ahn and Spooner,
101 2016; Polechońska and Samecka-Cymerman, 2016).Different mechanisms for removing various
102 classes of pollutant from surface water systems by macrophytes are considered below.

103 **Macronutrients**

104 It is important to note that elements targeted for phytoremediation may exist in a dissolved
105 phase, or in a particulate phase adhered to suspended material in the water column or bound to
106 sediment, which means there are different mechanisms for removal(Perk, 2006).Macronutrients,
107 including N and P, are essential elements required in relatively large concentrations for plant
108 metabolism (Hawkesford *et al.*, 2011). Therefore, when aquatic system are enriched with N and
109 P,phytoextraction (uptake and sequestration) is an important mechanism(Eid *et al.* 2012;
110 Mkandawire & Dudel, 2005). Particulate pollutants in the water column, such as P,can be stabilised
111 by phytofiltration(Tanner and Headley, 2011a; Olguín and Sá Nchez-Galvá, 2012), whereplant roots
112 may excrete exudates that assist phytoextraction of adsorbed elements(Jackson, 1998; Verkleij *et*
113 *al.*, 2009; Akeel, 2013). For N removal, phytodegradation may also be important in the water
114 column and sedimentas the oxygen and energy supplied to the root zone from macrophytes may
115 support nutrient-degrading microbial communities, including the simultaneouspresence ofboth
116 nitrifyingand denitrifying bacteria (Table 12.2)(Lu *et al.*, 2018).

117 **Micronutrients/metals**

118 Micronutrients are essential elements that are required by plants in relatively small
119 quantities, e.g. to regulate redox reactions, metabolism and cell integrity (Broadley *et al.*, 2011).
120 Essential micronutrients include iron (Fe), manganese (Mn), copper (Cu), zinc (Zn), molybdenum

121 (Mn) and boron (B); beneficial but non-essential micronutrients include sodium (Na), silicon (Si),
122 cobalt (Co), selenium (Se); while there are elements that can be found in plant tissue but are not
123 thought to be beneficial such as aluminium (Al) vanadium (V), titanium (Ti), lanthanum (La) and
124 cerium (Ce) (Broadley et al., 2011) (Table 12.1). Some of these elements may be enriched by industrial
125 pollution but can be reduced by phytoextraction through repeated harvesting of plant tissue,
126 following uptake in the water column through hydroponic growth (e.g. in FTWs) or where plants are
127 rooted in sediment (Ali et al., 2013) (Figure 12.2). The efficiency of phytoextraction as a
128 phytoremediation strategy depends upon the specific degree of essentiality of each element for plant
129 metabolism and is determined by specific mechanisms for uptake and translocation into plant
130 tissue (Dhir, 2013). Hyperaccumulators are plants that have a high affinity for certain elements and
131 through enhanced phytoextraction can sequester high concentrations of metals (Sarma, 2011; van
132 der Ent et al., 2013). Phytofiltration is important for soluble and particulate pollutants with
133 absorption/adsorption to plant roots (Olguín and Sá Nchez-Galva, 2012), and in some cases metals
134 can be bound and/or precipitated on the plant roots (Xian et al., 2010; Gomes et al., 2016) (Figure
135 12.2).

136 **Organic pollutants**

137 Organic pollutants are compounds containing carbon that are primarily synthetic,
138 environmentally persistent and potentially toxic. They include products such as pesticides, solvents
139 and pharmaceuticals and personal care products (PPCPs) (El-Shahawi et al., 2010) (Table 12.1).
140 Phytometabolism and rhizodegradation within the water column and sediment are integral
141 processes in the aquatic phytoremediation of organic compounds (Reinhold et al., 2010).
142 Phytometabolism can occur if organic compounds are more hydrophilic meaning they pass more
143 readily through the plant epidermis into plant cells (Lintelmann et al., 2003; Dettenmaier, Doucette
144 and Bugbee, 2009; Yamazaki et al., 2015) (Figure 12.2). Sequestered compounds undergo chemical
145 modification through oxidation, reduction or hydrolysis which makes them chemically more reactive

146 within plant cells; the less harmful metabolite is then conjugated/bound to sugars, amino acids or
147 glutathione to reduce its toxicity and hydrophobicity (Macek et al., 2000; Geissen et al., 2015).
148 These bound metabolites may then be either stored within the vacuole or excreted from the plant, or
149 can become insoluble by being covalently bound within the cell wall (Zhang et al., 2014).
150 Rhizodegradation can take place within sediment, and more hydrophobic compounds can serve as a
151 microbial carbon source where emergent macrophytes supply oxygen to the root zone (Figure 12.2).
152 The advantage of these two phytoremediation processes is that there is no need for repeated
153 harvests to extract the pollutant and thus disturbance to the aquatic system is reduced.

154 **Microbial pollutants**

155 Microbial water pollutants such as the bacteria *Escherichia coli* O157, the protozoan parasite
156 *Cryptosporidium* spp. and viruses such as norovirus can cause harm to humans and animals (Haack
157 et al., 2016; Fuhrmann et al., 2017) (Table 12.1). The ability of plants to directly take up microbial
158 pollutants is limited; however, there are some accounts of pathogens entering plant tissue through
159 the process of internalisation, although whether this is an active or passive process is unclear and
160 likely depends on the type of pathogen, plant and the local abiotic conditions (Hirneisen et al., 2012).
161 The primary mechanisms for removal of microbial pollutants from water are either, chemical, e.g.
162 oxidation, photodegradation, exposure to plant root biocides and adsorption to organic material and
163 biofilms; physical, e.g. through filtration and sedimentation; or biological, e.g. predation, natural die-
164 off, antibiosis and other biolytic processes (Decamp and Warren, 2000; Karathanasis et al., 2003;
165 Karim et al., 2004; Wand et al., 2006; Makvana and Sharma, 2013). Macrophyte planting systems,
166 particularly CWs, may promote these mechanisms and thus facilitate the degradation of microbial
167 pollutants.

168 **12.4 Macrophytes used in aquatic phytoremediation**

169

170 **12.4.1 Macronutrients**

171

172 Macrophytes uptake and sequester N primarily in the form of nitrate (NO_3^-) and ammonium (NH_4^+),
173 while P is taken up as phosphate (PO_4^{4-}). Studies vary in their focus on total amounts (i.e. including
174 particulate) versus the dissolved fraction of macronutrients, which makes comparing optimal
175 macrophyte accumulator species challenging (Table 12.3). Macrophytes that have the greatest
176 biomass production and/or fastest growth rates are some of the most effective
177 nutrient phyto-remediators (Keenen and Kirkwood, 2015), for example, *Eichhornia crassipes*, *Lemna*
178 *sp.* and *Typha latifolia* have growth rates of 60-110 t/ha/yr, 6-26 t/ha/yr and 8-61 t/ha/yr,
179 respectively (Gumbrecht, 1993).

180 Emergent species have received considerable attention in nutrient phyto-remediation and are
181 often deployed in CWs, with *Canna* spp. and *Cyperus* spp. showing some of the highest removal
182 efficiencies for ammonium (NH_4^+) of between 74-100% (Table 12.3). *Typha latifolia*, *Lolium*
183 *multiflorum* and *Polygonum hydropiperoides* showed high TP removal efficiency of 81-90% (Table
184 12.3). For floating macrophytes *Eichhornia crassipes*, *Lemna gibba* and *Pistia stratiotes* show good
185 potential for nutrient removal: *E. crassipes* can remove up to 92% NO_3^- and 81% NH_3^- whilst *L.*
186 *gibba* can remove 100% NO_3^- and 82% NH_3^- (Table 12.3). The same two species were also effective at
187 removing total phosphorus (TP) (Table 12.3). Submerged plants have received less attention for their
188 nutrient phyto-remediation capacity (Table 12.3). This may reflect the difficulty of cultivating and
189 harvesting submerged macrophytes, and the potentially lower biomass generated compared to
190 emergent plants (Du et al., 2017). *Ceratophyllum demersum* and *Myriophyllum aquaticum* are
191 potential candidates for the targeting of total nitrogen (TN) and TP with removal rates >41% (Table
192 12.3). *Potamogeton crispus* was deployed as part of a hybrid FTW experiment and was found to have
193 enhanced effects over the FTW comprised of only emergent plants; however, the individual removal
194 contribution from *P. crispus* was not quantified (Guo et al., 2014). Most submerged species are
195 rooted in sediment and may also remove nutrients from the water column through foliar absorption
196 (Eichert and Fernández, 2011). Hence they offer the dual ability to remove nutrients from water and
197 sediment, allowing the simultaneous remediation of sediments that have a pollutant legacy and

198 which may continue to release nutrients to the water column via internal loading even after external
199 loads have been reduced. However, the disturbance caused during harvesting can re-
200 suspend sediment-bound elements, and alter the macrophyte-equilibrium state to a potentially
201 undesirable phytoplankton-dominated state (Kuiper et al., 2017).

202 The phytoremediation potential of a macrophyte is influenced by biotic factors such as
203 competition, predation and developmental stage (Quilliam et al., 2015), and abiotic factors such as
204 temperature, pH, light availability, seasonality and nutrient loading (Ansari et al, 2014). For example,
205 Ayyasamy et al. (2009) found that the removal efficiency of by *E. crassipes* increased between
206 concentrations of 100 mg/l to 300 mg/l of NO_3^- , but decreased at higher concentrations of 400 and
207 500 mg/l of NO_3^- . Similarly, a mesocosm-based study of the effect of different temperature regimes
208 on N and P removal by *Nasturtium officinale* and *Oenanthe javanica* found that maximum net
209 accumulation of TN and TP occurred at an air temperature of 22°C but deteriorated thereafter (Hu et
210 al., 2010). Given the wide range of factors that may influence the ability of macrophytes to remove
211 contaminants, understanding the performance of some of the key macrophyte accumulators under
212 different environmental conditions is prudent in order to optimise species selection.

213 12.4.2 Metals

214
215 Macrophytes can also remove micronutrients (henceforth referred to as metals (Rai, 2009)) from
216 water and sediments, and hyperaccumulators are most appropriate for the phytoremediation of
217 metals (Ali et al., 2013). The search for hyperaccumulator species has been one of the primary foci
218 within the field given the widespread prevalence of past and current metal industrial effluents and
219 the ecological risks they carry (van der Ent et al., 2013); however, metal bioavailability can be reduced
220 by sedimentation and adsorption to clay particles (Kumar et al., 2008). Studies based on mesocosm-
221 scale CW experiments have been carried out on synthetic solutions with elevated metal
222 concentrations in domestic and industrial wastewaters to assess the potential of macrophytes of
223 different growth forms to act as hyperaccumulators (Fu & Wang, 2011; Kamal et al., 2004; Rai, 2009;

224 Rezania et al., 2016)(Table 12.4). Many species also have the capacity to take up multiple types of
225 metals meaning that some species could be more beneficial in phytoremediation (Table 12.4).

226 Macrophytes that have often been cited as hyperaccumulators with high biomass potential are
227 free-floating plants, such as members of the Lemnaceae (e.g. *Lemna minor*), *Pista stratiotes*,
228 *Eichhornia crassipes* and those from the genera *Salvinia* (Table 12.4). For example, *L. gibba* has been
229 reported to concentrate between 14,000mg/kg dry weight of Cd, whilst *E. crassipes* can concentrate
230 10,000mg/kg Zn (Low et al., 1994; Mkandawire et al., 2004). Furthermore, *Typha latifolia* and
231 *Ceratophyllum demersum* L. have also shown good potential (Osmolovskaya & Kurilenko, 2005;
232 Sunita et al., 2015). The main limitation for macrophyte metal uptake is the toxicity of the target
233 metal pollutant at higher concentrations (Landesman et al., 2011). However, detoxification
234 mechanisms also allow species to avoid the negative effects of these metals (Deng et al., 2004); for
235 example, more than 50% of the Ca, Cd, Co, Fe, Mg, Mn, and Zn recovered in the roots of *Pistia*
236 *stratiotes* were actually attached to the external surfaces indicating the ability of the plant to exclude
237 metals and thus maintain tolerable levels internally (Lu et al., 2011). Newete & Byrne (2016) also
238 state that the extent of the root system affects the ability of macrophytes to remove metal
239 pollutants, with fibrous root systems being superior due to their large surface area. Physio-chemical
240 factors are also important for uptake and accumulation of metals with temperature, light, pH and
241 salinity all having been shown to influence remediation performance (Rai, 2009).

242

243 12.4.3 Organic pollutants

244

245

246 Table 12.5 shows the wide range of studies that have been carried out in relation to the
247 phytoremediation of organic pollutants and some of the key macrophytes that may be utilised. For
248 pesticides, *Lemna minor* removed 95% of 2,4,5-trichlorophenol, whereas for isoproturon and
249 glyphosate *L. minor*'s removal efficiency was poor (25% and 8% respectively; Table 12.5). *Eichhornia*

250 *crassipes* also shows good phytoremediation potential, removing up to 81% of ethion within a water
251 mesocosm experiment (Table 12.5). The removal of DDT by macrophytes shows promise. For the DDT
252 isomers *o,p'*-DDT and *p,p'*-DDT: *Spirodela oligorrhiza* can remove 66% and 50% respectively;
253 whilst *Myriophyllum aquaticum* can remove 76% and 82% respectively (Gao et al., 2000). *Elodea*
254 *canadensis* also has the ability to remove 48% to 89% of *p,p'*-DDT (Gao et al., 2000; Garrison et al.,
255 2000). *Lemna gibba*, *L. minuta* and *Potamogeton crispus* have been demonstrated to be very
256 efficient at removing phenols from water (Barber et al., 1995; Hafez et al., 1998). However, *P. crispus*
257 is less efficient at removing two PAHs, phenanthrene (removal 18-34%) and pyrene (removal 14-24%)
258 (Meng et al., 2015).

259 There is great potential for phytoremediation of a wide variety of PPCPs such as anti-
260 inflammatory, hormonal replacement and anticonvulsant products (Zhang et al., 2014). CWs (section
261 12.7.1) planted with *Phragmites australis* demonstrated very efficient removal of the hormones
262 Estrone, 17 beta-estradiol and 17 alpha-ethinylestradiol from water (Table 12.5). In CWs the water
263 column/plant sediment matrix at a depth of c. 7.5 cm provided more efficient PPCP removal than
264 deeper depths of 30 cm (Zhang et al., 2014). This highlights the importance of oxygen for the removal
265 of waterborne hormone pollutants with vertical mixing from the surrounding atmosphere increasing
266 the aeration of plant roots and (Zhang et al., 2014). Plants such as *Typha latifolia* with more
267 extensive roots and rhizomes system may be favourable for deployment due to their capacity to
268 oxygenate water (Makvana and Sharma, 2013).

269 *Scirpus validus* displays mixed ability to remove anti-inflammatory pharmaceuticals with
270 very efficient removal of naproxen, compared to very poor removal of diclofenac (Zhang et al., 2012;
271 Zhang et al., 2013a). *Typha angustifolia* removed 27-91 % of anti-inflammatory drugs in a study by
272 Zhang et al. (2011). Chen et al. (2016) found that there is large variability in planted rural CWs in
273 terms of their removal efficiency of PPCPs with 11-100% removal of anti-inflammatories, 37-99% for
274 β -blockers and 18 - 95% for diuretics. Understanding this variability and identifying macrophytes for

275 the removal of PPCPs through laboratory studies and at the field-scale is important given the need
276 for lowcost removal solutions, especially in developing countries. There has been little focus on the
277 use of novel macrophyte planting systems(e.g.FTWs) for the removal of organic chemicals, and
278 future work on these systems would build flexibility into the deployment of different aquatic
279 phytoremediation schemes for tackling the problem of PPCP pollution. Importantly, the distribution
280 and storage of organic chemicals within plants, especially for PPCPs, requires further study in
281 order to avoid the problem of transferring pollutant from one place to another(sections 12.8 and
282 12.9).

283 12.4.4 Microbial pollutants

284
285 Most studies on the removal of microbial pollutants and their indicators of the presence
286 (e.g. *E.coli*, faecal coliforms and faecal streptococci) are focused on macrophytes within CWs,
287 therefore the following examples will mainly refer to this planting type (see section
288 12.7.1). Furthermore most studies show that CW planting systems remove microbial pollutants from
289 water via a combination of chemical, biological and physical mechanisms. A study of 12 CWs found
290 that over a year vegetated CWs removed between 95-97% of faecal coliforms and 93-98% of faecal
291 streptococci (Karathanasis et al., 2003). Similarly, in an experimental CW system, Makvana &
292 Sharma(2013) demonstrated removal rates of 94%, 87% and 94% for *Salmonella*, *Shigella* and
293 *Vibrio*, respectively. However, the removal of *Salmonella* and *E. coli* from water in unplanted control
294 mesocosms versus mesocosms containing *Typha latifolia*, *Cyperus papyrus*, *Cyperus alternifolius* and
295 *Phragmites australis* showed no significant difference in the removal rates (>98 %) between the two
296 treatments; furthermore, in general, unplanted mesocosms reached their maximum removal rate
297 before the planted mesocosms (with the exception of the *C.alternifolius* mesocosm) suggesting that
298 plants provide little additional benefit for removing biological pollutants over and above the effect of
299 standing water conditions (Kipasika et al., 2016). Similarly, a review comparing *Lemna sp.* treatment
300 ponds against unplanted treatment ponds showed that the latter had greater removal rates of
301 *E.coli* facilitated by the greater exposure of the water to UV light and the subsequent

302 photodegradation and microbial die-off (Ansa et al., 2015). However, Decamp & Warren (2000) have
303 shown that gravel beds planted with *Phragmites australis* remove *E. coli* more quickly compared to
304 unplanted soil beds, possibly as a result of the impact of antagonistic root exudates from *P.*
305 *australis* on *E. coli* survival.

306 The variability of the results obtained between planted and unplanted experiments suggests
307 that for each treatment system different mechanisms of microbial pollutant removal become
308 dominant. Within unplanted facultative systems or lagoons it is likely that oxygenation and
309 phytodegradation from UV light are the dominant methods of removal (Ansa et al., 2015).
310 Conversely, biological and chemical processes may become more important within planted systems,
311 for example, *Pistia stratiotes* facilitates presence of protozoa by providing structural habitat, which
312 can increase predation on *Salmonella* (Awuah, 2006). Conversely, predation from protozoa seemed
313 to have a negligible effect in systems planted with *Spirodela polyrhiza* (greater
314 duckweed), highlighting that removal mechanisms are probably related to below-ground
315 morphological attributes, with more extensive roots/rhizomes providing superior habitat for grazers
316 (Awuah and Gyasi, 2014). Increased root zone surface area also facilitates greater microbial biofilm
317 growth which is thought to be a key removal structure for bacterial adsorption and predator
318 microbial proliferation (Decamp and Warren, 2000). Therefore, smaller grasses such *Festuca*
319 *arundinacea* may have limited potential for microbial pollutant removal compared to large emergent
320 such as *Typha latifolia* (Decamp and Warren, 2000). Future research investigating the ability of
321 different macrophytes to remove microbial pollutants from water, especially outside of CW systems,
322 is clearly merited. Direct deployment of macrophytes for pathogen removal would be highly
323 beneficial in developing countries where low-cost options for remediation could provide accessible
324 water treatment.

325 Of the few experimental studies investigating potential for macrophyte removal of microbial
326 pollutants outside of CWs, Saeed et al. (2016) demonstrated a 72 % reduction of *E. coli* in FTWs

327 planted with *Phragmites australis* and *Canna indica*. However, during times of high *E.coli* loading,
328 induced by experimental 'shock phases' where hydraulic loading was increased between 5 to 14-fold
329 to simulate low frequency and high magnitude discharge events, the removal of *E. coli* was reduced
330 significantly to levels varying between 6-45%. The effect of hydraulic retention time is also important
331 for pathogen survival and die-off (Reinoso et al. 2008) and may have implications for the use of
332 phytoremediation (with FTWs) in lakes and rivers given the difference in hydraulic retention times.

333 12.5 Macrophyte phytoremediation communities

334

335 There has been considerable work focusing on the ability of individual plant species to remove
336 single pollutants from water (e.g. Zhou & Wang 2010), with the design of CWs also focusing on
337 monocultures of macrophytes (Kadlec, 2009). Conversely, there has been a lack of studies that
338 explicitly explore the ability of mixed plant assemblages to simultaneously take-up and degrade
339 multiple pollutants (Koelbener et al., 2008). A plant community-based approach provides the
340 opportunity to enhance the removal of both single pollutants, but also target multiple
341 contaminants. Studies that have looked specifically at phytoremediation using plant communities
342 have shown encouraging results (Fraser et al., 2004; Zhang et al., 2007; Liang et al., 2011; Türker et
343 al., 2016). For example, an experiment testing the removal of N and P from four different emergent
344 macrophytes in parallel (*Carex lacustris*, *Scirpus validus*, *Phalaris arundinacea* and *Typha latifolia*)
345 found that microcosms planted with all four macrophytes in equal proportion, either matched or
346 outperformed microcosms planted with a single species (Picardet al. 2005). Earlier studies also
347 suggest that plant polycultures have a greater removal potential for heavy metals and can reduce
348 biochemical oxygen demand (BOD) (Karpiscak et al., 1996; Scholes et al., 1999). However, Türker et al.
349 (2016) reported that boron removal from mine effluent was more effective in native emergent
350 monocultures compared to polycultures, although the opposite was true for NO_2^- removal. These
351 results suggest that there are probably optimal plant combinations for particular pollutants and

352 further experiments designed to identify these combinations would help to optimise the efficiency
353 of phytoremediation.

354 To assemble appropriate plant combinations there are several important factors to consider
355 including the functional diversity of the community. It has been reported that simply increasing
356 species diversity in a plant assemblage can increase nutrient removal, although polycultures
357 containing more than three species showed no further benefit (Ge et al. 2015; Geng et al. 2017). A
358 common theme among these studies is the importance of species identity in explaining variation in
359 nutrient removal, where specific combinations can more effectively remove pollutants. Therefore,
360 assembling appropriate plant communities based around the complementary phytoremediation
361 potential of individual species, and the interaction of those plants with others in the assemblage is
362 potentially more important than simply increasing species richness per se. However, the effect of
363 competition between plants is important to recognise as this may impact the community
364 composition, and therefore the ability to remove the targeted pollutants from water (Zhang et al.
365 2007). In a mesocosm experiment, containing the submerged macrophytes *Stuckenia pectinata*
366 (Sago pondweed), *Potamogeton natans* (broad-leaved pondweed), *Potamogeton crispus* (curled
367 pondweed) and *Zannichellia palustris* (horned pondweed), it was found that *S. pectinata* reduced the
368 biomass of the other species (Engelhardt & Ritchie, 2001). Reducing the biomass of certain species
369 will not necessarily compromise overall removal efficiency as uptake and sequestration potential will
370 vary with species. However, this highlights the need to understand interspecific interactions in order
371 to enhance removal efficiency, especially when considering targeting water bodies in a non-
372 equilibrium state where conditions favour the dominance of one particular species (Engelhardt &
373 Ritchie, 2002).

374 A field study employing plant communities revealed some of the benefits of combining multiple
375 macrophytes (Wang et al., 2009; Zhao et al., 2011). Nine macrophytes species (five floating, one
376 submerged and three emergent) deployed on FTWs and planted on river banks outside Jiaying City,
377 China, demonstrated removal rates of TN and TP at 16%-37% and 26%-43% respectively (Zhao et al.,

2011). Although the removal rates were relatively low, it was also highlighted that the plant community-based approach allows for species within the community to compensate for deficits in uptake of other species (Zhao et al., 2011). For example, the average P content of floating macrophytes was *ca.* 5.9g/m², whereas, emergent species including *Canna indica* and *Pontederiacordata* with higher biomass accumulation, stored P at a level of *ca.* 7.3g/m². Similarly, a phytoextraction study with emergent species (*Carex flava*, *Centaurea angustifolia* and *Salix caprea*) allowed the impact of facilitation across increasing concentration gradients to be seen (Koelbener et al., 2008). Here, the willow *S. caprea* attenuated the toxic effect of Zn on the relative growth rate of *C. flava* by lowering the availability of Zn, thus mitigating the negative effect of Zn on the sedge (Koelbener et al., 2008). This highlights that competitive effects may not always be negative and may produce positive effects through 'over yielding'. The consequences of competitive interactions between candidate macrophytes evidently deserve particular attention within the field of plant community-based phytoremediation.

As well as the potential enhanced removal of pollutants from plant communities with macrophytes of different life forms (Koelbener et al., 2008) there may also be the potential for generating ecosystem services from polycultures. A 2-year study by Wang et al. (2009) explored the potential restoration of Lake Taihu and Lake Machou by using a mosaic of macrophytes in successional stages highlighting the potential for spatial and temporal diversity in macrophyte deployment, and the provision of ecosystem services. Floating and emergent macrophytes were first introduced to reduce light availability for algal growth, facilitating the introduction of submerged species leading to removal rates of TN and TP of 60% and 72% (Wang et al., 2009). The provision of ecosystem services due to the different plant life forms was highlighted as an advantage by Wang et al. (2009) as increased patches of vegetation provided refuge for zooplankton that subsequently grazed phytoplankton. The added value of diverse plant communities is a factor that requires quantification to espouse the benefits of aquatic phytoremediation over and above water treatment.

403 Plant community-based approaches provide the opportunity to build temporally more
404 consistent treatment into phytoremediation by exploiting the differing phenology of plant species;
405 polyculture systems can thus offer the most consistent water treatment option with least
406 susceptibility to seasonal variation (Karathanasis et al., 2003). However, the temporal dynamics of
407 plant communities within the context of phytoremediation are under-researched, and there is a
408 need to explore the assembly of plants, e.g. in terms of differing phenologies, to extend the growing
409 season, especially in temperate regions where water treatment potential declines after senescence.

410

411

412 12.6 Issues in utilising invasive macrophytes

413

414 The most effective phytoremediators have fast growth rates and high biomass
415 accumulation; however, outside of their native range macrophyte species with these traits are often
416 considered to be invasive, and given their potential for rapid colonisation they can quickly
417 outcompete native macrophytes (Chambers et al., 2008). Species that are invasive in the UK, such as
418 *Azolla filiculoides* and *Hydrocotyle ranunculoides*, can clog waterways and have serious ecological
419 impacts on native flora and fauna (Schultz and Dibble, 2012). In the UK, the combined cost of
420 controlling invasive plants, together with their economic impact, is estimated to be £1.7 billion per
421 annum (The Great Britain Non-native Species Secretariat, 2015). Therefore, there is a
422 significant juxtaposition between using species of invasive plants in phytoremediation, and
423 management strategies to control invasive species (Rodríguez et al., 2012). Given that in many cases
424 the complete eradication of invasive aquatic macrophytes such as *Eichhornia crassipes* is unlikely, it
425 may be more appropriate to exploit these macrophytes as part of an integrated management
426 strategy that controls the spread of these species whilst at the same time effectively removing
427 nutrients and metals, capturing suspended sediment, and harvesting the biomass for economic gain
428 (Patel, 2012; Yan et al., 2017). A similar parallel can be drawn with non-native and invasive zebra

429 mussels (*Dreissena polymorpha*) which are often considered detrimental(Matsuzaki et al., 2009), but
430 have also widely been reported to stabilise the clear-water state of shallow lakes through filtering
431 phytoplankton and removing harmful cyanobacteria (Gulati et al., 2008).

432 Water bodies where invasive species are already present may be targeted for active
433 harvesting allowing periodical regrowth for continued phytoremediation(Xu et al., 2014). However,
434 there are important factors to consider including the containment of macrophytes to avoid
435 transferto other water bodies (e.g. via contaminated harvesting equipment or through downstream
436 spread of fragments), including the most appropriate harvesting technique, and the sustainability of
437 exploiting such an ecological engineering systems(Rodríguez et al., 2012; Yan et al., 2017). The site-
438 specific context will likely determine the appropriateness of active harvest of invasive aquatic plants
439 (Yan et al., 2017). In terms of introducing macrophytes into a freshwater system for
440 phytoremediation, it is inappropriate, and indeed possibly illegal, to deploy invasive species given
441 the potential for ecosystem damage and long terms effects. In these circumstances non-invasive or
442 native plants should therefore be employed, unless containment of invasive plants can be ensured,
443 such as in engineered CW systems.

444 12.7 Macrophyte planting systems

445

446 Macrophyte planting systems are effectivelyplanting strategies that are employed to facilitate
447 targeted phytoremediation of waters in different contexts in terms of point source and diffuse
448 source treatment and restoration. The following section details the key aspects of the three main
449 aquatic phytoremediation planting systems that have been developed; CWs, wild macrophyte
450 harvesting and planting, and FTWs.

451 12.7.1 Constructed wetlands

452

453 Phytoremediation has primarily been optimised for point source wastewater treatment in
454 the form of CWs. CWs have been used for the treatment of a variety of effluents including urban

455 storm water, sewage, mine tailing drainage, storm water treatment, landfill leachate treatment
456 systems and for wastewater polishing (Kivaisi, 2001; Nivala et al., 2007; Tanner, 1996; Vymazal, 2009;
457 Vymazal, 2011). CWs also show potential for treating wastewater containing emerging contaminants
458 of concern including pharmaceuticals and other endocrine disrupters (Vymazal, 2009).

459 CWs can be categorised as free water surface flow wetlands (FWSF) or sub-surface flow (SSF)
460 wetlands (Dhir, 2013) (Figure 12.3). FWSF wetlands contain emergent, floating and submerged
461 macrophytes growing in shallow ponds or lagoon waters over sandy or organic soils, which allows the
462 influent contaminated water to slowly flow through the emergent macrophyte stems for maximum
463 pollutant uptake and UV degradation (Kadlec, 2009). SSF wetlands are the most common type of CW
464 and comprise emergent macrophytes growing over a substrate of stone or gravel matrix enabling
465 water to come in direct contact with plant roots, rhizomes and biofilms, which promote aerobic
466 conditions (Vymazal, 2011). Several processes including physical filtering of the water, biological
467 processing of water by plants and microbial biofilms, and chemical changes due to redox state can
468 assist in pollutant removal in SSF systems (Faulwetter et al., 2009). The average SSF CW system is
469 100 times smaller than the FWSF CW system (Kadlec, 2009), therefore, FWSF are more common in
470 North America and Australia where a larger surface is available, whilst SSF wetlands are more
471 common in Europe where land availability is more limited (Vymazal, 2011). SSF wetlands are
472 frequently used to ameliorate the concentration of biologically derived organic material as indicated
473 by the lowering of biochemical oxygen demand (BOD) and chemical oxygen demand (COD) from
474 waste waters (Vymazal & Kröpfelová, 2009).

475 CWs are the most advanced form of macrophyte deployment within the umbrella of aquatic
476 phytoremediation (Kennen and Kirkwood, 2015). However, these systems can require high
477 investment costs and they are restricted primarily to pollutant point sources where there is
478 wastewater treatment such as tertiary sewage treatment and wastewater polishing before water
479 enters a natural waterway (Patiño Gómez and Lara-Borrero, 2012). This restricts the application of

480 CWs for the treatment of water containing pollutants from diffuse sources. Although CWs have the
481 potential to be utilised for treatment of a wide range of contaminants, their most widespread
482 application has been for sewage wastewater-related contaminants, including BOD, COD, N and P,
483 and often they are set up with crop monoculture to maximise plant uptake (Kadlec & Wallace, 2009;
484 Sundaravadivel & Vigneswaran, 2001; Vymazal, 2009).

485 CWs vary in level of design and engineering required for their development; FWSF wetlands
486 are generally low tech gravity-fed systems, whereas, SSF require more construction and
487 management to import the stone/gravel matrixes, and also may include bundsto separate different
488 treatments then requiring the use of electric pumps (Kadlec and Wallace, 2009). In both types of
489 CWs there are high investmentsin construction and operational costs. CW can also become clogged
490 with sediment, which impacts the functioning of the system and imposes additional costs for
491 excavation and removal of contaminated sediments, and the subsequent reinstatement of
492 macrophytes (Machado et al., 2016). According to design guidance for the treatment of urban waste
493 water and sewage, SSF CWs may require an area of around 5m² to 10 m² of CW per person
494 equivalent for adequate water purification(Tilley *et al.*, 2014). Therefore, given the potentially large
495 area required, CW-based phytoremediation may be unable to compete for limited land availability
496 with other more profitable land uses. Furthermore, in countries where vector-borne diseases, such
497 as malaria or dengue, are a public health issue the creation of open shallow wetland environments
498 may be undesirable as it has the potential to provide ideal conditions for the propagation of
499 mosquitoes and other disease vectors(Mwendera et al., 2017).

500 From both industry-based observations and from the available literature, the primary purpose
501 of CWs is water treatment and wastewater polishing. This however, ignores their potential to offer
502 ecosystem services such as sequestering and harvesting nutrients for reuse, provisioning for
503 biodiversity, pollination and carbon sequestration, and thus underplays the overall value of CWs.
504 There is great potential to develop different post-remediation 'streams' which have been relatively

505 unexplored, and which emphasise support for different ecosystem services (see section 12.10.2).
506 Aquatic phytoremediation is a promising technology for the treatment and remediation of polluted
507 water with the operational point-source based CW systems in place, but given the limitations of
508 these systems, including the lack of application for diffuse pollutants, investment costs and lack of
509 ecosystem focus there is an opportunity to further develop context-specific, sustainable
510 phytoremediation that provides ecosystem services within wider environmental systems.

511 12.7.2 Wild macrophyte harvesting

512
513 Most aquatic phytoremediation planting systems involve the deliberate deployment (FTW) or
514 engineering of planted systems (CWs). Harvesting of existing wild macrophytes from water bodies
515 such as shallow lakes can also be a phytoremediation strategy, and relies upon the opportunistic and
516 timely removal of macrophyte biomass in order to manage waterborne pollutants such as N and
517 P (Huser et al., 2016). A study of an urban shallow lake, showed that harvesting an annual amount of
518 3,600 kg dry weight of *Elodea canadensis* led to 16.4 kg P being removed from the system, equating
519 to around 53% of the TP load removed (Bartodziej et al., 2017). Although the estimated cost of
520 removal was \$670 per kg of TP, which was more expensive than chemical flocculating treatment, this
521 was still considerably less expensive than many catchment best management practices (Bartodziej et
522 al., 2017). Macrophyte harvesting is often carried out in lakes and waterways ostensibly to relieve
523 navigation, drainage, aesthetic or recreational problems, rather than for phytoremediation
524 purposes, but it is notable that nutrient export may be a collateral benefit of such harvesting. Other
525 case studies have shown that macrophyte harvesting for nutrient removal does not reduce nutrient
526 loading quite as favourably (Carpenter and Adams, 1977; Morency and Belnick, 1987), with Peterson
527 et al. (1974) estimating that plant harvesting only removed 1.4% of TP loading.

528 The variation between these case studies is possibly a result of the levels of nutrient loading,
529 with waters that receive extremely high inputs of nutrients leading to a poor offset by removal from
530 plant harvesting (Bartodziej et al., 2017). Another source of variability for nutrient removal is the

531 coverage of macrophytes across the particular water body; the reported optimal coverage of
532 macrophytes ranges from 5% to 40% (Portielje and Van der Molen, 1999; Dai *et al.*, 2012; Xu *et al.*,
533 2014). For environmental managers considering macrophyte harvesting as a mechanism for in-water
534 nutrient management, it is crucial that a scoping study is carried out to determine the base balance
535 of nutrient input/output and plant removal capacity, and to identify the need for upstream best
536 practices as part of an integrated management strategy.

537 The harvesting method itself is also an important element of harvesting wild macrophytes, e.g.
538 removal by hand, or mechanically via specialised boats equipped with cutting or raking apparatus
539 (Quilliam *et al.*, 2015). Hand removal is labour and time intensive, although it allows targeted
540 macrophyte removal and minimises disturbance (Quilliam *et al.*, 2015). Conversely, mechanical
541 removal allows more rapid and extensive removal but is non-selective and can lead to high levels of
542 turbidity due to the re-suspension of sediments. This can impact invertebrates and fish by removing
543 structural habitat and may ultimately drive the system from a desirable clear water macrophyte-
544 dominated state to a potentially unfavourable phytoplankton-dominated state (Dawson *et al.*, 1991;
545 Sayer *et al.*, 2010; Habib and AR, 2016).

546 In some circumstances it may be necessary to establish macrophytes in waterbodies by direct
547 planting through seeding or transplanting propagules (e.g. tubers/root crowns) if there are
548 no existing macrophytes, or if a particular species is required to target certain pollutants (Smart *et al.*,
549 1998; Hilt *et al.*, 2006). In addition to plant establishment there is also scope to
550 enhance macrophyte growth and biomass by engineering interventions such as the assembly of
551 polytunnels over vegetation, or enclosures to reduce grazing losses.

552 12.7.3 Floating treatment wetlands

553
554 Within aquatic phytoremediation one such novel ecological engineering solution that has been
555 developed is the FTW. The premise of this system is that highly productive emergent macrophytes
556 such as *Typha latifolia* are planted within a growth medium, which is supported by a buoyant frame

557 allowing the roots of the emergent macrophytes to be submerged in the water, thus enabling
558 rhizofiltration, phytoextraction and phytodegradation to take place hydroponically(Nichols et al.,
559 2016; Kiiskila et al., 2017) (Figure 12.4). Root uptake associated with FTWs is primarily applicable to
560 water-soluble contaminants within the water column only, although sediment-bound pollutants
561 canbe physically filteredfrom the water column by plant roots (Tanner and Headley, 2011b). FTWs
562 have recently gained increased attention and may also be referred to in the literature as artificial
563 floating islands, integrated ecological floating beds, floating plant bed system and hydroponic root
564 mats (Yeh et al., 2015).

565 FTWs can accommodate fluctuations in water levels, andthe stability of materials used to
566 construct the buoyant frame may include items such as polyvinyl chloride (PVC) pipes, foam sheets,
567 bottles and bamboo (Ladislav et al. 2013; Wang et al. 2015;Pavlineri et al. , 2017). However, it would
568 be useful within the literature if qualitative information and design challenges were also reported to
569 provide an idea of performance and usability of FTWs in practice, and although there are no
570 reported incidences of FTWs capsizing or other failures during pilot tests, this may simply reflect
571 publication bias.

572 Netting material or foam is generally used to support the growth medium in which the
573 macrophytes are grown (Yeh et al., 2015). Material previously used as substrate includes peat, soil,
574 cotton and coir fibre (Pavlineri et al., 2017). Furthermore, FTWs comprising foam with gaps to
575 support pots have also been designed (Lynch et al., 2015). Growth media physically supports the
576 planted macrophytes and provide nutrition, but the substrate can also enhance pollutant removal
577 through the stimulation of microbial activity (Tanner & Headley, 2011a). Macrophytes may be
578 established by transplanting of seedlings, cuttings or whole plants (Yang et al., 2008; Ning et al.,
579 2014). An advantage of using FTWs rather thandirect plantingof macrophytes is the ease in which
580 the biomass can be harvested from the frame, instead of having to remove plants from the
581 sediment. The quick and simple method of harvesting afforded by growing plants in FTW facilitates

582 recovering pollutants from plant biomass (Bartodziej et al., 2017). There is potential for quick re-
583 planting of the FTW for continued remediation and biomass removal (Wang et al., 2015; Ge et al.,
584 2016).

585 FTWs have been studied principally for their capacity to remove nutrients, but there have also
586 been attempts to assess heavy metal, pathogen and phytoplankton removal (Borne, 2014; Yeh et al.,
587 2015; Jones et al., 2017; Kiiskila et al., 2017). FTWs have been deployed at a variety of different
588 scales including microcosms, mesocosms, and as pilot trials within lagoons (Headley and Tanner,
589 2008; Ladislav et al., 2013; Chang et al., 2014; McAndrew et al., 2016; Nichols et al., 2016; Kiiskila et
590 al., 2017). Here the experimental polluted water used has included storm water, lake water, river
591 water, sewage effluents, domestic wastewaters, refinery wastewater, acid mine drainage, and
592 livestock effluents (Zhu et al., 2011; Li et al., 2012; Borne, 2014; Wang and Sample, 2014a; Abed,
593 Almukhtar and Scholz, 2017; Kiiskila et al., 2017). Mesocosm-scale studies are the most prominent
594 form of exploration into the effectiveness of FTW thus far (Chen et al., 2016), although there have
595 been a few examples of deployment at field-scale, such as Zhao et al. (2012) who demonstrated
596 that TN and TP concentrations could be reduced in a polluted Chinese river. Mesocosm studies
597 with synthetically produced experimental water allows full control of all input parameters.
598 However, they may not be representative of the real remediation performance given that polluted
599 waters contain a multitude of chemicals and microbes which may influence remediation (Javadi et
600 al., 2005). Therefore, further studies would benefit from testing the remediation of water sourced
601 from the environment.

602 Only a small handful of field-scale experiments have been carried out that assess the usefulness
603 of FTWs in successfully remediating pollutant-impacted waters (Zhu et al., 2012; McAndrew et al.,
604 2016; Nichols et al., 2016; Olgún et al., 2017). Of the available studies that assess FTW performance
605 within water bodies, including streams, urban and rural ponds, results focus on plant tissue element
606 accumulation rather than the arguably more pertinent issue of water quality improvement (Zhu et

607 al., 2012; Olguín et al., 2017; McAndrew et al., 2016; Nichols et al., 2016). Although plant tissue
608 sequestration is extremely important for assessing the bioaccumulation potential of macrophyte
609 species it does not explicitly demonstrate water quality improvement; this can only be proven
610 through monitoring water chemistry. Scaling up mesocosm scale experiments to assess actual field-
611 scale water quality improvement is challenging given the ideal of a control site with comparable
612 water chemistry and abiotic and biotic conditions, or high-temporal resolution baseline water quality
613 data for the experimental water body, both of which may be unavailable. Where there is a clear
614 opportunity for upstream and downstream water quality sampling near the experimental FTWs,
615 such as a stream, water quality changes are more likely to be attributed to the FTW intervention
616 between these points (Olguín et al., 2017). Similarly, more field studies longer than 2 years, ideally
617 up to 5 to 10 years, would lead to a better understanding of the longer-term performance of FTWs
618 and, crucially, reveal the actual remediation time (Yang et al., 2006). Furthermore, the influence of
619 inter-annual hydrological variability on FTW performance in terms of precipitation and evaporation
620 could also be evaluated. Despite the paucity of scientific studies at the field scale, commercial
621 companies now commonly offer FTWs as a water treatment solution, and as part of the aesthetic
622 enhancement of urban rivers. The phytoremediation research community must aim to keep pace
623 with the private sector to corroborate industry-advocated benefits of FTWs and avoid any potential
624 reputational damage to aquatic phytoremediation where expectations of these systems from
625 stakeholders are not met (Keenen and Kirkwood, 2015).

626 The remediation performance of FTWs is highly variable with reported minimum and maximum
627 removal efficiencies for TN values being 0.71 mg/l (4 %) and 51 mg/l (91 %) and 0.06 mg/l (1 %) and
628 18.85 mg/l (90 %) for TP (Figure 12.5). This high variability may be due to differences in FTW design,
629 macrophyte species employed, and the chemical composition of the experimental water. A further
630 example of variation in removal efficiency comes from Lynch et al. (2015) who compared two
631 commercial FTWs (Beemat and BioHaven®) planted with the rush *Juncus effusus* that had been
632 designed to treat storm water. It was found that Beemat FTW outperformed BioHaven® in both TN

633 and TP removal (Lynch et al. 2015). The difference in removal may have been due to the difference
634 in substrate (coir matting vs. sphagnum peat) or the physical design of FTW(Lynch et al. 2015).The
635 growth medium is indeed an important source of variability within FTW design. Rice straw used as
636 growth medium was found to enhance removal of TN, NH_4^+ and NO_3^- compared to plastic filling(Cao
637 and Zhang, 2014). Similarly, the FTW with straw filling had a greater total densityof nitrifying and
638 denitrifying bacteria which suggests thatthisorganic material was providing both a habitat and a
639 source of C for the growth of microorganisms, which were able contribute to pollutant metabolism
640 (Cao and Zhang, 2014).Commercial FTWs are still an expensive management option, and there is
641 currently a demand for more low-cost growth media that both provides a suitable substrate for
642 macrophytes and enhances pollutant removal; such examples includebiochar, activated carbons,
643 coffee waste and green compost (Tran et al., 2015). To date there has been no research
644 incorporating these materials into FTWs to assess the potential for enhanced remediation and the
645 potential value post-remediation.

646 HybridFTW planting systems are being developed in an attempted to enhance pollutant removal
647 and ecosystem restoration (Guo et al., 2014; Li et al., 2010; Lu et al. 2015). Such systems integrate a
648 new layer beneath the floating platform containing submerged macrophytes such as *Potamogeton*
649 *crispus*, and/or bivalves such as freshwater clams (*Corbicula fluminea*) (Guo et al., 2014; Li et al.,
650 2010) (Figure 12.6). Photovoltaic solar panels have also been attached to the frames of FTW to
651 power a submerged aerator to enhance oxygenation in the vicinity of the plant roots and associated
652 microorganisms, thus increasing the nutrient degradation process (Lu et al., 2015) (Figure
653 12.6).While these hybrid systems appear to enhance pollutant removal from the water column
654 compared to their macrophyte-only counterparts (Guo et al., 2014; Li et al., 2010), the added
655 complexity may impact on the utility of FTW as a phytoremediation system. With increasing
656 complexity of FTW design there is an increase in pollutant removal efficiency, cost and maintenance,
657 but a decrease in user uptake given the added management of submerged plants or solar PV
658 systems. A focus on maximising removal efficiency over the simplicity of the system may create

659 barriers for uptake by stakeholders such as farmers, land managers and government organisations
660 looking for low-cost low maintenance treatment options, especially within developing countries. A
661 useful exercise might be to compare the economics, maintenance requirements and user experience
662 of hybrid versus conventional FTWs to determine when increasing FTW complexity is appropriate.

663 The coverage of FTW over the target water body is also important, as indicated by a meta-
664 analysis showing that vegetation cover is significantly correlated with the removal of NH_4^- (Pavlineri
665 et al., 2017). Although increasing FTW coverage reduces atmospheric diffusion, oxygen is supplied to
666 water by emergent plants via root oxygenation (Xiao et al., 2016; Yeh et al., 2015). Furthermore, in
667 eutrophic waters this coverage may inhibit algal primary productivity, which may be beneficial for
668 mitigating the potential for occurrences of large algal blooms (Jones et al., 2017). The optimal
669 coverage of FTWs has been reported as 10-25% (Marimon et al., 2013), although generally there is
670 wide variation in the literature with values of between 100 %, 50 % and 5-8 % being reported as
671 acceptable for water treatment (Pavlineri et al., 2017). McAndrew & Ahn (2017) also note that
672 hydraulic retention time and plant productivity are important for determining removal efficiency.
673 Surface cover therefore needs to be considered in tandem with hydrology and macrophyte
674 selection. As the focus within the literature is on coverage, there has been no clear attempt to look
675 at the different surface arrangements of FTW on the water surface. For example, targeting of an
676 area, such as water inlet or outlet to a lake may be more beneficial than increased FTW coverage
677 over the target water body. Clearly, the coverage and area of FTW treatment is context-specific but
678 there is likely to be significant potential in investigating spatially targeted phytoremediation.

679 Finally, the poor design and management of FTWs is a topic that is rarely discussed within the
680 literature. FTWs have the potential to be pollutant sources should the biomass not be continually
681 harvested and removed, or if water birds attracted to the FTWs defecate into the water inputting
682 nutrients and microbial contaminants (guantrophication). Nutrient-rich growth media such as peat
683 may also leach nutrients into the target water body compared to more inert coir fibre (Lynch et al.,

684 2015). The placement of FTWs in watercourses must also be given full consideration as water birds
685 and recreational users may also use the target waterbody. FTWs potentially slow the velocity of
686 water in small water bodies such as ditches, which may conflict with farming interests where good
687 drainage is required. As with any good catchment management practice, appropriate consultation
688 with stakeholders is important for success.

689

690 12.8 Translocation and element storage in macrophytes

691

692 Understanding how and where nutrients and other pollutants are distributed within
693 macrophyte tissues is important to inform plant harvesting for removal of pollutants. The recovery of
694 nutrients is crucial for the value of post-harvest plant biomass, whilst ensuring correct plant parts
695 are harvested for effective removal of heavy metal and organic pollutants from the planting system.
696 Allometry of pollutants within plants varies according to species, but is also influenced by the
697 environmental conditions in terms of nutrient availability (Barrat-Segretain, 2001; Demars and
698 Edwards, 2007).

699 *Typha domingensis*, *Eichhornia crassipes*, *Pistia stratiotes* and *Myriophyllum*
700 *aquaticum* preferentially store N and P in the shoot compared to the roots or rhizome (Table 12.6),
701 although nutrients can be translocated through the plants leading to temporal dynamics in element
702 distribution driven by plant phenology and diurnal metabolism (Masclaux-Daubresse *et al.*, 2010;
703 Hawkesford *et al.*, 2011; Eid *et al.*, 2012). More than 50% of N can be stored in below-ground plant
704 parts by the end of a growing season (Vymazal, 2007). *Phragmites australis* grown in either natural
705 waters or a waste water infiltration pond demonstrated a clear seasonal pattern in the translocation
706 of nutrients from above-ground to below-ground parts as the end of the growing season
707 approached (Meuleman *et al.*, 2002). Early in the growing season N and P concentrations are higher

708 due to sink demand during active growth before concentrations decrease gradually through the
709 season as plants begin to senesce.

710 Coinciding with the decrease in nutrient concentrations in above-ground biomass, below-
711 ground concentrations of N and P increase, representing the preparation for plant senescence with
712 nutrient storage in the roots and rhizomes for the following season's growth(Garver et al., 1988).
713 Meuleman et al. (2002) suggested that harvesting during the winter meant that only 9% of N and 6%
714 of P associated with nutrient loading was removed, whereas, harvesting above-ground parts during
715 peak nutrient storage in summer enhanced removal to 40-50% of N and P. Seasonality is
716 important,although seasonal effects will differ between temperate, subtropical and tropical zones
717 with macrophytes in the latter two zones showing less element translocation and therefore enabling
718 multiple annual harvests(Vymazal, 2007).Macrophytes may perform poorly if nutrient translocation
719 to the rhizome is inhibited by harvesting during the active growing period (Tanaka et al., 2017),
720 although the issue of nutrient allocation is less problematic for floating macrophytes and emergent
721 macrophytes deployed in FTWs as the full plant can then be harvested(Wang et al. 2014).

722 Studies on element allocation tend to report absolute concentrations to determine if a
723 species is a better above-ground or below-ground accumulator. The potential for pollutant uptake
724 and removal by harvesting the areal parts is a function of both concentration and the biomass
725 produced (Polomski et al., 2009). For example, although shoot concentration of N in *Pistia*
726 *stratiotes*(13.93mg/g) was greater than in *Eichhornia crassipes*(10.16mg/g) in a study of nutrient
727 recovery, the total areal shoot storage of N for *Eichhornia crassipes* was over four times higher due
728 to its greater biomass(Polomski et al., 2009). This demonstrates that it is more effective to harvest
729 plants with greater above-ground biomass and moderate tissue concentrations of the pollutant of
730 interest, rather target plants with lower biomass but higher tissue concentrations (Duman et al.,
731 2007; Vymazal, 2016).

732 In eutrophic waters light is commonly the limiting factor for growth and plants therefore tend to
733 allocate nutrients to above-ground growth to maintain efficient light capture, while excessive
734 nutrient availability negates the requirement for belowground storage (Polomski *et al.*, 2009; Lynch
735 *et al.*, 2015); this also maintain intra-specific competitive advantages in these environments and can
736 be exploited as part of a phytoremediation management strategy . Where non-hyperaccumulator
737 plants are grown in a substrate where high concentrations of heavy metals and organic pollutants
738 are present, physiological mechanisms within these plants often limit the transport of these
739 compounds to above-ground tissue to mitigate damage to important cells, such as those responsible
740 for photosynthesis (Zhu et al., 1999;Verkleij et al. 2009).

741 The preference for below-ground storage by emergent macrophytes has been demonstrated in
742 multiple studies, as listed in Table 12.6. However, there are some occasions where metals are found
743 at greater concentration in aerial parts, such as Pb in *Cyperus esculentus*, Zn in *Glyceria maxima*, Mn
744 in *Phragmites australis* and Cu in *Phragmites australis* (Table 12.6), which suggests that specifically
745 classing species as above-ground or below-ground accumulators of specific pollutants may be
746 inappropriate. Furthermore, not all studies capture the full seasonal dynamics of nutrient or
747 pollutant translocation and allometry under different concentration regimes, and therefore, to
748 enable sound recommendations on harvesting during phytoremediation projects, further studies to
749 characterise chemical allocation over time of key species should be carried out to ensure pollutant
750 removal is appropriately targeted.

751 12.9 The role of microbial activity in aquatic phytoremediation

752
753 There is debate within the phytoremediation literature as to the relative importance of
754 macrophytes in removing pollutants compared to the independent microbial degradation. This
755 perspective primarily comes from observations showing that unplanted CWs can match or
756 outperform planted CWs in terms of pollutant removal (Cardinal et al., 2014). In addition to
757 microbial activity, processes such as sedimentation in P stabilisation and removal, and the

758 photodegradation of PPCPs have also been noted as important (Cardinal et al., 2014; Tanner &
759 Headley, 2011; Zhang et al., 2014). Microbial activity is also an important factor for
760 enabling phytodegradation of pollutants, however, the independent role of microbial communities is
761 now receiving much more attention (Houda et al., 2014). Improved understanding of how microbial
762 activity contributes to pollutant degradation is essential because it not only influences removal rates
763 but may have implications for the value of harvesting plant biomass and post-remediation resource
764 recovery if the actual plant uptake and sequestration (phytoextraction) of target pollutants is low.

765 There is an abundance of microorganisms associated with macrophyte roots that influence
766 the removal and degradation of pollutants (Stottmeister et al., 2003; Faulwetter et al., 2009). These
767 include bacteria that assist in nitrification and denitrification for the transformation and removal of
768 excess N, and biological mineralization of organic P (Valipour and Ahn, 2016). These processes are
769 integral to the efficient functioning of CWs but the role of macrophytes in facilitating and enhancing
770 the metabolic processes of these microorganisms is still not well understood, although it is likely
771 that the rhizosphere provides an energy source for microorganisms (Thijs et al., 2016). Redox state,
772 dissolved oxygen content and temperature are common limiting factors for different
773 microorganisms (Truu et al., 2009), and the potential for macrophytes to oxygenate the substrate
774 surrounding their below-ground organs can also facilitate the growth of microbes in the rhizosphere
775 (Pavlineri et al., 2017).

776 CWs are highly engineered, with multiple design elements that may influence the
777 abundance and diversity of microorganisms. Consequently carefully designed experiments are
778 required to explore the potential role of the plant microbiome in phytoremediation. Applying this
779 knowledge is particularly important for developing novel environmental engineering solutions such
780 as FTWs. The formation of microbial biofilms on the underside of FTWs and plant roots has been
781 suggested as a key removal pathway for nutrients and heavy metals (Tanner et al., 2011). Wang &
782 Sample (2014) found that unplanted FTWs had similar removal efficiencies compared to those

783 planted with monocultures of *Pontederia cordata* and *Schoenoplectus tabernaemontani* (Figure 12.7).
784 In this study, and elsewhere, temperature was a key factor in the performance of FTW which has
785 been related to changes in microbial activity (Van de Moortel, 2011; Wang & Sample, 2014b). In
786 contrast, Zhang et al. (2014) were unable to link microbial community traits associated with FTWs
787 biofilm such as ribotype number and diversity index to the removal efficiency of pollutants.

788 Given the conflicting evidence on the relative importance of plants and biofilms in
789 phytoremediation, a 'meta-organism' approach to phytoremediation is now required to appreciate
790 the multitude of factors and process at work (Thijs *et al.*, 2016; Feng *et al.*, 2017). Further studies are
791 required in these areas that employ suitable control treatments, along with adequate spatial and
792 temporal characterisation of microbial communities for different macrophytes in monoculture and
793 polyculture, and growth media. Furthermore, within these studies the mass balance of pollutant
794 allocation should be investigated to fully assess where and how pollutants are being stored and
795 translocated. Radio-labelled isotopes have been successfully employed to quantify cycling of
796 nutrients within CWs (Truu *et al.*, 2009). However, such techniques have not been employed during
797 FTW studies, where the application of radio-labelled isotopes would provide an opportunity to
798 understand the biochemical cycling with these novel systems. Finally, after adequate
799 characterisation of microbial communities and their relation to the plant and associated abiotic
800 environment, there may be new opportunities to enhance the microbial community to promote
801 pollutant removal (Glick, 2003; Thijs *et al.*, 2016).

802 **12.10 Added value of aquatic phytoremediation**

803 **12.10.1 Ecosystem services**

804

805 The process of phytoremediation has primarily been concerned with maximising the
806 efficiency of water treatment, whilst the benefits of phytoremediation over and above remediation
807 have essentially been overlooked. Clearly, water treatment is the primary ecosystem service in the
808 provision of safe and clean water; however, the planting of vegetation within the environment

809 creates new habitats for organisms (Zhu et al., 2011). For example, the presence of artificial floating
810 islands improved chick productivity of Black-throated Divers (*Gavia arctica*) by 44 % in waterbodies
811 with these structures (Hancock, 2000), indicating a potential combined role for FTWs in water
812 treatment and improved habitat connectivity. Similarly, a 15-year project investigating the
813 environmental benefits of creating treatment wetlands to ameliorate mine tailing effluents found
814 that there was a high abundance and diversity of protozoa, higher plants, terrestrial animals, and
815 birds (Yang et al., 2006).

816 In addition to habitat provisioning there is also the potential for facilitating pollination and
817 carbon sequestration (Nesshöver et al., 2017). The capacity for the latter may depend on the post-
818 remediation stage and the reuse of the biomass. Cultural services can also be provided by an
819 improvement in the aesthetic appeal of an area with increased vegetation (Masi et al., 2017). This is
820 most likely in urban waterways where FTW might provide attractive green infrastructure (Olguín et
821 al., 2017). There is a need to quantify and assess ecosystem services associated with
822 phytoremediation projects in order to better appreciate the multiple benefits generated from this
823 form of water treatment.

824 12.10.2 Resource recovery

825

826 The potential to generate large volumes of biomass through phytoremediation means that there
827 are opportunities for resource recovery within the process (Gomes, 2012). Post-remediation
828 biomass re-use streams (PBRs) are the disposal process and utilisation of the harvested plant tissues
829 of macrophytes used for phytoremediation (Gomes, 2012). As macrophytes are able to remove and
830 assimilate metals there is certainly potential for the recovery of metals such as gold, Cu and Ni
831 (phytomining) (Anderson et al. 2005). To date, most research in this area has focused on terrestrial
832 plants and soils contaminated through industrial mining (Rosenkranz et al., 2017). However, there
833 may be potential to explore metal-contaminated waters and sediments of wetlands used to treat
834 mine-tailing effluents. The usefulness of this process depends on the current market value of target

835 metals and the economic benefits associated with this form of phytoremediation(Sheoran et al.,
836 2009).

837 The use of macrophytes as biofuels is another possibility and is a feasible option to increase the
838 value of phytoremediation if there is a market for biomass. An economic assessment by Jiang et al.
839 (2015) found that high biomass production plants are required to make this a profitable venture.
840 However, different options need to be considered in pre-treatment, such as de-wetting and
841 briquetting, since fresh plant biomass comprises up to 90% water(Newete and Byrne,
842 2016). Macrophyte biomass may also be used for animal feed, or to make compost or
843 biochar(Quilliam et al., 2015; Tanaka et al. 2017). Quilliam et al. (2015) discussed in detail the issues
844 with these PBRs in terms of the transfer of pathogens, bio-magnification of heavy metals and
845 propagation of invasive species. A phytoremediation decision-making system that couples the target
846 pollutants and the PBRs would allow the resource recovery options to be established early in the
847 process (Song and Park, 2017). For example, the remediation of a eutrophic lake would seem to link
848 well with composting or animal feed PBRs given the potential for high nutritional content. However,
849 if heavy metal or pesticide contamination also is identified, then a biofuel or phytomining PBRs may
850 be more appropriate. Larger scale pilot tests of aquatic phytoremediation are required, and these
851 should explore the feasibility of using produced biomass in PBRs.

852 **12.11 Summary and future perspectives**

853
854 This chapter has outlined the potential of aquatic phytoremediation to provide efficient,
855 multi-targeted and sustainable remediation solutions for polluted waters. A summary of a proposed
856 research agenda required to fulfil the potential of these systems is presented in Table 12.7. Given
857 the wide range of organic, inorganic and biological pollutants that can impact surface waters there is
858 a need to steer phytoremediation towards a context-specific approach that allows the remediation
859 of multiple water body types, and waters affected by a range of pollutants.

860 With the development of novel ways to deploy macrophytes, such as by FTWs, there are
861 emerging options for spatial flexibility of applying phytoremediation, which are relatively
862 inexpensive. Larger scale pilot studies are required in this respect to assess the realistic
863 opportunities for use. At present there are a wide range of macrophytes of different growth forms
864 that have been established as efficient accumulators of pollutants. A further focus is required to
865 investigate the remediation potential of submerged species and to establish new accumulators that
866 may be used. Importantly, some of the key hyperaccumulators are considered invasive and would be
867 unsuitable to be deployed in natural surface waters. A proposed advancement for phytoremediation
868 systems is to consider the benefits of a plant community based-approach that assembles
869 polycultures of macrophytes with good accumulation capacity for different pollutants, enabling
870 multi-targeted remediation. Here, the need for a logical system of macrophyte selection based on
871 plant removal efficiencies and environmental tolerances, and target pollutant specifications,
872 requires development.

873 The process of macrophyte phytoremediation still requires a deeper understanding of how
874 to enhance removal efficiency and ensure sustainable harvesting of macrophytes. Understanding the
875 spatial and temporal dynamics of pollutant translocation within macrophytes is crucial for
876 permanent pollutant removal from water and for maintaining the economic value of different PBRs.
877 Furthermore, a 'meta-organism' approach needs to be considered in future phytoremediation
878 studies to establish the role of plant-associated microbial communities. There may be untapped
879 potential in manipulating these microbial communities for enhanced performance.

880 Finally, the focus of phytoremediation has been on the water treatment aspect, whilst there
881 is growing recognition of the capacity of these ecological engineering strategies to provide
882 ecosystem services such as carbon sequestration and biodiversity support. These benefits need to be
883 better quantified to determine the added-value of phytoremediation. With the waste management
884 sector shifting towards a life-cycle approach, there are clear opportunities for resource recovery

885 through identifying PBRs such as composting, biofuel production and animal feed. These PBRs
886 require further exploration in terms of their safety, value and ability to link directly with the target
887 pollutants removed (Figure 12.8). A life-cycle approach needs to be embedded in prospective aquatic
888 phytoremediation projects, to ensure that target pollutant(s) are being considered in tandem with
889 the PBR, whilst the frequency of harvest and replacement/regrowth of macrophytes is properly
890 linked into the remediation of the target pollutant (Figure 12.8).

891

892 **Acknowledgements**

893 Funding for this work was provided by the Scottish Government HydroNation Scholars Programme.

894

- Abed, S. N., Almuktar, S. A. and Scholz, M. (2017) 'Remediation of synthetic greywater in mesocosm—Scale floating treatment wetlands', *Ecological Engineering*. Elsevier B.V., 102, pp. 303–319. doi: 10.1016/j.ecoleng.2017.01.043.
- Afrous, A., Manshour, M., Liaghat, A., Pazira, E. and Sedghi, H. (2011) 'Mercury and arsenic accumulation by three species of aquatic plants in Dezful, Iran', *African Journal of Agricultural Research*, 6(24), pp. 5391–5397. doi: 10.5897/AJAR10.818.
- Akeel, K. (2013) *Empirical investigation of water pollution control through use of Phragmites australis A thesis submitted for the degree of Doctor of Philosophy by Khaled Al Akeel ETC, Graduate School Abstract*. Brunel University.
- Ali, H., Khan, E. and Sajad, M. A. (2013) 'Phytoremediation of heavy metals—Concepts and applications', *Chemosphere*, 91(7), pp. 869–881. doi: 10.1016/j.chemosphere.2013.01.075.
- Anderson, C., Moreno, F. and Meech, J. (2005) 'A field demonstration of gold phytoextraction technology', *Minerals Engineering*, 18(4), pp. 385–392. doi: 10.1016/j.mineng.2004.07.002.
- Anning, A. K., Korsah, P. E. and Addo-Fordjour, P. (2013) 'Phytoremediation of Wastewater with *Limnocharis Flava*, *Thalia Geniculata* and *Typha Latifolia* in Constructed Wetlands', *International Journal of Phytoremediation*, 15(5), pp. 452–464. doi: 10.1080/15226514.2012.716098.
- Ansa, E. D. O., Awuah, E., Andoh, A., Banu, R., Dorgbetor, W. H. K., Lubberding, H. J. and Gijzen, H. J. (2015) 'A Review of the Mechanisms of Faecal Coliform Removal from Algal and Duckweed Waste Stabilization Pond Systems', *American Journal of Environmental Sciences*, 11, pp. 28–34. doi: 10.3844/ajessp.2015.28.34.
- Ansari, A. A., Gill, S. S., Khan, F. A. and Naeem, M. (2014) 'Phytoremediation Systems for the Recovery of Nutrients from Eutrophic Waters', in *Eutrophication: Causes, Consequences and Control*. Dordrecht: Springer Netherlands, pp. 239–248. doi: 10.1007/978-94-007-7814-6_17.
- Arora, A., Saxena, S. and Sharma, D. K. (2006) 'Tolerance and phytoaccumulation of Chromium by three *Azolla* species', *World Journal of Microbiology & Biotechnology*, 22, pp. 97–100. doi: 10.1007/s11274-005-9000-9.
- Awuah, E. (2006) *The role of attachment in the removal of faecal bacteria from macrophyte and algal waste stabilization ponds. Pathogen removal mechanisms in macrophyte and algal waste stabilization ponds*. Wageningen University .
- Awuah, E. and Gyasi, S. (2014) 'Role of Protozoa on Faecal Bacteria Removal in Macrophyte and Algal Waste Stabilization Ponds', *Microbiology Journal*, 2, pp. 41–50. doi: 10.3923/mj.2014.41.50.
- Ayaz, S. Ç. and Saygin, O. (1996) *Hydroponic tertiary treatment*, *Water Research*. doi: 10.1016/0043-1354(95)00284-7.
- Ayyasamy, P. M., Rajakumar, S., Sathishkumar, M., Swaminathan, K., Shanthi, K., Lakshmanaperumalsamy, P. and Lee, S. (2009) 'Nitrate removal from synthetic medium and groundwater with aquatic macrophytes', *Desalination*, 242(1), pp. 286–296. doi: 10.1016/j.desal.2008.05.008.
- Barber, J. T., Sharma, H. A., Ensley, H. E., Polito, M. A. and Thomas, D. A. (1995) 'Detoxification of phenol by the aquatic angiosperm, *Lemna gibba*', *Chemosphere*. Pergamon, 31(6), pp. 3567–3574. doi: 10.1016/0045-6535(95)00205-M.

- Barrat-Segretain, M. (2001) 'Biomass allocation in three macrophyte species in relation to the disturbance level of their habitat', *Freshwater Biology*. Blackwell Science Ltd, 46(7), pp. 935–945. doi: 10.1046/j.1365-2427.2001.00728.x.
- Bartodziej, W. M., Blood, S. L. and Pilgrim, K. (2017) 'Aquatic plant harvesting: An economical phosphorus removal tool in an urban shallow lake', *J. Aquat. Plant Manage*, 55, pp. 26–34. Available at: <http://www.apms.org/wp/wp-content/uploads/japm-55-01-26.pdf> (Accessed: 27 February 2018).
- Bennicelli, R., Stępniewska, Z., Banach, A., Szajnocha, K. and Ostrowski, J. (2004) The ability of *Azolla caroliniana* to remove heavy metals (Hg(II), Cr(III), Cr(VI)) from municipal waste water, *Chemosphere*. doi: 10.1016/j.chemosphere.2003.11.015.
- Berger, E., Haase, P., Kuemmerlen, M., Leps, M., Schäfer, R. B. and Sundermann, A. (2017) 'Water quality variables and pollution sources shaping stream macroinvertebrate communities', *Science of the Total Environment*, 587588, pp. 1–10. doi: 10.1016/j.scitotenv.2017.02.031.
- Boonsong, K. and Chansiri, M. (2008) 'Efficiency of vetiver grass cultivated with floating platform technique in domestic wastewater treatment', *AU JOURNAL OF TECHNOLOGY*, 12(2), pp. 73–80.
- Borne, K. E. (2014) 'Floating treatment wetland influences on the fate and removal performance of phosphorus in stormwater retention ponds', *Ecological Engineering*, 69, pp. 76–82. doi: 10.1016/j.ecoleng.2014.03.062.
- Bouldin, J. L., Farris, J. L., Moore, M. T., Smith, S. and Cooper, C. M. (2006) 'Hydroponic uptake of atrazine and lambda-cyhalothrin in *Juncus effusus* and *Ludwigia peploides*', *Chemosphere*, 65(6), pp. 1049–1057. doi: 10.1016/j.chemosphere.2006.03.031.
- Broadley, M., Brown, P., Cakmak, I., Rengel, Z. and Zhao, F. (2011) *Function of Nutrients: Micronutrients, Marschner's Mineral Nutrition of Higher Plants: Third Edition*. doi: 10.1016/B978-0-12-384905-2.00007-8.
- Cao, W. and Zhang, Y. (2014) Removal of nitrogen (N) from hypereutrophic waters by ecological floating beds (EFBs) with various substrates, *Ecological Engineering*. doi: 10.1016/j.ecoleng.2013.10.018.
- Carbonell, A. ., Aarabi, M. ., DeLaune, R. ., Gambrell, R. and Patrick Jr, W. (1998) 'Arsenic in wetland vegetation: Availability, phytotoxicity, uptake and effects on plant growth and nutrition', *Science of The Total Environment*, 217(3), pp. 189–199. doi: 10.1016/S0048-9697(98)00195-8.
- Cardinal, P., Anderson, J. C., Carlson, J. C., Low, J. E., Challis, J. K., Beattie, S. A., Bartel, C. N., Elliott, A. D., Montero, O. F., Lokesh, S., Favreau, A., Kozlova, T. A., Knapp, C. W., Hanson, M. L. and Wong, C. S. (2014) 'Macrophytes may not contribute significantly to removal of nutrients, pharmaceuticals, and antibiotic resistance in model surface constructed wetlands', *Science of The Total Environment*, 482, pp. 294–304. doi: 10.1016/j.scitotenv.2014.02.095.
- Carpenter, S. R. and Adams, M. S. (1977) 'The macrophyte tissue nutrient pool of a hardwater eutrophic lake: Implications for macrophyte harvesting', *Aquatic Botany*. Elsevier, 3, pp. 239–255. doi: 10.1016/0304-3770(77)90026-2.
- Chambers, P. A., Lacoul, A. P., Murphy, A. K. J., Thomaz, A. S. M., Lacoul, P., Murphy, K. J. and Thomaz, S. M. (2008) 'Global diversity of aquatic macrophytes in freshwater', *Hydrobiologia*, 595, pp. 9–26. doi: 10.1007/s10750-007-9154-6.

- Chandra, R. and Yadav, S. (2011) 'Phytoremediation of CD, CR, CU, MN, FE, NI, PB and ZN from Aqueous Solution Using Phragmites Cummunis, Typha Angustifolia and Cyperus Esculentus', *International Journal of Phytoremediation*. Taylor & Francis Group, 13(6), pp. 580–591. doi: 10.1080/15226514.2010.495258.
- Chang, Y.-H., Ku, C.-R. and Lu, H.-L. (2014) 'Effects of aquatic ecological indicators of sustainable green energy landscape facilities', *Ecological Engineering*, 71, pp. 144–153. doi: 10.1016/j.ecoleng.2014.07.051.
- Chen, Y., Vymazal, J., Březinová, T., Koželuh, M., Kule, L., Huang, J. and Chen, Z. (2016) 'Occurrence, removal and environmental risk assessment of pharmaceuticals and personal care products in rural wastewater treatment wetlands', *Science of the Total Environment*, xxx. doi: 10.1016/j.scitotenv.2016.06.069.
- Chen, Z., Cuervo, D., Müller, J., Wiessner, A., Köser, H., Vymazal, J., Kästner, M. and Kuschik, P. (2016) 'Hydroponic root mats for wastewater treatment—a review', *Environmental Science and Pollution Research*. Springer Berlin Heidelberg, 23(16), pp. 15911–15928. doi: 10.1007/s11356-016-6801-3.
- Cheng, S. (2003) 'Heavy metal pollution in China: Origin, pattern and control', *Environmental Science and Pollution Research*. Ecomed, 10(3), pp. 192–198. doi: 10.1065/espr2002.11.141.1.
- Coleman, J., Hensch, K., Garbutt, K., Sexstone, A., Bissonnette, G. and Skousen, J. (2001) 'Treatment of Domestic Wastewater by Three Plant Species in Constructed Wetlands', *Water, Air, and Soil Pollution*. Kluwer Academic Publishers, 128(3/4), pp. 283–295. doi: 10.1023/A:1010336703606.
- Dai, Y., Jia, C., Liang, W., Hu, S. and Wu, Z. (2012) Effects of the submerged macrophyte *Ceratophyllum demersum* L. on restoration of a eutrophic waterbody and its optimal coverage, *Ecological Engineering*. doi: 10.1016/j.ecoleng.2011.12.023.
- Dawson, F., Clinton, E. and Ladle, M. (1991) 'Invertebrates on cut weed removed during weed-cutting operations along an English river, the River Frome, Dorset', *Aquaculture Research*. Blackwell Publishing Ltd, 22(1), pp. 113–132. doi: 10.1111/j.1365-2109.1991.tb00500.x.
- Decamp, O. and Warren, A. (2000) Investigation of *Escherichia coli* removal in various designs of subsurface flow wetlands used for wastewater treatment, *Ecological Engineering*. doi: 10.1016/S0925-8574(99)00007-5.
- Demars, B. O. L. and Edwards, A. C. (2007) 'Tissue nutrient concentrations in freshwater aquatic macrophytes: High inter-taxon differences and low phenotypic response to nutrient supply', *Freshwater Biology*, 52(11), pp. 2073–2086. doi: 10.1111/j.1365-2427.2007.01817.x.
- Deng, H., Ye, Z. . and Wong, M. . (2004) 'Accumulation of lead, zinc, copper and cadmium by 12 wetland plant species thriving in metal-contaminated sites in China', *Environmental Pollution*, 132(1), pp. 29–40. doi: 10.1016/j.envpol.2004.03.030.
- Denny, P. (1972) 'Sites of Nutrient Absorption in Aquatic Macrophytes', *Source Journal of Ecology*, 60(3), pp. 819–829. Available at: <http://www.jstor.org/stable/2258568> (Accessed: 4 January 2018).
- Dettenmaier, E., Doucette, W. and Bugbee, B. (2009) 'Chemical Hydrophobicity and Uptake by Plant Roots', *Environmental science & technology*, 43, pp. 324–329. doi: 10.1021/es801751x.
- Dhir, B. (2013) *Phytoremediation: Role of Aquatic Plants in Environmental Clean-Up*. Springer India. doi: 10.1007/978-81-322-1307-9.

- Dhir, B., Sharmila, P. and Saradhi, P. P. (2008) 'Photosynthetic performance of *Salvinia natans* exposed to chromium and zinc rich wastewater', *Brazilian Journal of Plant Physiology*. Sociedade Brasileira de Fisiologia Vegetal, 20(1), pp. 61–70. doi: 10.1590/S1677-04202008000100007.
- Dhote, S. and Dixit, S. (2009) 'Water quality improvement through macrophytes - A review', *Environmental Monitoring and Assessment*, 152(1–4), pp. 149–153. doi: 10.1007/s10661-008-0303-9.
- Dosnon-Olette, R., Couderchet, M., Oturan, M. A., Oturan, N. and Eullaffroy, P. (2011) 'Potential Use of *Lemna Minor* for the Phytoremediation of Isoproturon and Glyphosate', *International Journal of Phytoremediation*, 13(6), pp. 601–612. doi: 10.1080/15226514.2010.525549.
- Du, W., Li, Z., Zhang, Z., Jin, Q., Chen, X. and Jiang, S. (2017) 'Composition and Biomass of Aquatic Vegetation in the Poyang Lake, China', *Scientifica*. doi: 10.1155/2017/8742480.
- Duman, F., Mehmet, A., Ae, C. and Sezen, G. (2007) 'Seasonal changes of metal accumulation and distribution in common club rush (*Schoenoplectus lacustris*) and common reed (*Phragmites australis*)'. doi: 10.1007/s10646-007-0150-4.
- Dunn, S., Brown, I., Sample, J. and Post, H. (2012) 'Relationships between climate, water resources, land use and diffuse pollution and the significance of uncertainty in climate change', *Journal of Hydrology*, (434–435), pp. 19–35.
- Edwards, P. (2015) 'Aquaculture environment interactions: Past, present and likely future trends', *Aquaculture*, 447, pp. 2–14. doi: 10.1016/j.aquaculture.2015.02.001.
- Eichert, T. and Fernández, V. (2011) *Uptake and Release of Elements by Leaves and Other Aerial Plant Parts, Marschner's Mineral Nutrition of Higher Plants: Third Edition*. Elsevier Ltd. doi: 10.1016/B978-0-12-384905-2.00004-2.
- Eid, E. M., Shaltout, K. H., El-Sheikh, M. A. and Asaeda, T. (2012) 'Seasonal courses of nutrients and heavy metals in water, sediment and above- and below-ground *Typha domingensis* biomass in Lake Burullus (Egypt): Perspectives for phytoremediation', *Flora - Morphology, Distribution, Functional Ecology of Plants*, 207(11), pp. 783–794. doi: 10.1016/j.flora.2012.09.003.
- El-Kheir, W., Ismail, G., Farid, A. E., Tarek, T. and Hammad, D. (2007) 'Assessment of the Efficiency of Duckweed (*Lemna gibba*) in Wastewater Treatment', *International Journal of Agriculture and Biology*, 9(5), pp. 681–687.
- El-Shahawi, M. S., Hamza, A., Bashammakh, A. S. and Al-Saggaf, W. T. (2010) 'An overview on the accumulation, distribution, transformations, toxicity and analytical methods for the monitoring of persistent organic pollutants', *Talanta*, 80(5), pp. 1587–1597. doi: 10.1016/j.talanta.2009.09.055.
- Engelhardt, K. A. M. and Ritchie, M. E. (2002) 'The Effect of Aquatic Plant Species Richness on Wetland Ecosystem Processes', *Ecology*, 83(10), pp. 2911–2924.
- Engelhardt, K. a and Ritchie, M. E. (2001) 'Effects of macrophyte species richness on wetland ecosystem functioning and services.', *Nature*, 411(6838), pp. 687–689. doi: 10.1038/35079573.
- van der Ent, A., Baker, A. J. M., Reeves, R. D., Pollard, A. J. and Schat, H. (2013) 'Hyperaccumulators of metal and metalloid trace elements: Facts and fiction', *Plant and Soil*. Springer Netherlands, 362(1–2), pp. 319–334. doi: 10.1007/s11104-012-1287-3.

- Faulwetter, J. L., Gagnon, V., Sundberg, C., Chazarenc, F., Burr, M. D., Brisson, J., Camper, A. K. and Stein, O. R. (2009) 'Microbial processes influencing performance of treatment wetlands: A review', *Ecological Engineering*, 35(6), pp. 987–1004. doi: 10.1016/j.ecoleng.2008.12.030.
- Feng, N.-X., Yu, J., Zhao, H.-M., Cheng, Y.-T., Mo, C.-H., Cai, Q.-Y., Li, Y.-W., Li, H. and Wong, M.-H. (2017) 'Efficient phytoremediation of organic contaminants in soils using plant–endophyte partnerships', *Science of The Total Environment*. doi: 10.1016/j.scitotenv.2017.01.075.
- Fernandez, R. T., Whitwell, T., Riley, M. B. and Bernard, C. R. (1999) 'Evaluating semiaquatic herbaceous perennials for use in herbicide phytoremediation', *Journal of the American Society for Horticultural Science*, 124(5), pp. 539–544.
- Fraser, L. H., Carty, S. M. and Steer, D. (2004) 'A test of four plant species to reduce total nitrogen and total phosphorus from soil leachate in subsurface wetland microcosms', *Bioresource Technology*, 94(2), pp. 185–192. doi: 10.1016/j.biortech.2003.11.023.
- Fu, F. and Wang, Q. (2011) 'Removal of heavy metal ions from wastewaters: A review', *Journal of Environmental Management*, 92, pp. 407–418. doi: 10.1016/j.jenvman.2010.11.011.
- Fuhrmann, S., Nauta, M., Pham-Duc, P., Tram, N. T., Nguyen-Viet, H., Utzinger, J., Cissé, G. and Winkler, M. S. (2017) 'Disease burden due to gastrointestinal infections among people living along the major wastewater system in Hanoi, Vietnam', *Advances in Water Resources*. Elsevier, 108, pp. 439–449. doi: 10.1016/J.ADVWATRES.2016.12.010.
- Gabrielson, J. O., Perkins, M. A. and Welch, E. B. (1984) 'The uptake, translocation and release of phosphorus by *Elodea densa*', *Hydrobiologia*. Kluwer Academic Publishers, 111(1), pp. 43–48. doi: 10.1007/BF00007379.
- Gao, J., Garrison, A. W., Hoehamer, C., Mazur, C. S. and Wolfe, N. L. (2000) 'Uptake and phytotransformation of o,p'-DDT and p,p'-DDT by axenically cultivated aquatic plants', *Journal of Agricultural and Food Chemistry*, 48(12), pp. 6121–6127. doi: 10.1021/jf990956x.
- Garrison, A. W., Nzungu, V. A., Avents, J. K., Ellington, J. J., Jones, W. J., Rennels, D. and Wolfe, N. L. E. E. (2000) 'Phyrodgradation of p,p'-DDT and the Enantiomers of o,p'-DDT', *Environ. Sci. Technol.*, 34(9), pp. 1663–1670.
- Garver, E. G., Dubbe, D. R. and Pratt, D. C. (1988) 'Seasonal patterns in accumulation and partitioning of biomass and macronutrients in *Typha* spp.', *Aquatic Botany*. Elsevier, 32(1–2), pp. 115–127. doi: 10.1016/0304-3770(88)90092-7.
- Ge, Y., Han, W., Huang, C., Wang, H., Liu, D., Chang, S. X., Gu, B., Zhang, C., Gu, B., Fan, X., Du, Y. and Chang, J. (2015) 'Positive effects of plant diversity on nitrogen removal in microcosms of constructed wetlands with high ammonium loading', *Ecological Engineering*. Elsevier, 82, pp. 614–623. doi: 10.1016/J.ECOLENG.2015.05.030.
- Ge, Z., Feng, C., Wang, X. and Zhang, J. (2016) 'Seasonal applicability of three vegetation constructed floating treatment wetlands for nutrient removal and harvesting strategy in urban stormwater retention ponds', *International Biodeterioration & Biodegradation*. Elsevier, 112, pp. 80–87. doi: 10.1016/J.IBIOD.2016.05.007.
- Geissen, V., Mol, H., Klumpp, E., Umlauf, G., Nadal, M., van der Ploeg, M., van de Zee, S. E. A. T. M. and Ritsema, C. J. (2015) 'Emerging pollutants in the environment: A challenge for water resource management', *International Soil and Water Conservation Research*, 3(1), pp. 57–65. doi:

10.1016/j.iswcr.2015.03.002.

Geng, Y., Han, W., Yu, C., Jiang, Q., Wu, J., Chang, J. and Ge, Y. (2017) 'Effect of plant diversity on phosphorus removal in hydroponic microcosms simulating floating constructed wetlands', *Ecological Engineering*, 107, pp. 110–119. doi: 10.1016/j.ecoleng.2017.06.061.

Glick, B. R. (2003) 'Phytoremediation: synergistic use of plants and bacteria to clean up the environment', *Biotechnology Advances*, 21(5), pp. 383–393. doi: 10.1016/S0734-9750(03)00055-7.

Gomes, H. I. (2012) 'Phytoremediation for bioenergy: challenges and opportunities', *Environmental Technology Reviews*. Taylor & Francis, 1(1), pp. 59–66. doi: 10.1080/09593330.2012.696715.

Gomes, M. A. da C., Hauser-Davis, R. A., de Souza, A. N. and Vitória, A. P. (2016) 'Metal phytoremediation: General strategies, genetically modified plants and applications in metal nanoparticle contamination', *Ecotoxicology and Environmental Safety*, 134, pp. 133–147. doi: 10.1016/j.ecoenv.2016.08.024.

Gulati, R. D., Dionisio Pires, L. M. and Van Donk, E. (2008) 'Lake restoration studies: Failures, bottlenecks and prospects of new ecotechnological measures', *Limnologica - Ecology and Management of Inland Waters*. Urban & Fischer, 38(3–4), pp. 233–247. doi: 10.1016/J.LIMNO.2008.05.008.

Gumbricht, T. (1993) 'Review Nutrient removal processes in freshwater submersed macrophyte systems', *Ecological Engineering Elsevier Science Publishers B.V*, 2, pp. 1–30.

Guo, Y., Liu, Y., Zeng, G., Hu, X., Li, X., Huang, D., Liu, Y. and Yin, Y. (2014) 'A restoration-promoting integrated floating bed and its experimental performance in eutrophication remediation', *Journal of Environmental Sciences*. Elsevier, 26(5), pp. 1090–1098. doi: 10.1016/S1001-0742(13)60500-8.

Ha, N. T. H., Sakakibara, M. and Sano, S. (2009) 'Phytoremediation of Sb, As, Cu, and Zn from contaminated water by the aquatic macrophyte *Eleocharis acicularis*', *Clean - Soil, Air, Water*, 37(9), pp. 720–725. doi: 10.1002/clen.200900061.

Haack, S. K., Duris, J. W., Kolpin, D. W., Focazio, M. J., Meyer, M. T., Johnson, H. E., Oster, R. J. and Foreman, W. T. (2016) 'Contamination with bacterial zoonotic pathogen genes in U.S. streams influenced by varying types of animal agriculture', *Science of the Total Environment*, 563–564, pp. 340–350. doi: 10.1016/j.scitotenv.2016.04.087.

Habib, S. and AR, Y. (2016) 'Impact of different Harvesting Techniques on the Population of Macrophyte-associated-Invertebrate Community in an Urban Lake', *Journal of Pollution Effects & Control*. OMICS International, 4(2), pp. 1–3. doi: 10.4172/2375-4397.1000158.

Hafez, N., Abdalla, S. and Ramadan, Y. S. (1998) 'Accumulation Of Phenol By *Potamogeton crispus* from Aqueous Industrial Waste', *Bull. Environ. Contam. Toxicol*, 60, pp. 944–948.

Hancock, M. (2000) 'Artificial floating islands for nesting Black-throated Divers *Gavia arctica* in Scotland: construction, use and effect on breeding success', *Bird Study*, 47, pp. 165–175.

Hawkesford, M., Horst, W., Kichey, T., Lambers, H., Schjoerring, J., Miller, I. S. and White, P. (2011) *Functions of Macronutrients, Marschner's Mineral Nutrition of Higher Plants: Third Edition*. Elsevier Ltd. doi: 10.1016/B978-0-12-384905-2.00006-6.

Haygarth, P., Jarvie, H. and Powers, S. (2014) 'Sustainable Phosphorus Management and the Need

for a Long-Term Perspective: The Legacy Hypothesis', *Environmental Science & Technology*, 48, pp. 8417–8419.

Headley, T. R. and Tanner, C. C. (2008) 'Floating Treatment Wetlands: an Innovative Option for Stormwater Quality Applications', *11th International Conference on Wetland Systems for Water Pollution Control*.

Heathwaite, A. L. (2010) 'Multiple stressors on water availability at global to catchment scales: Understanding human impact on nutrient cycles to protect water quality and water availability in the long term', *Freshwater Biology*, 55(SUPPL. 1), pp. 241–257. doi: 10.1111/j.1365-2427.2009.02368.x.

Heisler, J., Glibert, P. M., Burkholder, J. M., Anderson, D. M., Cochlan, W., Dennison, W. C., Dortch, Q., Gobler, C. J., Heil, C. A., Humphries, E., Lewitus, A., Magnien, R., Marshall, H. G., Sellner, K., Stockwell, D. A., Stoecker, D. K. and Suddleson, M. (2008) 'Eutrophication and harmful algal blooms: A scientific consensus', *Harmful Algae*. Elsevier, 8(1), pp. 3–13. doi: 10.1016/J.HAL.2008.08.006.

Hilt, S., Gross, E. M., Hupfer, M., Morscheid, H., Malmann, J., Melzer, A., Poltz, J., Sandrock, S., Scharf, E.-M., Schneider, S. and Van De Weyer H, K. (2006) 'Restoration of submerged vegetation in shallow eutrophic lakes – A guideline and state of the art in Germany ARTICLE IN PRESS', *Limnologia*, 36, pp. 155–171. doi: 10.1016/j.limno.2006.06.001.

Hirneisen, K. A., Sharma, M. and Kniel, K. E. (2012) 'Human Enteric Pathogen Internalization by Root Uptake into Food Crops', *Foodborne Pathogens and Disease*, 9(5), pp. 396–405. doi: 10.1089/fpd.2011.1044.

Houda, N., Hanene, C., Ines, M., Said Myriam, B., Imen, D. and Abdennaceur, H. (2014) 'African Journal of Microbiology Research Isolation and characterization of microbial communities from a constructed wetlands system: A case study in Tunisia', 8(6), pp. 529–538. doi: 10.5897/AJMR2013.6493.

Hu, C., Zhang, L., Hamilton, D., Zhou, W., Yang, T. and Zhu, D. (2007) 'Physiological responses induced by copper bioaccumulation in *Eichhornia crassipes* (Mart.)', *Hydrobiologia*. Kluwer Academic Publishers, 579(1), pp. 211–218. doi: 10.1007/s10750-006-0404-9.

Hu, M.-H., Yuan, J.-H., Yang, X.-E. and He, Z.-L. (2010) 'Effects of temperature on purification of eutrophic water by floating eco-island system', *Acta Ecologica Sinica*, 30(6), pp. 310–318. doi: 10.1016/j.chnaes.2010.06.009.

Huser, B. J., Futter, M., Lee, J. T. and Perniel, M. (2016) 'In-lake measures for phosphorus control: The most feasible and cost-effective solution for long-term management of water quality in urban lakes', *Water Research*, 97, pp. 142–152. doi: 10.1016/j.watres.2015.07.036.

Islam, M. S., Ueno, Y., Sikder, M. T. and Kurasaki, M. (2013) 'Phytofiltration of Arsenic and Cadmium From the Water Environment Using *Micranthemum Umbrosum* (J.F. Gmel) S.F. Blake As A Hyperaccumulator', *International Journal of Phytoremediation*. Taylor & Francis Group, 15(10), pp. 1010–1021. doi: 10.1080/15226514.2012.751356.

Jackson, L. . (1998) 'Paradigms of metal accumulation in rooted aquatic vascular plants', *Science of The Total Environment*, 219(2), pp. 223–231. doi: 10.1016/S0048-9697(98)00231-9.

Javadi, E., Moattar, F., Karbassi, A. R. and Monavari, S. M. (2005) 'Removal of lead, cadmium and manganese from liquid solution using water lily (*Nymphaea alba*)', *Journal of Food, Agriculture & Environment Journal of Food Agriculture & Environment*, 88(4), pp. 1220–1225.

- Jones, D. L., Cross, P., Withers, P. J. a., DeLuca, T. H., Robinson, D. a., Quilliam, R. S., Harris, I. M., Chadwick, D. R. and Edwards-Jones, G. (2013) 'REVIEW: Nutrient stripping: the global disparity between food security and soil nutrient stocks', *Journal of Applied Ecology*. Edited by P. Kardol, 50(4), pp. 851–862. doi: 10.1111/1365-2664.12089.
- Jones, T. G., Willis, N., Gough, R. and Freeman, C. (2017) 'An experimental use of floating treatment wetlands (FTWs) to reduce phytoplankton growth in freshwaters', *Ecological Engineering*. Elsevier B.V., 99, pp. 316–323. doi: 10.1016/j.ecoleng.2016.11.002.
- Kadlec, R. H. (2009) 'Comparison of free water and horizontal subsurface treatment wetlands', *Ecological Engineering*, 35(2), pp. 159–174. doi: 10.1016/j.ecoleng.2008.04.008.
- Kadlec, R. H. and Wallace, S. D. (2009) *Treatment Wetlands, Second Edition, Treatment Wetlands, Second Edition*. doi: 10.1201/9781420012514.
- Kamal, M., Ghaly, A. E., Mahmoud, N. and Côté, R. (2004) 'Phytoaccumulation of heavy metals by aquatic plants', *Environment International*, 29(8), pp. 1029–1039. doi: 10.1016/S0160-4120(03)00091-6.
- Kamarudzaman, A. and Ismail, N. (2011) 'Removal of nutrients from landfill leachate using subsurface flow constructed wetland planted with *Limncharis flava* and *Scirpus atrovirens*', ... on *Environmental and ...*, 19, pp. 79–83.
- Kansiime, F., Oryem-Origa, H. and Rukwago, S. (2005) 'Comparative assessment of the value of papyrus and cocoyams for the restoration of the Nakivubo wetland in Kampala, Uganda', *Physics and Chemistry of the Earth, Parts A/B/C*, 30(11), pp. 698–705. doi: 10.1016/j.pce.2005.08.010.
- Kara, Y. (2010) 'Bioaccumulation of nickel by aquatic macrophytes', *Desalination and Water Treatment*, 19, pp. 325–328.
- Karathanasis, A. D., Potter, C. L. and Coyne, M. S. (2003) 'Vegetation effects on fecal bacteria, BOD, and suspended solid removal in constructed wetlands treating domestic wastewater', *Ecological Engineering*, 20(2), pp. 157–169. doi: 10.1016/S0925-8574(03)00011-9.
- Karim, M. R., Manshadi, F. D., Karpiscak, M. M. and Gerba, C. P. (2004) 'The persistence and removal of enteric pathogens in constructed wetlands', *Water Research*, 38(7), pp. 1831–1837. doi: 10.1016/j.watres.2003.12.029.
- Karnchanawong, S. (1995) 'Comparative study of domestic wastewater treatment efficiencies between facultative pond and water spinach pond', *Water Science and Technology*, 32(3), pp. 263–270. doi: 10.1016/0273-1223(95)00627-3.
- Karpiscak, M. M., Gerba, C. P., Watt, P. M., Foster, K. E. and Falabi, J. A. (1996) 'Multi-species plant systems for wastewater quality improvements and habitat enhancement', *Water Science and Technology*. 33(10), pp. 231–236. doi: 10.1016/0273-1223(96)00424-6.
- Keenen, K. and Kirkwood, N. (2015) *PHYTO Principles and resources for site remediation and landscape design*. 1st edn. Oxton: Routledge.
- Keizer-Vlek, H. E., Verdonschot, P. F. M., Verdonschot, R. C. M. and Dekkers, D. (2014) 'The contribution of plant uptake to nutrient removal by floating treatment wetlands', *Ecological Engineering*, 73, pp. 684–690. doi: 10.1016/j.ecoleng.2014.09.081.

Kennen, K. and Kirkwood, N. (2015) *Phyto: Principles and Resources for Site Remediation and Landscape Design*. 1st edn. Oxon: Routledge.

Kiiskila, J. D., Sarkar, D., Feuerstein, K. A. and Datta, R. (2017) 'A preliminary study to design a floating treatment wetland for remediating acid mine drainage-impacted water using vetiver grass (*Chrysopogon zizanioides*) of variance DO Dissolved oxygen EC Electric conductivity FTWs Floating treatment wetlands HSD Honest', *Environ Sci Pollut Res*, 24, pp. 27985–27993. doi: 10.1007/s11356-017-0401-8.

Kintu Sekiranda, S. B. and Kiwanuka, S. (1997) 'A study of nutrient removal efficiency of *Phragmites mauritianus* in experimental reactors in Uganda', *Hydrobiologia*. Kluwer Academic Publishers, 364(1), pp. 83–91. doi: 10.1023/A:1003166924903.

Kipasika, H. J., Buza, J., Smith, W. A. and Njau, K. N. (2016) 'African Journal of Microbiology Research Removal capacity of faecal pathogens from wastewater by four wetland vegetation: *Typha latifolia*, *Cyperus papyrus*, *Cyperus alternifolius* and *Phragmites australis*', 10(19), pp. 654–661. doi: 10.5897/AJMR2016.7931.

Kivaisi, A. K. (2001) 'The potential for constructed wetlands for wastewater treatment and reuse in developing countries: A review', *Ecological Engineering*, 16(4), pp. 545–560. doi: 10.1016/S0925-8574(00)00113-0.

Koelbener, A., Ramseier, D. and Suter, M. (2008) 'Competition alters plant species response to nickel and zinc', *Plant and Soil*, 303(1–2), pp. 241–251. doi: 10.1007/s11104-007-9503-2.

Körner, S. and Vermaat, J. E. (1998) 'The relative importance of *Lemna gibba* L., bacteria and algae for the nitrogen and phosphorus removal in duckweed-covered domestic wastewater', *Water Research*, 32(12), pp. 3651–3661. doi: 10.1016/S0043-1354(98)00166-3.

Kuiper, J. J., Verhofstad, M. J. J. M., Louwers, E. L. M., Bakker, E. S., Brederveld, R. J., van Gerven, L. P. A., Janssen, A. B. G., de Klein, J. J. M. and Mooij, W. M. (2017) 'Mowing Submerged Macrophytes in Shallow Lakes with Alternative Stable States: Battling the Good Guys?', *Environmental management*, 59(4), pp. 619–634. doi: 10.1007/s00267-016-0811-2.

Kumar Mishra, V. and Tripathi, B. (2008) 'Concurrent removal and accumulation of heavy metals by the three aquatic macrophytes'. doi: 10.1016/j.biortech.2008.01.002.

Kutty, S. R. M., Ngatenah, S. N. I., Isa, M. H. and Malakahmad, A. (2009) 'Nutrients removal from municipal wastewater treatment plant effluent using *Eichhornia Crassipes*', *World Academy of Science, Engineering and Technology*, 36(12), pp. 828–833.

Kyambadde, J., Kansiime, F., Gumaelius, L. and Dalhammar, G. (2004) 'A comparative study of *Cyperus papyrus* and *Miscanthidium violaceum*-based constructed wetlands for wastewater treatment in a tropical climate', *Water Research*, 38(2), pp. 475–485. doi: 10.1016/j.watres.2003.10.008.

Ladislav, S., Gérente, C., Chazarenc, F., Brisson, J. and André, Y. (2013) 'Performances of Two Macrophytes Species in Floating Treatment Wetlands for Cadmium, Nickel, and Zinc Removal from Urban Stormwater Runoff', *Water Air Soil Pollution*, 224, pp. 1408–1418. doi: 10.1007/s11270-012-1408-x.

Lam, Q. D., Schmalz, B., Fohrer, N. and Schmalz, B. (2011) 'The impact of agricultural Best Management Practices on water quality in a North German lowland catchment', *Environ Monit*

Assess, 183(183). doi: 10.1007/s10661-011-1926-9.

Landesman, L., Fedler, C. and Duan, R. (2011) 'Plant Nutrient Phytoremediation Using Duckweed', in Ansari, A. and Al., E. (eds) *Eutrophication: Causes, Consequences and Control*. Springer Science+Business Media, pp. 341–354. doi: 10.1007/9789048196258_17.

Lang Martins, A. P., Reissmann, C. B., Boeger, M. R. T., De Oliveira, E. B. and Favaretto, A. N. (2010) 'Efficiency of *Polygonum hydropiperoides* for Phytoremediation of Fish Pond Effluents Enriched with N and P', *J. Aquat. Plant Manage*, 48(48), pp. 116–120.

Lawford, R., Bogardi, J., Marx, S., Jain, S., Wostl, C. P., Knü Ppe, K., Ringler, C., Lansigan, F. and Meza, F. (2013) 'Basin perspectives on the Water–Energy–Food Security Nexus', *Current Opinion in Environmental Sustainability*, 5, pp. 607–616. doi: 10.1016/j.cosust.2013.11.005.

Lesage, E., Mundia, C., Rousseau, D. P. L., Van De Moortel, A. M. K., Du Laing, G., Tack, F. M. G., De Pauw, N. and Verloo, M. G. (2008) 'Removal of Heavy Metals from Industrial Effluents by the Submerged Aquatic Plant *Myriophyllum spicatum* L. 19.2 Material and Methods', in *Wastewater Treatment, Plan Dynamics and Management*, pp. 211–221.

Li, H., Hao, H., Yang, X., Xiang, L., Zhao, F., Jiang, H. and He, Z. (2012) 'Purification of Refinery Wastewater By Different Perennial Grasses Growing in a Floating Bed', *Journal of Plant Nutrition*, 35(1), pp. 93–110. doi: 10.1080/01904167.2012.631670.

Li, X.-N., Song, H.-L., Li, W., Lu, X.-W. and Nishimura, O. (2010) 'An integrated ecological floating-bed employing plant, freshwater clam and biofilm carrier for purification of eutrophic water', *Ecological Engineering*, 36(4), pp. 382–390. doi: 10.1016/j.ecoleng.2009.11.004.

Liang, M.-Q., Zhang, C.-F., Peng, C.-L., Lai, Z.-L., Chen, D.-F. and Chen, Z.-H. (2011) 'Plant growth, community structure, and nutrient removal in monoculture and mixed constructed wetlands', *Ecological Engineering*, 37(2), pp. 309–316. doi: 10.1016/j.ecoleng.2010.11.018.

Liebs, M. and Carsten Von Der Ohe, P. (2005) 'Analyzing effects of pesticides on invertebrate communities in streams', *Environmental Toxicology and Chemistry*, 24(4), pp. 954–965.

Lintelmann, J., Katayama, A., Kurihara, N., Shore, L. and Wenzel, A. A. (2003) 'Endocrine disruptors in the environment (IUPAC Technical Report) Endocrine disruptors in the environment', *Pure Appl. Chem. Schejbal (Czech Republic)*, 75(5), pp. 631–681.

Liu, Y., Sanguanphun, T., Yuan, W., Cheng, J. J. and Meetam, M. (2017) 'The biological responses and metal phytoaccumulation of duckweed *Spirodela polyrhiza* to manganese and chromium', *Environmental Science and Pollution Research*, 24(23), pp. 19104–19113. doi: 10.1007/s11356-017-9519-y.

Low, K. S., Lee, C. K. and Tai, C. H. (1994) 'Biosorption of copper by water hyacinth roots', *Journal of Environmental Science and Health . Part A: Environmental Science and Engineering and Toxicology*. Taylor & Francis Group , 29(1), pp. 171–188. doi: 10.1080/10934529409376028.

Lu, B., Xu, Z., Li, J. and Chai, X. (2018) 'Removal of water nutrients by different aquatic plant species: An alternative way to remediate polluted rural rivers', *Ecological Engineering*. Elsevier, 110(September 2017), pp. 18–26. doi: 10.1016/j.ecoleng.2017.09.016.

Lu, H.-L., Ku, C.-R. and Chang, Y.-H. (2015) 'Water quality improvement with artificial floating islands', *Ecological Engineering*, 74, pp. 371–375. doi: 10.1016/j.ecoleng.2014.11.013.

- Lu, Q., He, Z. L., Graetz, D. A., Stoffella, P. J. and Yang, X. (2010) 'Phytoremediation to remove nutrients and improve eutrophic stormwaters using water lettuce (*Pistia stratiotes* L.)', *Environmental Science and Pollution Research*, 17, pp. 84–96. doi: 10.1007/s11356-008-0094-0.
- Lu, Q., He, Z. L., Graetz, D. A., Stoffella, P. J. and Yang, X. (2011) 'Uptake and distribution of metals by water lettuce (*Pistia stratiotes* L.)', *Environmental Science and Pollution Research*. Springer-Verlag, 18(6), pp. 978–986. doi: 10.1007/s11356-011-0453-0.
- Lynch, J., Fox, L. J., Owen Jr., J. S. and Sample, D. J. (2015) 'Evaluation of commercial floating treatment wetland technologies for nutrient remediation of stormwater', *Ecological Engineering*, 75, pp. 61–69. doi: 10.1016/j.ecoleng.2014.11.001.
- Macek, T., Macková, M. and Káš, J. (2000) 'Exploitation of plants for the removal of organics in environmental remediation', *Biotechnology Advances*, 18(1), pp. 23–34. doi: 10.1016/S0734-9750(99)00034-8.
- Machado, A. I., Beretta, M., Fragoso, R. and Duarte, E. (2016) 'Overview of the state of the art of constructed wetlands for decentralized wastewater management in Brazil', *Journal of Environmental Management*. doi: 10.1016/j.jenvman.2016.11.015.
- Maine, M. A., Suñé, N. L. and Lager, S. C. (2004) 'Chromium bioaccumulation: comparison of the capacity of two floating aquatic macrophytes', *Water Research*, 38(6), pp. 1494–1501. doi: 10.1016/j.watres.2003.12.025.
- Makvana, K. S. and Sharma, M. K. (2013) 'Assessment of Pathogen Removal Potential of Root Zone Technology from Domestic Wastewater', *Universal Journal of Environmental Research and Technology* www.environmentaljournal.org, 3(3), pp. 401–406.
- Manios, T., Stentiford, E. I. and Millner, P. (2003) 'Removal of heavy metals from a metaliferous water solution by *Typha latifolia* plants and sewage sludge compost', *Chemosphere*. Pergamon, 53(5), pp. 487–494. doi: 10.1016/S0045-6535(03)00537-X.
- Marimon, Z. A., Xuan, Z. and Chang, N.-B. (2013) 'System dynamics modeling with sensitivity analysis for floating treatment wetlands in a stormwater wet pond', *Ecological Modelling*. Elsevier, 267, pp. 66–79. doi: 10.1016/J.ECOLMODEL.2013.07.017.
- Masclaux-Daubresse, C., Daniel-Vedele, F., Dechorgnat, J., Chardon, F., Gaufichon, L. and Suzuki, A. (2010) 'Review: part of a special issue on plant nutrition Nitrogen uptake, assimilation and remobilization in plants: challenges for sustainable and productive agriculture', *Annals of Botany*, 105, pp. 1141–1157. doi: 10.1093/aob/mcq028.
- Masi, F., Rizzo, A. and Regelsberger, M. (2017) 'The role of constructed wetlands in a new circular economy, resource oriented, and ecosystem services paradigm', *Journal of Environmental Management*. doi: 10.1016/j.jenvman.2017.11.086.
- Matsuzaki, S. S., Usio, N., Takamura, N. and Washitani, I. (2009) 'Contrasting impacts of invasive engineers on freshwater ecosystems: an experiment and meta-analysis', *Oecologia*. Springer-Verlag, 158(4), pp. 673–686. doi: 10.1007/s00442-008-1180-1.
- McAndrew, B. and Ahn, C. (2017) 'Developing an ecosystem model of a floating wetland for water quality improvement on a stormwater pond', *Journal of Environmental Management*. Academic Press, 202, pp. 198–207. doi: 10.1016/J.JENVMAN.2017.07.035.

- McAndrew, B., Ahn, C. and Spooner, J. (2016) 'Nitrogen and Sediment Capture of a Floating Treatment Wetland on an Urban Stormwater Retention Pond—The Case of the Rain Project', *Sustainability*, 8(10), p. 972. doi: 10.3390/su8100972.
- Meals, D. W., Dressing, S. A. and Davenport, T. E. (2010) 'Lag Time in Water Quality Response to Best Management Practices: A Review', *Journal of Environment Quality*, 39(1), p. 85. doi: 10.2134/jeq2009.0108.
- Meng, F., Huang, J., Liu, H. and Chi, J. (2015) 'Remedial effects of *Potamogeton crispus* L. on PAH-contaminated sediments', *Environmental Science and Pollution Research*, 22, pp. 7547–7556.
- Meuleman, A. F. M., Beekman, H. and Verhoeven, J. T. A. (2002) 'Nutrient retention and nutrient-use efficiency in phragmites australis stands after wastewater application', *WETLANDS*, 22(4), pp. 712–721.
- Miretzky, P., Saralegui, A. and Cirelli, A. F. (2004) 'Aquatic macrophytes potential for the simultaneous removal of heavy metals (Buenos Aires, Argentina)', *Chemosphere*, 57(8), pp. 997–1005. doi: 10.1016/j.chemosphere.2004.07.024.
- Mkandawire, M. and Dudel, E. G. (2007) 'Are Lemna spp. Effective Phytoremediation Agents?', *Bioremediation, Biodiversity and Bioavailability*.
- Mkandawire, M. and Dudel, E. G. (2005) 'Accumulation of arsenic in Lemna gibba L. (duckweed) in tailing waters of two abandoned uranium mining sites in Saxony, Germany', *Science of The Total Environment*, 336(1), pp. 81–89. doi: 10.1016/j.scitotenv.2004.06.002.
- Mkandawire, M., Lyubun, Y. V., Kosterin, P. V. and Dudel, E. G. (2004) 'Toxicity of arsenic species to Lemna gibba L. and the influence of phosphate on arsenic bioavailability', *Environmental Toxicology*. Wiley Subscription Services, Inc., A Wiley Company, 19(1), pp. 26–34. doi: 10.1002/tox.10148.
- Mkandawire, M., Taubert, B. and Dudel, E. G. (2004) 'Capacity of Lemna gibba L. (Duckweed) for Uranium and Arsenic Phytoremediation in Mine Tailing Waters', *International Journal of Phytoremediation*, 6(4), pp. 347–362. doi: 10.1080/16226510490888884.
- Molisani, M. M. and Lacerda, L. D. (2006) 'Mercury contents in aquatic macrophytes from two reservoirs in the Paraíba Do Sul: guandú river system, Se Brazil', *Brazilian Journal of Biology*, 66(1A), pp. 101–107.
- Moore, M. T., Locke, M. A. and Kröger, R. (2016) 'Using aquatic vegetation to remediate nitrate, ammonium, and soluble reactive phosphorus in simulated runoff', *Chemosphere*, 160, pp. 149–154. doi: 10.1016/j.chemosphere.2016.06.071.
- Van de Moortel, A. M. K., Du Laing, G., De Pauw, N. and Tack, F. M. G. (2011) 'Distribution and Mobilization of Pollutants in the Sediment of a Constructed Floating Wetland Used for Treatment of Combined Sewer Overflow Events', *Water Environment Research*, 83(5), pp. 427–439. doi: 10.2175/106143010X12851009156169.
- Morency, D. and Belnick, T. (1987) 'CONTROL OF INTERNAL PHOSPHORUS LOADING IN TWO SHALLOW LAKES BY ALUM AND AQUATIC PLANT HARVESTING', *Lake and Reservoir Management*, 3(1), pp. 31–37.
- Mwendera, C., De Jager, C., Longwe, H., Hongoro, C., Phiri, K. and Mutero, C. M. (2017)

'Development of a framework to improve the utilisation of malaria research for policy development in Malawi', *Health Research Policy and Systems*, 15. doi: 10.1186/s12961-017-0264-y.

Nesshöver, C., Assmuth, T., Irvine, K. N., Rusch, G. M., Waylen, K. A., Delbaere, B., Haase, D., Jones-Walters, L., Keune, H., Kovacs, E., Krauze, K., Külvik, M., Rey, F., van Dijk, J., Vistad, O. I., Wilkinson, M. E. and Wittmer, H. (2017) 'The science, policy and practice of nature-based solutions: An interdisciplinary perspective', *Science of The Total Environment*, pp. 1215–1227. doi: 10.1016/j.scitotenv.2016.11.106.

Newete, S. W. and Byrne, M. J. (2016) 'The capacity of aquatic macrophytes for phytoremediation and their disposal with specific reference to water hyacinth.', *Environmental science and pollution research international*, 23(11), pp. 10630–43. doi: 10.1007/s11356-016-6329-6.

Nichols, P., Lucke, T., Drapper, D. and Walker, C. (2016) 'Performance Evaluation of a Floating Treatment Wetland in an Urban Catchment', *Water*. Multidisciplinary Digital Publishing Institute, 8(6), p. 244. doi: 10.3390/w8060244.

Ning, D., Huang, Y., Pan, R., Wang, F. and Wang, H. (2014) 'Effect of eco-remediation using planted floating bed system on nutrients and heavy metals in urban river water and sediment: A field study in China', *Science of The Total Environment*, 485, pp. 596–603. doi: 10.1016/j.scitotenv.2014.03.103.

Nivala, J., Hoos, M. B., Cross, C., Wallace, S. and Parkin, G. (2007) 'Treatment of landfill leachate using an aerated, horizontal subsurface-flow constructed wetland', *Science of The Total Environment*, 380(1), pp. 19–27. doi: 10.1016/j.scitotenv.2006.12.030.

Ohe, T., Watanabe, T. and Wakabayashi, K. (2004) 'Mutagens in surface waters: a review', *Mutation Research/Reviews in Mutation Research*, 567(2), pp. 109–149. doi: 10.1016/j.mrrev.2004.08.003.

Olguín, E. J. and Sá Nchez-Galva, G. (2012) 'Heavy metal removal in phytofiltration and phycoremediation: the need to differentiate between bioadsorption and bioaccumulation', *New BIOTECHNOLOGY*, 30(1), pp. 3–8. doi: 10.1016/j.nbt.2012.05.020.

Olguín, E. J., Sanchez-Galvan, G., Melo, F. J., Hernandez, V. J. and Gonzalez-Portela, R. E. (2017) 'Long-term assessment at field scale of Floating Treatment Wetlands for improvement of water quality and provision of ecosystem services in a eutrophic urban pond', *Science of The Total Environment*. Elsevier B.V. doi: 10.1016/j.scitotenv.2017.01.072.

Oren Benaroya, R., Tzin, V., Tel-Or, E. and Zamski, E. (2004) 'Lead accumulation in the aquatic fern *Azolla filiculoides*', *Plant Physiology and Biochemistry*, 42(7), pp. 639–645. doi: 10.1016/j.plaphy.2004.03.010.

Ormerod, S. J., Dobson, M., Hildrew, A. G. and Townsend, C. R. (2010) 'Multiple stressors in freshwater ecosystems', *Freshwater Biology*, 55(SUPPL. 1), pp. 1–4. doi: 10.1111/j.1365-2427.2009.02395.x.

Osmolovskaya, N. and Kurilenko, V. (2005) 'Macrophytes in phytoremediation of heavy metal contaminated water and sediments in urban inland ponds', *Geophysical Research Abstracts*, 7.

Padmavathiamma, P. K. and Li, L. Y. (2007) 'Phytoremediation Technology: Hyper-accumulation Metals in Plants', *Water, Air, and Soil Pollution*, 184(1–4), pp. 105–126. doi: 10.1007/s11270-007-9401-5.

Paisio, C. E., Fernandez, M., Gonzalez, P. S., Talano, M. A., Medina, M. I. and Agostini, E. (2018)

'Simultaneous phytoremediation of chromium and phenol by *Lemna minuta* Kunth: a promising biotechnological tool', *International Journal of Environmental Science and Technology*. Springer Berlin Heidelberg, 15(1), pp. 37–48. doi: 10.1007/s13762-017-1368-1.

Panich-pat, T. (2005) 'Electron Microscopic Studies on Localization of Lead in Organs of *Typha angustifolia* Grown on Contaminated Soil', *ScienceAsia*, 31, pp. 49–53.

Parzych, A., Sobisz, Z. and Cymer, M. (2016a) 'Preliminary research of heavy metals content in aquatic plants taken from surface water (Northern Poland)', *Desalination and Water Treatment*. Taylor & Francis, 57(3), pp. 1451–1461. doi: 10.1080/19443994.2014.1002275.

Parzych, A., Sobisz, Z. and Cymer, M. (2016b) 'Preliminary research of heavy metals content in aquatic plants taken from surface water (Northern Poland)', *Desalination and Water Treatment*, 57(3), pp. 1451–1461. doi: 10.1080/19443994.2014.1002275.

Patel, S. (2012) 'Threats, management and envisaged utilizations of aquatic weed *Eichhornia crassipes*: an overview'. doi: 10.1007/s11157-012-9289-4.

Patiño Gómez, J. M. and Lara-Borrero, J. A. (2012) 'Investment, operation and maintenance costs (2012) for natural wastewater treatment systems in small communities in Colombia', *European Water*, 40, pp. 19–30.

Pavlineri, N., Skoulikidis, N. T. and Tsihrintzis, V. A. (2017) 'Constructed Floating Wetlands: A review of research, design, operation and management aspects, and data meta-analysis', *Chemical Engineering Journal*, 308, pp. 1120–1132. doi: 10.1016/j.cej.2016.09.140.

Perk, M. van der (2006) *Soil and Water Contamination from molecular to catchment scale*. ast. London: Taylor & Francis Group.

Peterson, S. A., Smith, W. L. and Malueg, K. W. (1974) 'Full-Scale Harvest of Aquatic Plants: Nutrient Removal from a Eutrophic Lake', *Source Journal (Water Pollution Control Federation)*, 46(4), pp. 697–707.

Phetsombat, S., Kruatrachue, M., Pokethitiyook, P. and Upatham, S. (2006) 'Toxicity and bioaccumulation of cadmium and lead in *Salvinia cucullata*', *Journal of Environmental Biology Enterprises*, 27(4), pp. 645–652.

Picard, C. R., Fraser, L. H. and Steer, D. (2005) 'The interacting effects of temperature and plant community type on nutrient removal in wetland microcosms', *Bioresource Technology*, 96(9), pp. 1039–1047. doi: 10.1016/j.biortech.2004.09.007.

Polechońska, L. and Samecka-Cymerman, A. (2016) 'Bioaccumulation of macro-and trace elements by European frogbit (*Hydrocharis morsus-ranae* L.) in relation to environmental pollution', *Environmental Science and Pollution*, pp. 3469–3480. doi: 10.1007/s11356-015-5550-z.

Polomski, R. F., Taylor, M. D., Bielenberg, D. G., Bridges, W. C., Klaine, S. J. and Whitwell, T. (2009) 'Nitrogen and Phosphorus Remediation by Three Floating Aquatic Macrophytes in Greenhouse-Based Laboratory-Scale Subsurface Constructed Wetlands', *Water, Air, and Soil Pollution*. Springer Netherlands, 197(1–4), pp. 223–232. doi: 10.1007/s11270-008-9805-x.

Portielje, R. and Van der Molen, D. T. (1999) 'Relationships between eutrophication variables: from nutrient loading to transparency', in *Shallow Lakes '98*. Dordrecht: Springer Netherlands, pp. 375–387. doi: 10.1007/978-94-017-2986-4_42.

- Pratas, J., Paulo, C., Favas, P. J. C. and Venkatachalam, P. (2014) 'Potential of aquatic plants for phytofiltration of uranium-contaminated waters in laboratory conditions', *Ecological Engineering*, 69, pp. 170–176. doi: 10.1016/j.ecoleng.2014.03.046.
- Qian, J.-H., Zayed, A., Zhu, Y.-L., Yu, M. and Terry, N. (1999) 'Phytoaccumulation of Trace Elements by Wetland Plants: III. Uptake and Accumulation of Ten Trace Elements by Twelve Plant Species', *Journal of Environment Quality*. American Society of Agronomy, Crop Science Society of America, and Soil Science Society of America, 28(5), p. 1448. doi: 10.2134/jeq1999.00472425002800050009x.
- Quilliam, R. S., van Niekerk, M. A., Chadwick, D. R., Cross, P., Hanley, N., Jones, D. L., Vinten, A. J. A., Willby, N. and Oliver, D. M. (2015) 'Can macrophyte harvesting from eutrophic water close the loop on nutrient loss from agricultural land?', *Journal of Environmental Management*. Elsevier Ltd, 152, pp. 210–217. doi: 10.1016/j.jenvman.2015.01.046.
- Rahman, M. A. and Hasegawa, H. (2011) 'Aquatic arsenic: Phytoremediation using floating macrophytes', *Chemosphere*. Pergamon, 83(5), pp. 633–646. doi: 10.1016/J.CHEMOSPHERE.2011.02.045.
- Rai, P. K. (2009) *Heavy Metal Phytoremediation from Aquatic Ecosystems with Special Reference to Macrophytes*, *Critical Reviews in Environmental Science and Technology*. doi: 10.1080/10643380801910058.
- Rai, U. N., Tripathi, R. D., Vajpayee, P., Pandey, N., Ali, M. B. and Gupta, D. K. (2003) 'Cadmium accumulation and its phytotoxicity in potamogeton pectinatus L. (Potamogetonaceae)', *Bulletin of Environmental Contamination and Toxicology*, 70(3), pp. 566–575. doi: 10.1007/s00128-003-0023-3.
- Reinhold, D., Vishwanathan, S., Park, J. J., Oh, D. and Michael Saunders, F. (2010) 'Assessment of plant-driven removal of emerging organic pollutants by duckweed', *Chemosphere*, 80(7), pp. 687–692. doi: 10.1016/j.chemosphere.2010.05.045.
- Reinoso, R., Torres, L. A. and Bécares, E. (2008) 'Efficiency of natural systems for removal of bacteria and pathogenic parasites from wastewater', *Science of The Total Environment*, 395(2), pp. 80–86. doi: 10.1016/j.scitotenv.2008.02.039.
- Rezania, S., Taib, S. M., Fadhil, M., Din, M., Dahalan, F. A. and Kamyab, H. (2016) 'Comprehensive review on phytotechnology: Heavy metals removal by diverse aquatic plants species from wastewater', *Journal of Hazardous Materials*, 318, pp. 587–599. doi: 10.1016/j.jhazmat.2016.07.053.
- Rockström, J., Falkenmark, M., Allan, T., Folke, C., Gordon, L., Jägerskog, A., Kummu, M., Lannerstad, M., Meybeck, M., Molden, D., Postel, S., Savenije, H. H. G., Svedin, U., Turton, A. and Varis, O. (2014) 'The unfolding water drama in the Anthropocene: Towards a resilience-based perspective on water for global sustainability', *Ecohydrology*, 7(5), pp. 1249–1261. doi: 10.1002/eco.1562.
- Rodríguez, M., Brisson, J., Rueda, G. and Rodríguez, M. S. (2012) 'Water Quality Improvement of a Reservoir Invaded by an Exotic Macrophyte', *Invasive Plant Science and Management*, 5(2), pp. 290–299. doi: 10.1614/IPSM-D-11-00023.1.
- Rosenkranz, T., Kisser, J., Wenzel, W. W. and Puschenreiter, M. (2017) 'Waste or substrate for metal hyperaccumulating plants — The potential of phytomining on waste incineration bottom ash', *Science of The Total Environment*, 575, pp. 910–918. doi: 10.1016/j.scitotenv.2016.09.144.
- Saeed, T., Paul, B., Afrin, R., Al-Muyeed, A. and Sun, G. (2016) 'Floating constructed wetland for the treatment of polluted river water: A pilot scale study on seasonal variation and shock load', *Chemical*

Engineering Journal, 287, pp. 62–73. doi: 10.1016/j.cej.2015.10.118.

Sarma, H. (2011) 'Metal hyperaccumulation in plants: a review focusing on phytoremediation technology', *Journal of Environmental Science and Technology*. 4: 118-138.

Sayer, C., Davidson, T. and Jones, J. (2010) 'Seasonal dynamics of macrophytes and phytoplankton in shallow lakes: a eutrophication-driven pathway from plants to plankton?', *Freshwater Biology*. Blackwell Publishing Ltd, 55(3), pp. 500–513. doi: 10.1111/j.1365-2427.2009.02365.x.

Scholes, L. N. L., Shutes, R. B. E., Revitt, D. M., Purchase, D. and Forshaw, M. (1999) 'The removal of urban pollutants by constructed wetlands during wet weather', *Water Science and Technology*. No longer published by Elsevier, 40(3), pp. 333–340. doi: 10.1016/S0273-1223(99)00467-9.

Schultz, R. and Dibble, E. (2012) 'Effects of invasive macrophytes on freshwater fish and macroinvertebrate communities: the role of invasive plant traits', *Hydrobiologia*. Springer Netherlands, 684(1), pp. 1–14. doi: 10.1007/s10750-011-0978-8.

Sheoran, V., Sheoran, A. S. and Poonia, P. (2009) 'Phytomining: A review', *Minerals Engineering*, 22(12), pp. 1007–1019. doi: 10.1016/j.mineng.2009.04.001.

Singh, N., Pandey, G., Rai, U., Tripathi, R., Singh, H. and Gupta, D. (2005) 'Metal Accumulation and Ecophysiological Effects of Distillery Effluent on *Potamogeton pectinatus* L', *Bulletin of Environmental Contamination and Toxicology*, 74, pp. 857–863. doi: 10.1007/s00128-005-0660-9.

Sivaci, E., Sivaci, A. and Sokman, M. (2004) 'Biosorption of cadmium by *Myriophyllum spicatum* and *Myriophyllum triphyllum* orchard', *Chemosphere*, 56, pp. 1043–1048.

Smart, R. M., Dick, G. O. and Doyle, A. R. D. (1998) 'Techniques for Establishing Native Aquatic Plants', *J. Aquat. Plant Manage*, 36(36), pp. 44–49.

Song, H.-L., Nakano, K., Taniguchi, T., Nomura, M. and Nishimura, O. (2009) 'Estrogen removal from treated municipal effluent in small-scale constructed wetland with different depth', *Bioresource Technology*, 100(12), pp. 2945–2951. doi: 10.1016/j.biortech.2009.01.045.

Song, U. and Park, H. (2017) 'Importance of biomass management acts and policies after phytoremediation', *Journal of Ecology and Environment*, 41(13), pp. 1–6. doi: 10.1186/s41610-017-0033-4.

Souza, F. A., Dzedzic, M., Cubas, S. A. and Maranhão, L. T. (2013) 'Restoration of polluted waters by phytoremediation using *Myriophyllum aquaticum* (Vell.) Verdc., Haloragaceae', *Journal of Environmental Management*, 120, pp. 5–9. doi: 10.1016/j.jenvman.2013.01.029.

Srivastava, S., Shrivastava, M., Suprasanna, P. and D'Souza, S. F. (2011) *Phytofiltration of arsenic from simulated contaminated water using *Hydrilla verticillata* in field conditions*, *Ecological Engineering*. doi: 10.1016/j.ecoleng.2011.06.012.

Stottmeister, U., Wießner, A., Kusch, P., Kappelmeyer, U., Kästner, M., Bederski, O., Müller, R. A. and Moormann, H. (2003) 'Effects of plants and microorganisms in constructed wetlands for wastewater treatment', *Biotechnology Advances*, 22(1), pp. 93–117. doi: 10.1016/j.biotechadv.2003.08.010.

Sun, L., Liu, Y. and Jin, H. (2009) 'Nitrogen removal from polluted river by enhanced floating bed grown canna', *Ecological Engineering*, 35(1), pp. 135–140. doi: 10.1016/j.ecoleng.2008.09.016.

- Sundaravadivel, M. and Vigneswaran, S. (2001) 'Constructed Wetlands for Wastewater Treatment', *Critical Review in Environmental Science and Technology*, 31(4). doi: 10.3390/w2030530.
- Sunita Sharma, Bikram Singh, V. K. M. (2015) 'Phytoremediation: role of terrestrial plants and aquatic macrophytes in the remediation of radionuclides and heavy metal contaminated soil and water', *Environ Sci Pollut Res*, 22, pp. 946–962.
- Taghi, M., Khosravi, M. and Rakhshae, R. (2005) 'Biosorption of Pb, Cd, Cu and Zn from the wastewater by treated *Azolla filiculoides* with H₂O₂/MgCl₂', *International Journal of Environmental Science & Technology*, 1(4), pp. 265–271.
- Tanaka, T. S. T., Irbis, C., Kumagai, H., Wang, P., Li, K. and Inamura, T. (2017) 'Effect of *Phragmites japonicus* harvest frequency and timing on dry matter yield and nutritive value', *Journal of Environmental Management*, 187, pp. 436–443. doi: 10.1016/j.jenvman.2016.11.008.
- Tanner, C. C. (1996) 'Plants for constructed wetland treatment systems - A comparison of the growth and nutrient uptake of eight emergent species', *Ecological Engineering*, 7(1), pp. 59–83. doi: 10.1016/0925-8574(95)00066-6.
- Tanner, C. C. and Headley, T. R. (2011a) 'Components of floating emergent macrophyte treatment wetlands influencing removal of stormwater pollutants', *Ecological Engineering*, 37(3), pp. 474–486. doi: 10.1016/j.ecoleng.2010.12.012.
- Tanner, C. C. and Headley, T. R. (2011b) 'Components of floating emergent macrophyte treatment wetlands influencing removal of stormwater pollutants', *Ecological Engineering*. Elsevier B.V., 37(3), pp. 474–486. doi: 10.1016/j.ecoleng.2010.12.012.
- Tanner, C. C., Sukias, J. P. S., Park, J., Yates, C. and Headley, T. (2011) *Floating treatment wetlands: a new tool for nutrient management in lakes and waterways, Adding to the knowledge base for the nutrient manager*.
- The Great Britain Non-native Species Secretariat (2015) *The Great Britain Invasive Non-Native Species Strategy*. York. Available at: www.nationalarchives.gov.uk/doc/open-government-licence/version/3/ (Accessed: 3 March 2018).
- Thijs, S., Sillen, W., Rineau, F., Weyens, N. and Vangronsveld, J. (2016) 'Towards an Enhanced Understanding of Plant-Microbiome Interactions to Improve Phytoremediation: Engineering the Metaorganism.', *Frontiers in microbiology*. p. 341. doi: 10.3389/fmicb.2016.00341.
- Tilley, E., Ulrich, L., Luthi, C., Reymond, P. and Zurbrugg, C. (2014) *Compendium of Sanitation Systems and Technologies. 2nd Revised Edition*. Available at: http://www.sswm.info/category/implementation-tools/wastewater-treatment/hardware/semi-centralised-wastewater-treatments/h#reference_book7934 (Accessed: 12 December 2016).
- Tilman, D., Balzer, C., Hill, J. and Befort, B. L. (2011) 'Global food demand and the sustainable intensification of agriculture.', *Proceedings of the National Academy of Sciences of the United States of America*. National Academy of Sciences, 108(50), pp. 20260–4. doi: 10.1073/pnas.1116437108.
- Tran, V. S., Ngo, H. H., Guo, W., Zhang, J., Liang, S., Ton-That, C. and Zhang, X. (2015) 'Typical low cost biosorbents for adsorptive removal of specific organic pollutants from water', *Bioresource Technology*. pp. 353–363. doi: 10.1016/J.BIORTECH.2015.02.003.
- Tront, J. M. and Saunders, F. M. (2006) 'Role of plant activity and contaminant speciation in aquatic

- plant assimilation of 2,4,5-trichlorophenol', *Chemosphere*, 64(3), pp. 400–407. doi: 10.1016/j.chemosphere.2005.12.025.
- Truu, M., Juhanson, J. and Truu, J. (2009) 'Microbial biomass, activity and community composition in constructed wetlands', *Science of The Total Environment*, 407(13), pp. 3958–3971. doi: 10.1016/j.scitotenv.2008.11.036.
- Turgut, C. (2005) 'Uptake and modeling of pesticides by roots and shoots of parrotfeather (*Myriophyllum aquaticum*)', *Environmental Science and Pollution Research*, 12(6), pp. 342–346. doi: 10.1065/espr2005.05.256.
- Türker, O. C., Türe, C., Böcük, H. and Yakar, A. (2016) 'Phyto-management of boron mine effluent using native macrophytes in mono-culture and poly-culture constructed wetlands', *Ecological Engineering*. Elsevier B.V., 94, pp. 65–74. doi: 10.1016/j.ecoleng.2016.05.043.
- Tyler, H. L., Moore, M. T. and Locke, M. A. (2012) 'Potential for Phosphate Mitigation from Agricultural Runoff by Three Aquatic Macrophytes', *Water Air Soil Pollution*, 223, pp. 4557–4564.
- Ulén, B., Bechmann, M., Fölster, J., Jarvie, H. P. and Tunney, H. (2007) 'Agriculture as a phosphorus source for eutrophication in the north-west European countries, Norway, Sweden, United Kingdom and Ireland: a review', *Soil Use and Management*. Blackwell Publishing Ltd, 23, pp. 5–15. doi: 10.1111/j.1475-2743.2007.00115.x.
- Valipour, A. and Ahn, Y.-H. (2016) 'Constructed wetlands as sustainable ecotechnologies in decentralization practices: a review', *Environmental Science and Pollution Research*. Heidelberg, 23(1), pp. 180–197. doi: 10.1007/s11356-015-5713-y.
- Verkleij, J. A. C., Golan-Goldhirsh, A., Antosiewicz, D. M., Schwitzguébel, J.-P. and Schröder, P. (2009) 'Dualities in plant tolerance to pollutants and their uptake and translocation to the upper plant parts', *Environmental and Experimental Botany*, 67(1), pp. 10–22. doi: 10.1016/j.envexpbot.2009.05.009.
- Vymazal, J. (2007) 'Removal of nutrients in various types of constructed wetlands', *Science of the Total Environment*, 380(1–3), pp. 48–65. doi: 10.1016/j.scitotenv.2006.09.014.
- Vymazal, J. (2009) 'The use constructed wetlands with horizontal sub-surface flow for various types of wastewater', *Ecological Engineering*, 35(1), pp. 1–17. doi: 10.1016/j.ecoleng.2008.08.016.
- Vymazal, J. (2011) 'Constructed Wetlands for Wastewater Treatment: Five Decades of Experience [†]', *Environmental Science & Technology*. American Chemical Society, 45(1), pp. 61–69. doi: 10.1021/es101403q.
- Vymazal, J. (2016) *Concentration is not enough to evaluate accumulation of heavy metals and nutrients in plants*, *Science of The Total Environment*. doi: 10.1016/j.scitotenv.2015.12.011.
- Vymazal, J. and Kröpfelová, L. (2009) 'Removal of organics in constructed wetlands with horizontal sub-surface flow: A review of the field experience', *Science of The Total Environment*, 407(13), pp. 3911–3922. doi: 10.1016/j.scitotenv.2008.08.032.
- Wand, H., Vacca, G., Kuschik, P., Krü Ger, M. and Kä Stner, M. (2006) 'Removal of bacteria by filtration in planted and non-planted sand columns'. doi: 10.1016/j.watres.2006.08.024.
- Wang, C.-Y. and Sample, D. J. (2014a) 'Assessment of the nutrient removal effectiveness of floating

treatment wetlands applied to urban retention ponds', *Journal of Environmental Management*, 137, pp. 23–35. doi: 10.1016/j.jenvman.2014.02.008.

Wang, C.-Y. and Sample, D. J. (2014b) 'Assessment of the nutrient removal effectiveness of floating treatment wetlands applied to urban retention ponds', *Journal of Environmental Management*, 137, pp. 23–35. doi: 10.1016/j.jenvman.2014.02.008.

Wang, C.-Y., Sample, D. J. and Bell, C. (2014) 'Vegetation effects on floating treatment wetland nutrient removal and harvesting strategies in urban stormwater ponds', *Science of The Total Environment*. Elsevier, 499, pp. 384–393. doi: 10.1016/J.SCITOTENV.2014.08.063.

Wang, C. Y., Sample, D. J., Day, S. D. and Grizzard, T. J. (2015) 'Floating treatment wetland nutrient removal through vegetation harvest and observations from a field study', *Ecological Engineering*. Elsevier B.V., 78, pp. 15–26. doi: 10.1016/j.ecoleng.2014.05.018.

Wang, G. X., Zhang, L. M., Chua, H., Li, X. D., Xia, M. F. and Pu, P. M. (2009) 'A mosaic community of macrophytes for the ecological remediation of eutrophic shallow lakes', *Ecological Engineering*, 35(4), pp. 582–590. doi: 10.1016/j.ecoleng.2008.06.006.

Wang, T., Weissman, J., Ramesh, G., Varadarajan, R. and Benemann, J. (1996) 'Parameters for removal of toxic heavy metals by water Milfoil (*Myriophyllum spicatum*)', *Bulletin of Environmental Contamination and Toxicology*, 57, pp. 779–786.

Windham, L., Weis, J. . and Weis, P. (2003) 'Uptake and distribution of metals in two dominant salt marsh macrophytes, *Spartina alterniflora* (cordgrass) and *Phragmites australis* (common reed)', *Estuarine, Coastal and Shelf Science*, 56(1), pp. 63–72. doi: 10.1016/S0272-7714(02)00121-X.

Xia, H. and Ma, X. (2006) 'Phytoremediation of ethion by water hyacinth (*Eichhornia crassipes*) from water', *Bioresource Technology*, 97(8), pp. 1050–1054. doi: 10.1016/j.biortech.2005.04.039.

Xian, Q., Hu, L., Chen, H., Chang, Z. and Zou, H. (2010) 'Removal of nutrients and veterinary antibiotics from swine wastewater by a constructed macrophyte floating bed system', *Journal of Environmental Management*, 91(12), pp. 2657–2661. doi: 10.1016/j.jenvman.2010.07.036.

Xiao, J., Chu, S., Tian, G., Thring, R. W. and Cui, L. (2016) 'An Eco-tank system containing microbes and different aquatic plant species for the bioremediation of N,N-dimethylformamide polluted river waters', *Journal of Hazardous Materials*, 320, pp. 564–570. doi: 10.1016/j.jhazmat.2016.07.037.

Xing, W., Wu, H., Hao, B. and Liu, G. (2013) 'Metal accumulation by submerged macrophytes in eutrophic lakes at the watershed scale', *Environmental Science and Pollution Research*, 20(10), pp. 6999–7008. doi: 10.1007/s11356-013-1854-z.

Xu, Z. H., Yin, X. A. and Yang, Z. F. (2014) 'An optimisation approach for shallow lake restoration through macrophyte management', *Hydrol. Earth Syst. Sci*, 18, pp. 2167–2176. doi: 10.5194/hess-18-2167-2014.

Yamazaki, K., Tsuruta, H. and Inui, H. (2015) 'Different uptake pathways between hydrophilic and hydrophobic compounds in lateral roots of *Cucurbita pepo*', *Journal of Pesticide Science*. Pesticide Science Society of Japan, 40(3), pp. 99–105. doi: 10.1584/jpestics.D14-081.

Yan, S.-H., Song, W. and Guo, J.-Y. (2017) 'Critical Reviews in Biotechnology Advances in management and utilization of invasive water hyacinth (*Eichhornia crassipes*) in aquatic ecosystems – a review Advances in management and utilization of invasive water hyacinth (*Eichhornia crassipes*)

- in aquatic', *Critical Reviews in Biotechnology Crit Rev Biotechnol*, 37(372), pp. 218–228. doi: 10.3109/07388551.2015.1132406.
- Yang, B., Lan, C. Y., Yang, C. S., Liao, W. B., Chang, H. and Shu, W. S. (2006) 'Long-term efficiency and stability of wetlands for treating wastewater of a lead/zinc mine and the concurrent ecosystem development', *Environmental Pollution*, 143, pp. 499–512. doi: 10.1016/j.envpol.2005.11.045.
- Yang, Z., Zheng, S., Chen, J. and Sun, M. (2008) 'Purification of nitrate-rich agricultural runoff by a hydroponic system', *Bioresource Technology*. Elsevier, 99(17), pp. 8049–8053. doi: 10.1016/J.BIORTECH.2008.03.040.
- Ye, Z., Baker, A., Wong, M. and Willis, A. (1997) 'Zinc, lead and cadmium tolerance, uptake and accumulation by *Typha latifolia*', *New Phytologist*. Wiley/Blackwell (10.1111), 136(3), pp. 469–480. doi: 10.1046/j.1469-8137.1997.00759.x.
- Yeh, N., Yeh, P. and Chang, Y.-H. (2015) 'Artificial floating islands for environmental improvement', *Renewable and Sustainable Energy Reviews*, 47, pp. 616–622. doi: 10.1016/j.rser.2015.03.090.
- Zarate, F. M., Schulwitz, S. E., Stevens, K. J. and Venables, B. J. (2012) 'Bioconcentration of triclosan, methyl-triclosan, and triclocarban in the plants and sediments of a constructed wetland', *Chemosphere*, 88, pp. 323–329. doi: 10.1016/j.chemosphere.2012.03.005.
- Zayed, A., Gowthaman, S. and Terry, N. (1998) 'Phytoaccumulation of Trace Elements by Wetland Plants: I. Duckweed', *Journal of Environment Quality*. American Society of Agronomy, Crop Science Society of America, and Soil Science Society of America, 27(3), p. 715. doi: 10.2134/jeq1998.00472425002700030032x.
- Zayed, A., Pilon-Smits, E., de Souza, M., Lin, Z. and Terry, N. (2000) 'Remediation of selenium polluted soils and waters by phytovolatilization', in Terry, N. and Banuelos, G. (eds) *Phytoremediation of contaminated soil and water*. 1st edn. Boca Raton.
- Zhang, C.-B., Liu, W.-L., Pan, X.-C., Guan, M., Liu, S.-Y., Ge, Y. and Chang, J. (2014) 'Comparison of effects of plant and biofilm bacterial community parameters on removal performances of pollutants in floating island systems', *Ecological Engineering*, 73, pp. 58–63. doi: 10.1016/j.ecoleng.2014.09.023.
- Zhang, D., Gersberg, R. M., Ng, W. J. and Tan, S. K. (2014) 'Removal of pharmaceuticals and personal care products in aquatic plant-based systems: A review', *Environmental Pollution*, 184, pp. 620–639. doi: 10.1016/j.envpol.2013.09.009.
- Zhang, D. Q., Hua, T., Gersberg, R. M., Zhu, J., Ng, W. J. and Tan, S. K. (2012) 'Fate of diclofenac in wetland mesocosms planted with *Scirpus validus*', *Ecological Engineering*, 49, pp. 59–64. doi: 10.1016/j.ecoleng.2012.08.018.
- Zhang, D. Q., Hua, T., Gersberg, R. M., Zhu, J., Ng, W. J. and Tan, S. K. (2013a) 'Carbamazepine and naproxen: Fate in wetland mesocosms planted with *Scirpus validus*', *Chemosphere*, 91(1), pp. 14–21. doi: 10.1016/j.chemosphere.2012.11.018.
- Zhang, D. Q., Hua, T., Gersberg, R. M., Zhu, J., Ng, W. J. and Tan, S. K. (2013b) *Fate of caffeine in mesocosms wetland planted with Scirpus validus*, *Chemosphere*. doi: 10.1016/j.chemosphere.2012.09.059.
- Zhang, D. Q., Tan, S. K., Gersberg, R. M., Sadreddini, S., Zhu, J. and Tuan, N. A. (2011) 'Removal of

pharmaceutical compounds in tropical constructed wetlands', *Ecological Engineering*, 37(3), pp. 460–464. doi: 10.1016/j.ecoleng.2010.11.002.

Zhang, X., Hu, Y., Liu, Y. and Chen, B. (2011) 'Arsenic uptake, accumulation and phytofiltration by duckweed (*Spirodela polyrhiza* L.)', *Journal of Environmental Sciences*, 23(4), pp. 601–606. doi: 10.1016/S1001-0742(10)60454-8.

Zhang, Z., Rengel, Z. and Meney, K. (2007) 'Nutrient removal from simulated wastewater using *Canna indica* and *Schoenoplectus validus* in mono- and mixed-culture in wetland microcosms', *Water, Air, and Soil Pollution*, 183(1–4), pp. 95–105. doi: 10.1007/s11270-007-9359-3.

Zhang, Z., Rengel, Z., Meney, K., Zhang, Z., Rengel, Z. and Meney, K. (2007) 'Nutrient Removal from Simulated Wastewater Using *Canna indica* and *Schoenoplectus validus* in Mono-and Mixed-Culture in Wetland Microcosms', *Water Air Soil Pollut*, 183, pp. 95–105. doi: 10.1007/s11270-007-9359-3.

Zhao, F., Xi, S., Yang, X., Yang, W., Li, J., Gu, B. and He, Z. (2011) 'Purifying eutrophic river waters with integrated floating island systems', *Ecological Engineering*, 40, pp. 53–60. doi: 10.1016/j.ecoleng.2011.12.012.

Zhao, F., Yang, W., Zeng, Z., Li, H., Yang, X., He, Z., Gu, B., Rafiq, M. T. and Peng, H. (2012) 'Nutrient removal efficiency and biomass production of different bioenergy plants in hypereutrophic water', *Biomass and Bioenergy*, 42, pp. 212–218. doi: 10.1016/j.biombioe.2012.04.003.

Zhou, X. and Wang, G. (2010) 'Nutrient concentration variations during *Oenanthe javanica* growth and decay in the ecological floating bed system', *Journal of Environmental Sciences*, 22(11), pp. 1710–1717. doi: 10.1016/S1001-0742(09)60310-7.

Zhu, J., Hu, W., Hu, L., Deng, J., Li, Q. and Gao, F. (2012) 'Variation in the Efficiency of Nutrient Removal in a Pilot-Scale Natural Wetland', *Wetlands*, 32, pp. 11–319. doi: 10.1007/s13157-011-0261-9.

Zhu, L., Li, Z. and Ketola, T. (2011) 'Biomass accumulations and nutrient uptake of plants cultivated on artificial floating beds in China's rural area', *Ecological Engineering*, 37(10), pp. 1460–1466. doi: 10.1016/j.ecoleng.2011.03.010.

Zhu, Y. L., Zayed, A. M., Qian, J. H., de Souza, M. and Terry, N. (1999) 'Phytoaccumulation of trace elements by wetland plants: II. Water hyacinth', *Journal of Environmental Quality*, 28(1), pp. 339–344. doi: 10.2134/jeq1999.00472425002800010042x.

Zimmerman, J. B., Mihelcic, J. R. and Smith, and J. (2008) 'Global Stressors on Water Quality and Quantity', *Environmental Science & Technology*. American Chemical Society, 42(12), pp. 4247–4254. doi: 10.1021/es0871457.

Table 12.1: Key pollutants impacting the aquatic environment, organised by pollutant category, type and providing examples of the pollutants, their sources and impacts

Pollutant category	Pollutant Type	Example pollutant	Sources	Potential impacts
Organic	Persistent organic pollutants (POPs)/Xenobiotics	Dioxins, organochlorides, Polycyclic aromatic hydrocarbons (PAH), Polychlorinated biphenyls	Industry Agriculture	Toxicity Endocrine disrupting effects
	Pesticides	Glyphosate Hexachlorocyclohexane Fenhexamid Deltamethrin	Agriculture Aquaculture	Toxicity Endocrine disrupting effects
	Pharmaceutical and personal care products (PPCPs)	Antibiotics Hormones Pain relief medication	Domestic Agriculture Aquaculture	Endocrine disrupting effects Antibiotic resistance Destabilising microbial communities
	Algal toxins	Microcystin-LR	Cyanobacterial algal blooms	Acute/chronic toxicity
Inorganic	Nutrients	Nitrogen (N) Phosphorus (P) Potassium (K)	Agriculture Aquaculture Septic tank inputs	Nutrient enrichment/eutrophication
	Metalloid elements	Iron (Fe) Aluminium (Al) Lead (Pb) Nickel (Ni) Cadmium (Cd)	Agriculture Industry (mining and combustion of fossil fuels) Al mobilisation	Toxicity Endocrine disrupting effects

		Copper (Cu) Uranium (U)	through acid rain	
Microbial	Pathogens and parasites	<i>E.coli</i> O157 <i>Cryptosporidium parvum</i>	Agriculture Aquaculture Domestic	Human illness (intestinal infection)



Figure 12.1: Photo examples of floating, submerged and emergent macrophyte life forms. From left to right: *Persicaria amphibia* (floating), *Ceratophyllum demersum* (Submerged) and *Sparganium erectum* (emergent)

Table 12.2: Phytoremediation mechanisms, adapted from Dhir (2013) and Rezania et al. (2016).

Mechanism	Medium	Contaminant category	Description	Accumulation Part	Example genera
Rhizofiltration/phytofiltration	Water	Organics/inorganics /heavy metals	Extraction from contaminated water by adsorption/absorption	Shoots/roots	<i>Lemna, Hydrocharis, Eichhornia</i>
Phytoextraction/phytoaccumulation	Soil/water	Inorganics/heavy metals	Uptake by roots and translocation to upper parts	Shoots	<i>Juncus, Schoenoplectus</i>
Phytostabilisation	Soil/sediment	Inorganics/heavy metals	Rendering contaminants immobile within soil matrix due to plant root action	Reduction in rhizosphere	<i>Chenopodium</i>
Phytovolatilization	Soil/sediment/ water (less common)	Organics	Conversion of contaminants to volatile form	Atmospheric release	<i>Phragmites</i>
Phytodegradation	Soil/sediment/ Water	Organics/inorganics /microbiological	Degradation in Rhizosphere through microbial degradation or by metabolism within plant	Degradation in rhizosphere/pollutant degraded in plant to less harmful metabolite	<i>Typha, Phragmites, Myriophyllum</i>

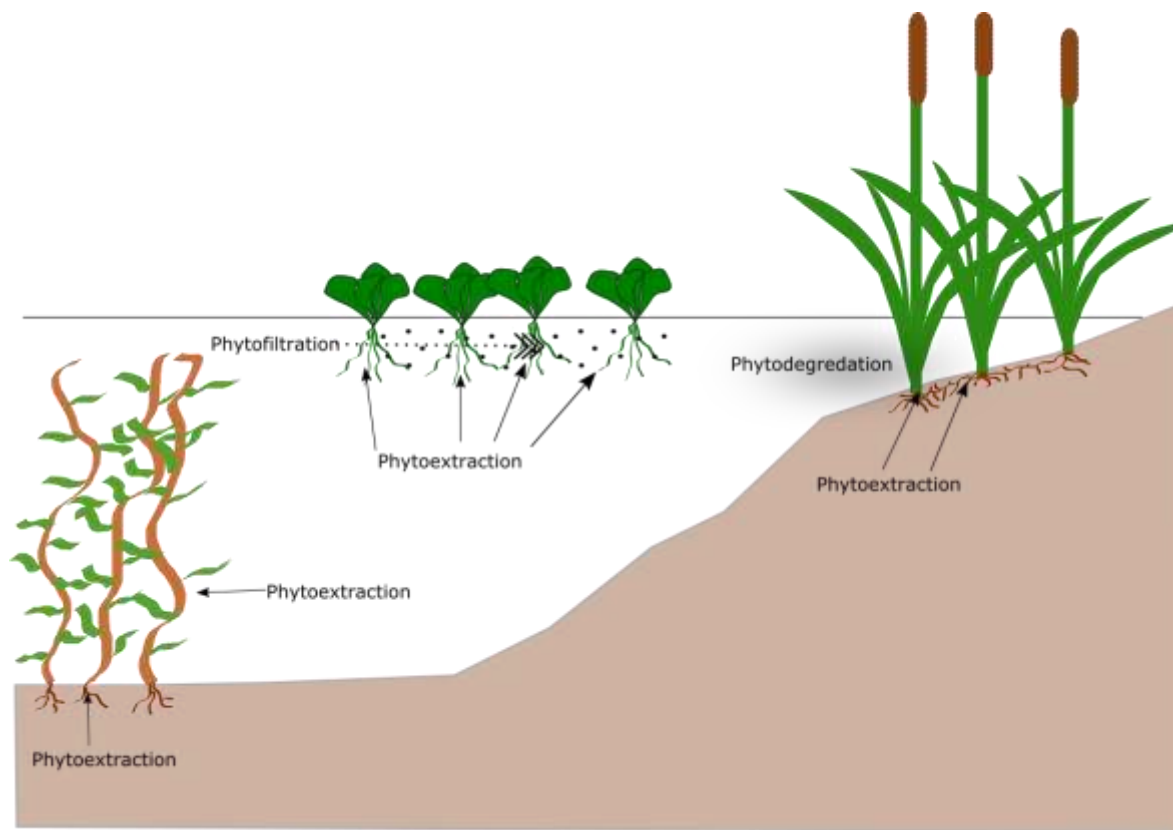


Figure 12.2 Phytoremediation mechanisms used to degrade/remove waterborne pollutants, by growth form.

Table 12.3: Removal efficiencies (%) of macrophyte species investigated in this review of nutrients phyto remediation

Species	Life Form	Removal Efficiency (%)					Macrophyte Deployment	Experiment	Reference
		Total Nitrogen	Nitrate (NO ₃)	Ammonia (NH ₃)	Ammonium (NH ₄)	Total Phosphorus			
<i>Canna sp.</i>	Emergent	50			100		FTW	Mesocosm	Sun et al (2009)

Species	Life Form	Removal Efficiency (%)					Macrophyte Deployment	Experiment	Reference	
		Total Nitrogen	Nitrate (NO ₃)	Ammonia (NH ₃)	Ammonium (NH ₄)	Total Phosphorus				
<i>Cyperus sp.</i>	Emergent				42			FTW	Mesocosm	Ayaz & Saygin (1996)
					33			FTW	Mesocosm	Ayaz & Saygin (1996)
		72			75			Constructed wetland	Constructed wetland	Kyambadde et al. (2004)
<i>Polygonum hydropiperoides</i>	Emergent	57			63	54.09		FTW	Microcosm	Kansiime et al. (2005)
		74				81		Direct planting	Mesocosm	Lang Martins et al. (2010)
<i>Echinodorus cordifolius</i>	Emergent		45		49.9	10.85		Direct planting	Mesocosm	Moore et al. (2016)
<i>Ipomoea aquatica</i>	Emergent	76						FTW	Mesocosm	Karnchanawong (1995)
		36-46				36-47		FTW	Mesocosms	Li et al. (2010)
<i>Juncus effusus</i>	Emergent	61.94			48	62		FTW	Mesocosm	Li et al. (2010)
		48		50		63		Constructed wetland	Constructed wetland	Coleman et al. (2001)
<i>Leersia oryzoides</i>	Emergent					51		Direct planting	Mesocosm	Tyler et al. (2012)
<i>Limnocharis flava</i>	Emergent			92				Constructed wetland	Constructed wetland	Kamarudzaman & Ismail (2011)
<i>Lolium multiflorum</i>	Emergent	81				90		FTW	Mesocosm	Xian et al. (2010)
<i>Miscanthidium violaceum</i>	Emergent	57			47	41		Constructed wetland	Constructed wetland	Kyambadde et al. (2004)
<i>Oenanthe javanica</i>	Emergent	91		97		76		FTW	Mesocosm	Zhou & Wang (2010)
<i>Panicum hemitomon</i>	Emergent		60		54	28		Direct planting	Mesocosm	Moore et al. (2016)

Species	Life Form	Removal Efficiency (%)					Macrophyte Deployment	Experiment	Reference	
		Total Nitrogen	Nitrate (NO ₃)	Ammonia (NH ₃)	Ammonium (NH ₄)	Total Phosphorus				Phosphate
<i>Phragmites</i>	Emergent				98			FTW	Mesocosm	Kintu Sekiranda & Kiwanuka, (1997)
<i>Saururus cernuus</i>	Emergent		35		-3	-13		Direct planting	Mesocosm	Moore et al. (2016)
<i>Scirpus atrovirens</i>	Emergent			91			82	Constructed wetland	Constructed wetland	Kamarudzaman & Ismail (2011)
<i>Scirpus validus</i>	Emergent	25		25		48		Constructed wetland	Constructed wetland	Coleman et al. (2001)
<i>Sparganium americanum</i>	Emergent					14		Direct planting	Mesocosm	Tyler et al. (2012)
<i>Thalia dealbata</i>	Emergent		46		31	4		Direct planting	Mesocosm	Moore et al. (2016)
<i>Typha angustifolia</i>	Emergent	57				23		FTW	Mesocosm pots	Keizer-Vlek et al. (2014)
<i>Typha latifolia</i>	Emergent	62		62		81		Constructed wetland	Constructed wetland	Coleman et al. (2001)
						53		Direct planting	Mesocosm	Tyler et al. (2012)
<i>Vetiveria zizanoides</i>	Emergent	49	32		17	12		Direct planting	Mesocosm	Moore et al. (2016)
				50		21		FTW	Mesocosm	Boonsong & Chansiri (2008)
<i>Eichhornia crassipes</i>	Floating		61-83					Direct planting	Mesocosm	Ayyasamy et al. (2009)
			92	81		67		Direct planting	Mesocosm	Kutty et al. (2009)
<i>Pistia stratiotes</i>	Floating	50				14-31		Direct planting	Ponds (storm water detention)	Lu et al. (2010)
			31-51					Direct planting	Mesocosm	Ayyasamy et al. (2009)

Species	Life Form	Removal Efficiency (%)						Macrophyte Deployment	Experiment	Reference
		Total Nitrogen	Nitrate (NO ₃)	Ammonia (NH ₃)	Ammonium (NH ₄)	Total Phosphorus	Phosphate			
<i>Salvinia molesta</i>	Floating		18-36					Direct planting	Mesocosm	Ayyasamy et al. (2009)
<i>Lemna gibba</i>	Floating	97	100	82		99		Direct planting	mesocosm-wastwater	Körner & Vermaat (1998)
							64	Sewage water system	Sewage water system	El-Kheir et al. (2007)
<i>Ceratophyllum demersum</i>	Submerged	42			65	73		Direct planting	Mesocosms	Dai et al. (2012)
<i>Myriophyllum aquaticum</i>	Submerged	88	45		35	94		Direct planting	Mesocosm	Souza et al. (2013)
						7		Direct planting	Mesocosm	Moore et al. (2016)

Species	Life Form	Removal Efficiency (%)						Macrophyte Deployment	Experiment	Reference
		Total Nitrogen	Nitrate (NO ₃)	Ammonia (NH ₃)	Ammonium (NH ₄)	Total Phosphorus	Phosphate			
<i>Canna sp.</i>	Emergent	50		100				FTW	Mesocosm	Sun et al (2009)
<i>Cyperus sp.</i>	Emergent			42				FTW	Mesocosm	Ayaz & Saygin(1996)
		72		33				FTW	Mesocosm	Ayaz & Saygin(1996)
				75				Constructed wetland	Constructed wetland	Kyambadde et al.(2004)
		57		63		54.09		FTW	Microcosm	Kansiime et al.(2005)
<i>Polygonum hydropiperoides</i>	Emergent	74				81		Direct planting	Mesocosm	Lang Martins et al.(2010)
<i>Echinodorus cordifolius</i>	Emergent		45		49.9	10.85		Direct planting	Mesocosm	Moore et al.(2016)

Species	Life Form	Removal Efficiency (%)					Macrophyte Deployment	Experiment	Reference	
		Total Nitrogen	Nitrate (NO ₃)	Ammonia (NH ₃)	Ammonium (NH ₄)	Total Phosphorus				
<i>Ipomoea aquatica</i>	Emergent	76					FTW	Mesocosm	Karnchanawong(1995)	
		36-46				36-47	FTW	Mesocosms	Li et al. (2010)	
		61.94		48		62	FTW	Mesocosm	Li et al. (2010)	
<i>Juncus effusus</i>	Emergent	48		50		63	Constructed wetland	Constructed wetland	Coleman et al.(2001)	
<i>Leersia oryzoides</i>	Emergent					51	Direct planting	Mesocosm	Tyler et al.(2012)	
<i>Limnocharis flava</i>	Emergent			92			96	Constructed wetland	Constructed wetland	Kamarudzaman & Ismail(2011)
<i>Lolium multiflorum</i>	Emergent	81				90	FTW	Mesocosm	Xian et al.(2010)	
<i>Miscanthidium violaceum</i>	Emergent	57		47		41	Constructed wetland	Constructed wetland	Kyambadde et al.(2004)	
<i>Oenanthe javanica</i>	Emergent	91		97		76	FTW	Mesocosm	Zhou & Wang (2010)	
<i>Panicum hemitomon</i>	Emergent		60		54	28	Direct planting	Mesocosm	Moore et al.(2016)	
<i>Phragmites</i>	Emergent			98			FTW	Mesocosm	Kintu Sekiranda & Kiwanuka, (1997)	
<i>Saururus cernuus</i>	Emergent		35		-3	-13	Direct planting	Mesocosm	Moore et al.(2016)	
<i>Scirpus atrovirens</i>	Emergent			91			82	Constructed wetland	Constructed wetland	Kamarudzaman & Ismail(2011)
<i>Scirpus validus</i>	Emergent	25		25		48	Constructed wetland	Constructed wetland	Coleman et al.(2001)	
<i>Sparganium americanum</i>	Emergent					14	Direct planting	Mesocosm	Tyler et al. (2012)	

Species	Life Form	Removal Efficiency (%)					Macrophyte Deployment	Experiment	Reference	
		Total Nitrogen	Nitrate (NO ₃)	Ammonia (NH ₃)	Ammonium (NH ₄)	Total Phosphorus				Phosphate
<i>Thalia dealbata</i>	Emergent		46		31	4		Direct planting	Mesocosm	Moore et al.(2016)
<i>Typha angustifolia</i>	Emergent	57				23		FTW	Mesocosm pots	Keizer-Vlek et al.(2014)
<i>Typha latifolia</i>	Emergent	62		62		81		Constructed wetland	Constructed wetland	Coleman et al.(2001)
						53		Direct planting	Mesocosm	Tyler et al. (2012)
<i>Vetiveria zizanoides</i>	Emergent	49	32	50	17	12		Direct planting	Mesocosm	Moore et al.(2016)
						21		FTW	Mesocosm	Boonsong & Chansiri (2008)
<i>Eichhornia crassipes</i>	Floating		61-83					Direct planting	Mesocosm	Ayyasamy et al.(2009)
			92	81		67		Direct planting	Mesocosm	Kutty et al. (2009)
<i>Pistia stratiotes</i>	Floating	50				14-31		Direct planting	Ponds (storm water detention)	Lu et al.(2010)
			31-51					Direct planting	Mesocosm	Ayyasamy et al.(2009)
<i>Salvinia molesta</i>	Floating		18-36					Direct planting	Mesocosm	Ayyasamy et al.(2009)
<i>Lemna gibba</i>	Floating	97				99		Direct planting	mesocosm-wastwater	Körner & Vermaat (1998)
			100	82			64	Sewage water system	Sewage water system	El-Kheir et al. (2007)
<i>Ceratophyllum demersum</i>	Submerged	42			65	73		Direct planting	Mesocosms	Dai et al. (2012)
<i>Myriophyllum</i>	Submerge	88				94		Direct planting	Mesocosm	Souzaet al.(2013)

Species	Life Form	Removal Efficiency (%)					Macrophyte Deployment	Experiment	Reference	
		Total Nitrogen	Nitrate (NO ₃)	Ammonia (NH ₃)	Ammonium (NH ₄)	Total Phosphorus				
<i>aquaticum</i>	d		45		35	7		Direct planting	Mesocosm	Moore et al.(2016)

Table 12.4: Key macrophyte metal accumulators reported in the literature

Species	Life Form	Metals	Reference
<i>Ceratophyllum submersum</i>	Submerged	Ni	Kara (2010)
<i>Ceratophyllum demersum</i>	Submerged	Cr, Pb	Osmolovskaya and Kurilenko (2005)
<i>Potamogeton natans</i>	Submerged	U	Pratas et al. (2014)
<i>Myriophyllum spicatum</i>	Submerged	Co,Cu, Mn, Pb, Zn	Wang et al. (1996);Sivaci et al., (2004); Lesage et al. (2008)
<i>Potamogeton pectinatus</i>	Submerged	Cd, Cu, Mn, Pb, Zn	Rai et al. (2003);Singh et al. (2005)
<i>Hydrilla verticillata</i>	Submerged	As, Cu	Srivastava et al. (2011)
<i>Lemnocharis flava</i>	Emergent	Cu, Fe, Hg, Pb, Zn	Anninget al.(2013)
<i>Glyceria maxima</i>	Emergent	Cu, Zn	Parzych et al.(2016)
<i>Typha latifolia</i>	Emergent	As, Cu, Ni, Zn	Ye et al. (1997);Ha et al.(2009);Manios et al.(2003); Qian et al. (1999)
<i>Typha angustifolia</i>	Emergent	Pb	Panich-pat (2005)
<i>Elodea densa</i>	Emergent	Hg	Molisani and Lacerda(2006)
<i>Phalaris arundinacea</i>	Emergent	Fe, Mn, Ni	Parzych et al. (2016)
<i>Phargmites australis</i>	Emergent	As, Hg	Windham et al.(2003); Afrous et al. (2011)
<i>Scirpus maritimus</i>	Emergent	As,	Afrous et al. (2011)
<i>Spartina alterniflora</i>	Emergent	As,	Carbonell et al. (1998)
<i>Spartina patens</i>	Emergent	Cd	Zayed et al. (2000)
<i>Azolla filiculoides</i>	Floating	Cd, Cr, Ni, Pb, Zn	Oren Benaroya et al. (2004);Aroraet al.(2006):Taghi et al.(2005); Zayed et al.(1998)
<i>Azolla caroliniana</i>	Floating	As, Cr, Cu, Hg	Rahman and Hasegawa(2011); Bennicelli et al. (2004)
<i>Pista stratiotes</i>	Floating	Cr,Cu, Hg	Miretzky et al.(2004);Molisani et al. (2006); Maine et al.(2004)
<i>Salvinia cucullata</i>	Floating	Cd, Pb	Phetsombat et al. (2006)
<i>Salvinia natans</i>	Floating	Cr, Zn	Dhir et al.(2008)
<i>Spirodela polyrhiza</i>	Floating	As	Zhang et al. (2011)
<i>Eichhornia crassipes</i>	Floating	Cd, Cr, Cu, Hg, Ni, Zn	Zhu et al. (1999):Hu et al. (2007); Molisani et al. (2006); Low et al. (1994)
<i>Lemna gibba</i>	Floating	As, Cd, Ni	Mkandawire and Dudel(2005); Mkandawire et al. (2004); Mkandawire et al.(2004)

Table 12.5: Removal efficiencies of macrophyte species investigated in phytoremediation studies of organic pollutants

Organic Pollutant	Species	Life Form	Target pollutant	Experimental situation	Removal (%)	Reference
Pesticides	<i>Cannaxgeneralise</i>	Emergent	Isoxaben, oryzalin	Mesocosm	n/a	Fernandez et al. (1999)
	<i>Pontaaderia cordata</i>	Emergent	Isoxaben, oryzalin	Mesocosm	n/a	Fernandez et al. (1999)
	<i>Iris L.x'Charjoys Jan'</i>	Emergent	Isoxaben, oryzalin	Mesocosm	n/a	Fernandez et al. (1999)
	<i>Eichhornia crassipes</i>	Floating	Ethion	Mesocosm	81	Xia & Ma(2006)
	<i>Juncus effusus</i>	Emergent	Atrazine, Lambda-cyhalothrin	Mesocosm	n/a	Bouldin et al.(2006)
	<i>Ludwigia peploides</i>	Emergent	Atrazine, Lambda-cyhalothrin	Mesocosm	n/a	Bouldin et al.(2006)
	<i>Lemna minor</i>	Floating	2,4,5-trichlorophenol	Mesocosm	95	Tront & Saunders(2006)
			Isoproturon, Glyphosate	Mesocosm	25, 8	Dosnon-Olette et al. (2011)
	<i>Spirodela oligorrhiza</i>	Floating	DDT (OP,PP-DDT)	Mesocosm	66, 50	Gao et al. (2000)
	<i>Elodea canadensis</i>	Submerged	DDT (OP,PP-DDT)	Mesocosm	31, 48	Gao et al. (2000)
	<i>Mariophyllum aquaticum</i>	Submerged	DDT (OP,PP-DDT) Trifluralin, cycloxdim, Atrazine, Terbutryn	Mesocosm	76, 82	Gao et al. (2000)
		<i>Elodea canadensis</i>	Submerged	DDT (OP,PP-DDT)	Mesocosm	89
POP	<i>Lemna gibba</i>	Floating	Phenol	Mesocosm	90	Barber et al. (1995)
	<i>Lemna minuta</i>	Floating	Phenol	Mesocosm	100	Paisio et al. (2018)
	<i>Potamogeton crispus</i>	Submerged	Phenol	Mesocosm	70-100	Hafez et al.(1998)
	PAHs (phenanthrene and pyrene)		Mesocosm (sediment pots)	18-34 , 14.-24	Meng et al (2015)	

included)						
PPCP	<i>Phragmites australis</i>	Emergent	Estrone, 17 beta-estradiol, 17 alpha-ethynylestradiol	Constructed wetland	68-84	Song et al.(2009)
	<i>Scirpus validus</i>	Emergent	Diclofenac	Mesocosm	1-7%	Zhang et al. (2012)
			Naproxen, Carbamazepine,	Constructed wetland	97-99,53-60	Zhang et al. (2013a)
			Caffeine	Mesocosm	>99.7	Zhang et al.(2013b)
	<i>Typha angustifolia</i>	Emergent	Carbamazepine, Naproxen, Diclofenac, Ibuprofen	Constructed wetland	27, 91, 55,80	Zhang et al. (2011)
	<i>Pontederia cordata</i>	Emergent	Troclosan, methyl-triclosan & Triclocarbon	Constructed wetland	n/a	Zarate et al.(2012)
	<i>Sagittaria graminea</i>	Emergent	Troclosan, methyl-triclosan & Triclocarbon	Constructed wetland	n/a	Zarate et al.(2012)
	<i>Typha latifolia</i>	Emergent	Troclosan, methyl-triclosan & Triclocarbon	Constructed wetland	n/a	Zarate et al. (2012)

Note

1. n/a refers to studies where the removal efficiencies are not reported

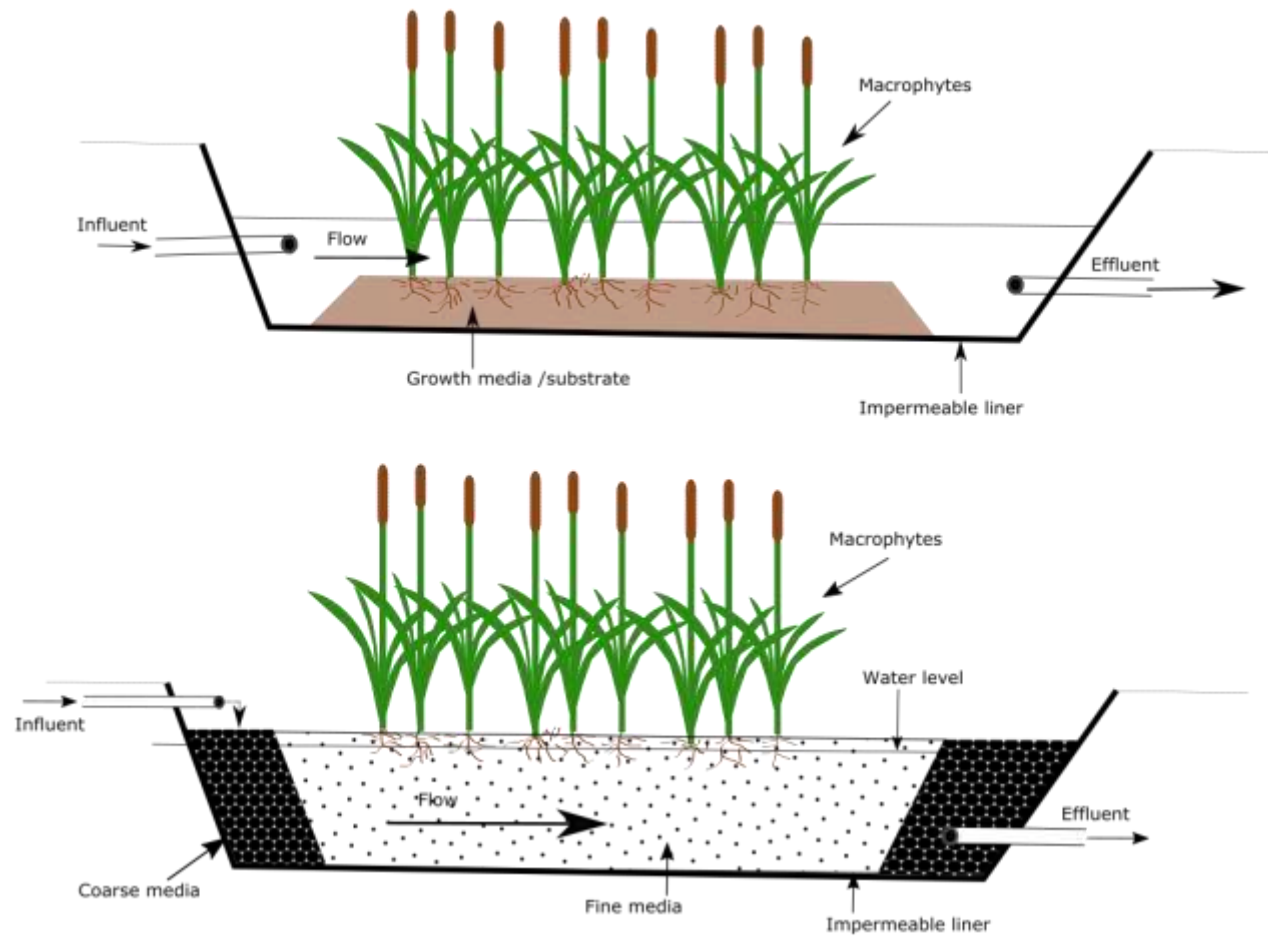


Figure 12.3: Top: Key elements of a free water surface flow wetlands (FWSF) constructed wetland. Bottom: Key elements of a or sub-surface flow (SSF) constructed wetland.

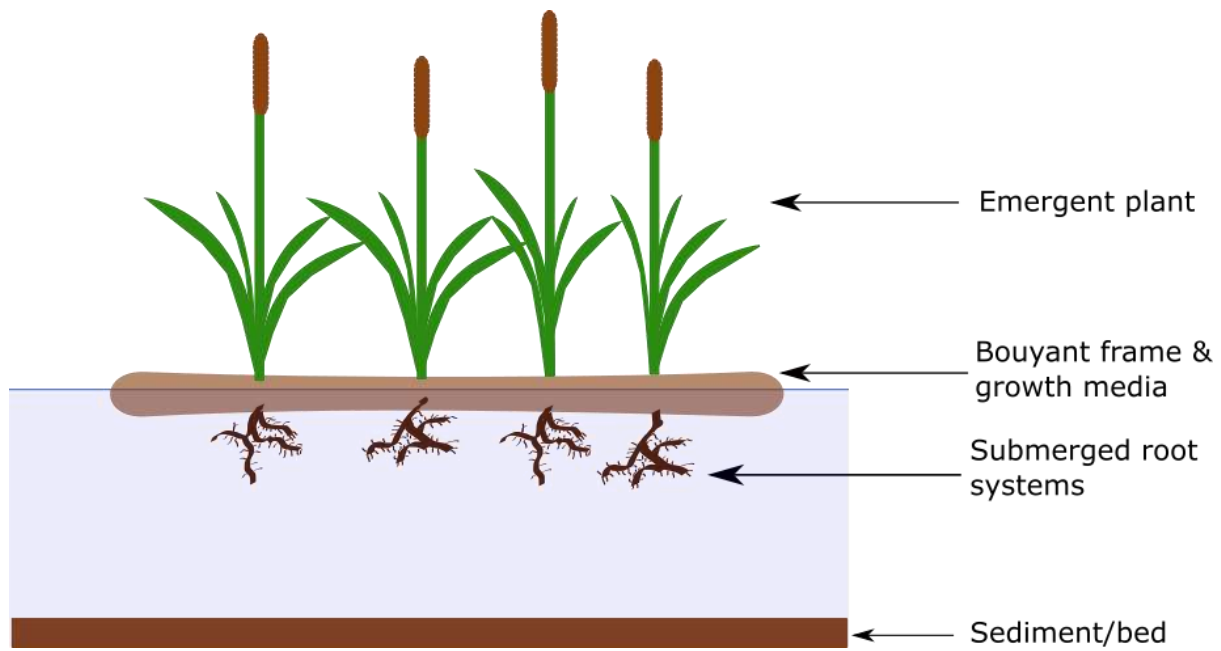


Figure 12.4: Schematic view of a FTW.

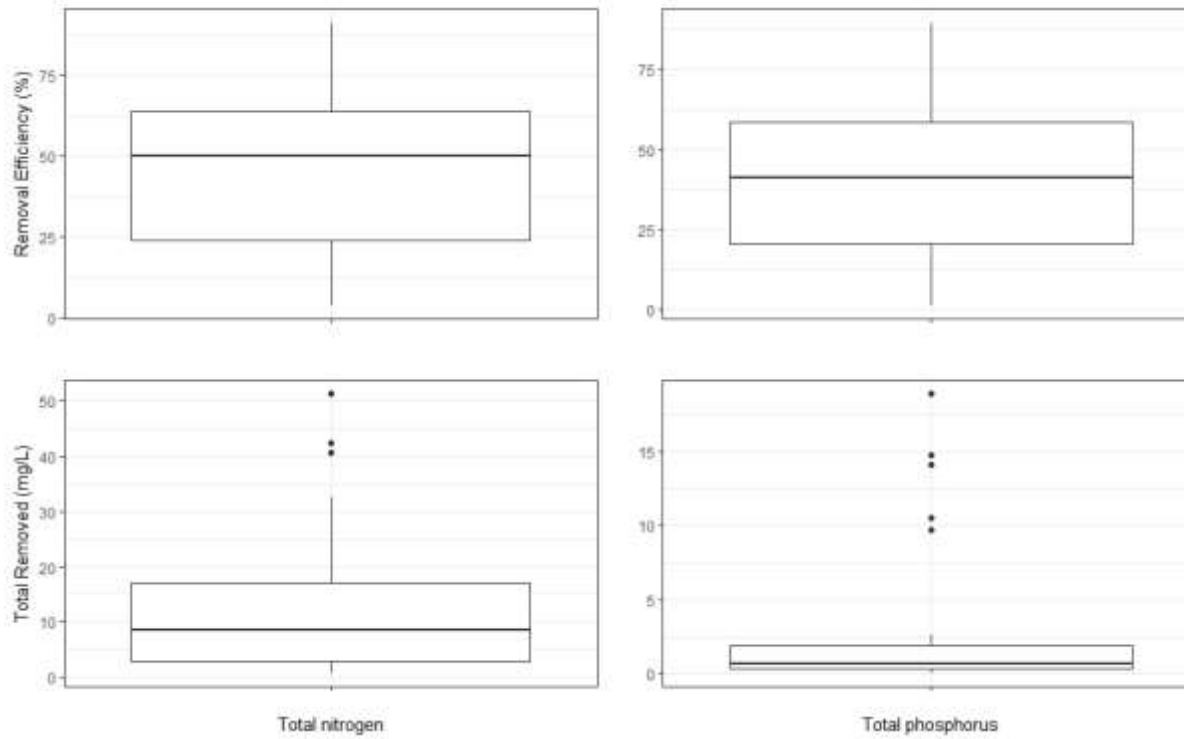


Figure 12.5: Boxplots of removal efficiencies (%) and total removed (mg/l) of Total Nitrogen (TN) ($n=44$) and Total phosphorus (TP) ($n=28$), raw data taken from literature reviewed by Pavlineri et al. (2017).

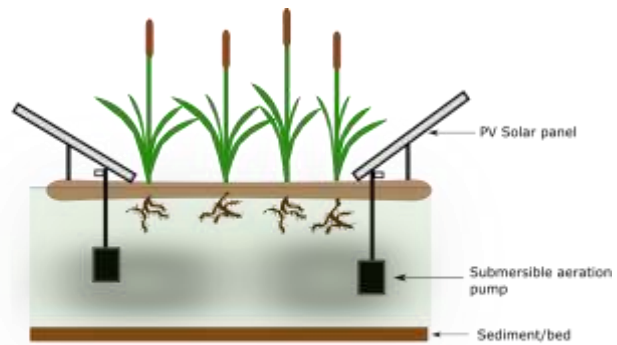
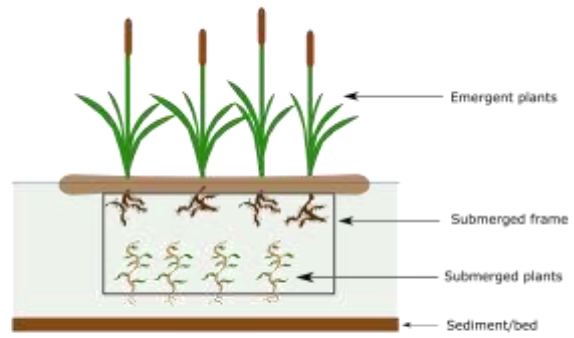


Figure 12.6: Top, a schematic representation of a hybrid FTW including submerged vegetation.

Bottom schematic representation of a FTW incorporating solar technology to power an aeration device.

Table 12.6: Plant allocations of pollutants in selected emergent, floating and submerged macrophytes

Species	Growth form	Plant allocation of pollutant		Reference
		Above-ground	Below-ground	
<i>Cyperus riparia</i>	Emergent	Cd, Ni, Zn		Ladislav et al. (2013)
<i>Cyperus esculentus</i>	Emergent	Cd, Cr, Cu, Fe, Mn, Ni	Pb	Chandra & Yadav(2011)
<i>Glyceria maxima</i>	Emergent	Cu, Fe, Mn, Ni, Zn		Parzych et al. (2016)
<i>Juncus effusus</i>	Emergent	Cd, Ni	Zn	Ladislav et al. (2013)
<i>Phalaris arundinacea</i>	Emergent	Cu, Fe, Mn, Ni, Zn		Parzych et al. (2016)
<i>Phragmites australis</i>	Emergent	Cu, Fe, Ni, Zn	Mn	Parzych et al. (2016)
		Cr, Cu, Mn, Ni, Zn		Duman et al.(2007)
<i>Phragmites australis</i>	Emergent	Cd, Cu, Zn	Cr, Fe, Mn, Pb	Chandra & Yadav(2011)
<i>Schoenoplectus lacustris</i>	Emergent	Cu, Ni, Pb, Zn		Duman et al.(2007)
<i>Typha angustifolia</i>	Emergent	Cd, Cr, Cu, Fe, Mn, Ni, Pb	Zn	Chandra & Yadav(2011)
<i>Typha domingensis</i>	Emergent	Ca, Cu, Fe, P, Zn	N	Eid et al. (2012)
<i>Typha latifolia</i>	Emergent	Cu, Fe, Ni, Zn	Mn	Parzych et al. (2016)
<i>Eichhornia crassipes</i>	Floating		N, P	Polomski et al. (2009)
<i>Pistia stratiotes</i>	Floating		N, P	Polomski et al. (2009)
	Floating	Al, Cd, Co, Cr, Cu, Fe, K, Mg, Na	Ca	Lu et al.(2011)
<i>Micranthemum umbrosum</i>	Submerged	Cd	As	Islam et al.(2013)
<i>Myriophyllum aquaticum</i>	Submerged		N, P	Polomski et al. (2009)

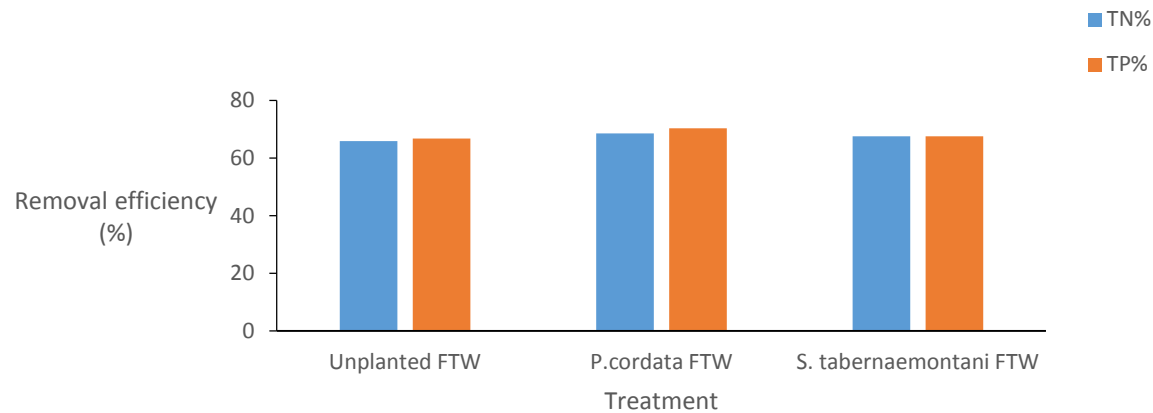


Figure 12.7: Removal efficiencies of TN and TP for an unplanted FTW, a *P. cordata* planted FTW and an *S. tabernaemontani* FTW. Raw data taken from Wang & Sample (2014)

Table 12.7: Summary of the aquatic phytoremediation research agenda required to deliver efficient, multi-targeted and suitable phytoremediation. Research areas, specific lines of investigation and their priority are highlighted.

Research area	Lines of investigation	High priority (0-2 years)	Medium priority (2-5 years)	Low priority (5-10 years)
Identify new macrophyte accumulators for emerging pollutants	To what extent can macrophytes assimilate and degrade PPCPs and pathogens?			
Plant community-based remediation	Evaluate potential for multi-targeted remediation in plant polyculture incorporating temporal/phonological differences and asses plant competitive effects			

Research area	Lines of investigation	High priority (0-2 years)	Medium priority (2-5 years)	Low priority (5-10 years)
Investigate the role of microbial communities on pollutant uptake/removal	<p>Adopt a 'Metaorganism' approach to address the role of microorganisms and biofilms in phytoremediation by ensuring studies have suitable control treatments, assess spatial and temporal variation in microbial communities in order to fully characterise the bacteria by their functions.</p> <p>Investigate how microbes can maximise the phytoremediation process by different plant associations and FTW growth media.</p> <p>Mas balance studies required, potentially incorporating radiolabelled tracers.</p>			
Assess provision of phytoremediation to provide ecosystem services	Identify and quantify ecosystem services associated with phytoremediation to appreciate the value of method over and above water treatment.			
Develop a system for macrophyte selection	Develop a suitable system for macrophyte selection to provide context-specific phytoremediation as a tool for environmental agencies and stakeholders.			
Identify accumulation zones of pollutants within macrophytes	Further studies into the allocation and translocation of pollutants within plants with temporal assessments of the optimum time to harvest biomass.			
Explore novel ways of deploying macrophytes in the environment for phytoremediation	<p>Explore new ways to deploy macrophytes into aquatic environment, especially by developing aquatic-aquatic attenuation and inducing growth in native flora.</p> <p>Undertake large scale studies of FTWs that assess remediation and FTW surface spatial arrangement.</p> <p>Assess stakeholder usability of novel phytoremediation methods.</p>			
Determine the effect of different growth media on pollutant removal	Assess influence of different FTW growth media e.g. biochar.			
Determine post-remediation re-use streams for resource recovery	Investigate feasible options for resource recovery and identify context-specific post-remediation biomass re-use streams that link with target pollutants e.g. biomass as fertilizers.			

Research area	Lines of investigation	High priority (0-2 years)	Medium priority (2-5 years)	Low priority (5-10 years)
Testing macrophytes for individual accumulators	Continue testing new macrophytes for phytoremediation for inorganic, organic and biological pollutants. Focus on finding non-invasive plants.			

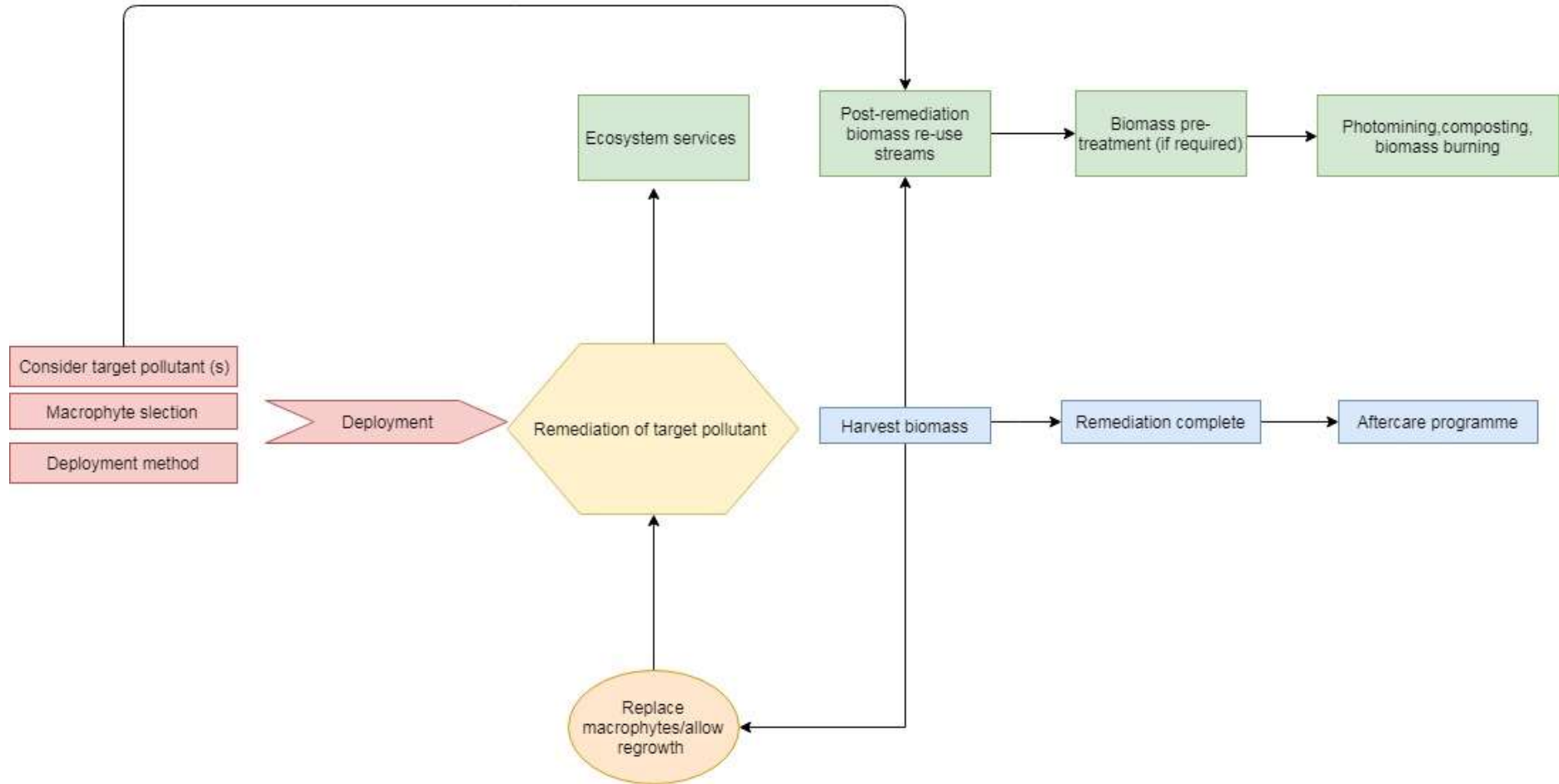


Figure 12.8: Process diagram illustrating the proposed phytoremediation process in its entirety

