

Environmental performance of blue foods

Jessica A. Gephart^{1*+}, Patrik JG Henriksson^{2,3,4+}, Robert W.R. Parker^{5,17+}, Alon Shepon^{6,7,10+}, Kelvin D. Gorospe¹, Kristina Bergman⁸, Gidon Eshel⁹, Christopher D. Golden¹⁰, Benjamin S. Halpern^{12,13}, Sara Hornborg⁸, Malin Jonell^{2,4}, Marc Metian¹⁴, Kathleen Mifflin⁵, Richard Newton¹⁵, Peter Tyedmers⁵, Wenbo Zhang¹⁶, Friederike Ziegler⁸, Max Troell^{2,4}

¹ Dept. of Environmental Science, American University, Washington, DC 20016, USA

² Stockholm Resilience Centre, Kräftriket 2B, 114 19 Stockholm, Sweden

³ WorldFish, Jalan Batu Maung, 11960 Penang, Malaysia

⁴ Beijer Institute of Ecological Economics, The Royal Swedish Academy of Science, Box 50005, 104 05 Stockholm, Sweden

⁵ School for Resource and Environmental Studies, Dalhousie University, Halifax, Nova Scotia, Canada

⁶ Dept. of Environmental Studies, The Porter School of the Environment and Earth Sciences, Tel Aviv University, Israel

⁷ The Steinhardt Museum of Natural History, Tel Aviv University, Israel

⁸ Dept. of Agriculture and Food, RISE Research Institutes of Sweden, Göteborg, Sweden

⁹ Dept. of Environmental Science, Bard College, Campus Road, PO Box 5000

¹⁰ Dept. of Nutrition, Harvard T.H. Chan School of Public Health, Boston, MA 02115, USA

¹¹ Dept. of Environmental Health, Harvard T.H. Chan School of Public Health, Boston, MA 02115, USA

¹² National Center for Ecological Analysis and Synthesis, University of California, Santa Barbara, CA 93101 USA

¹³ Bren School of Environmental Science and Management, University of California, Santa Barbara, CA 93106 USA

¹⁴ International Atomic Energy Agency –Environment Laboratories (IAEA-EL), Radioecology Laboratory, 4a Quai Antoine 1er, 98000 Principality of Monaco, Monaco

¹⁵ Institute of Aquaculture, University of Stirling, Stirling, FK9 4LA

¹⁶ College of Fisheries and Life Science, Shanghai Ocean University, Shanghai 201306, China

¹⁷ Aquaculture Stewardship Council, Daalseplein 101, 3511 SX Utrecht, The Netherlands

* Corresponding author, jgephart@american.edu

+ Denotes First Co-Authorship

1 **Fish and other aquatic foods (blue foods) present an opportunity for more sustainable**
2 **diets^{1,2}. Yet comprehensive comparison has been limited due to sparse inclusion of blue**
3 **foods in environmental impact studies^{3,4} relative to the vast diversity of production⁵. We**
4 **provide standardized estimates of greenhouse gas, nitrogen, phosphorus, freshwater, and**
5 **land stressors for species groups covering nearly three quarters of global production. We**
6 **find that across all blue foods, farmed bivalves and seaweeds generate the lowest stressors.**
7 **Capture fisheries predominantly generate greenhouse gas emissions, with small pelagic**
8 **fishes generating lower emissions than all fed aquaculture, but flatfish and crustaceans**
9 **generating the highest. Among farmed finfish and crustaceans, silver and bighead carps**
10 **have the lowest greenhouse gas, nitrogen, and phosphorus emissions, but highest water use,**
11 **while farmed salmon and trout use the least land and water. Finally, we model intervention**
12 **scenarios and find improving feed conversion ratios reduces stressors across all fed groups,**
13 **increasing fish yield reduces land and water use by up to half, and optimizing gears**
14 **reduces capture fishery emissions by more than half for some groups. Collectively, our**
15 **analysis identifies high performing blue foods, highlights opportunities to improve**
16 **environmental performance, advances data-poor environmental assessments, and informs**
17 **sustainable diets.**

18 **MAIN TEXT**

19 The food system is a major driver of environmental change, emitting a quarter of all
20 greenhouse gas emissions, occupying half of all ice-free land, and responsible for three quarters
21 of global consumptive water use and eutrophication^{3,6}. Yet, it still fails to meet global nutrition
22 needs⁷, with 820 million people lacking sufficient food⁸ and with one in three people globally
23 overweight or obese⁹. As a critical source of nutrition^{8,10} generating relatively low average
24 environmental pressures^{1,2,11,12}, blue foods present an opportunity to improve nutrition with
25 lower environmental burdens, in line with the Sustainable Development Goals to improve
26 nutrition (Goal 2), ensure sustainable consumption and production (Goal 12), and sustainably use
27 marine resources (Goal 14).

28 Blue foods, however, are underrepresented in food system environmental assessments¹³
29 and the stressors considered are limited⁴ such that we have some understanding of greenhouse
30 gas emissions^{14,15}, but less of others such as land or freshwater use¹⁶. Where blue foods are
31 included, they are typically represented by only one or a few broad categories (e.g., ^{3,17,18}),
32 masking the vast diversity within blue food production. Finally, estimates combining results of
33 published life cycle assessments undertaken for different purposes and consequently employing
34 incompatible methodologies^{19,20}, cannot be compared reliably. It is therefore critical to examine
35 the environmental performance across the diversity of blue foods in a robust, methodologically
36 consistent manner to serve as a benchmark within the rapidly evolving sector as blue food
37 demand increases²¹, production shifts toward aquaculture and production technologies advance.

38 Here, we provide standardized estimates of greenhouse gas (GHG) emissions,
39 consumptive freshwater use (water use), terrestrial land occupation (land use), and nitrogen (N)
40 and phosphorus (P) emissions for blue foods, reported per tonne of edible weight. We identify a
41 set of key life cycle inventory data (i.e., material and energy input and farm-level performance
42 data) from published studies and datasets to which a harmonized methodology is applied. We
43 draw on studies that collectively report data from over 1,690 farms and 1,000 unique fishery
44 records around the world. The 23 species groups represented in our results cover over 70% of

45 global blue food production. We then discuss environmental impacts not covered by the standard
46 stressors, most notably biodiversity loss. Finally, we leverage our model to identify and quantify
47 improvement opportunities and discuss public and private policy options to realize these
48 improvements. In doing so, these results help identify current and future opportunities for blue
49 foods within sustainable diets.

50 *Blue food environmental stressors*

51 Reducing food system GHG emissions is central to meeting global emission targets⁸. Fed
52 aquaculture emissions result primarily from feeds²², while fuel use drives capture fisheries
53 emissions¹¹. Across assessed blue foods, farmed seaweeds and bivalves generate the lowest
54 emissions, followed by small pelagic capture fisheries, while flatfish and crustacean fisheries
55 produce the highest (Fig 1). For fed aquaculture, feed production is responsible for >70% of
56 emissions for most groups (Fig S6). Farmed bivalves and shrimp produce lower average
57 emissions than their capture counterparts (bivalves: 1362 versus 11,400 kg CO₂-eq t⁻¹; shrimps:
58 9090 versus 11956 kg CO₂-eq t⁻¹; results expressed in terms of edible weight), while
59 salmon/trout are similar whether farmed or fished (5173– 5379 versus 6881 kg CO₂-eq t⁻¹).

60 Land use, especially conversion of natural areas, results in a range of context-dependent
61 biodiversity impacts and GHG emissions²³ and creates potential trade-offs with alternate uses,
62 including production of other foods. On-farm land use is low (<1000 m²a t⁻¹; <10%) for most
63 systems and highest (3737–8689 m²a t⁻¹) for extensive ponds (e.g., milkfish, shrimp, and
64 silver/bighead carp). Generally, most land use is associated with feed production for fed systems
65 and explains the overall rankings (Fig 1), though milkfish uses the highest amount of on- and
66 off-farm land.

67 Freshwater increasingly constrains agriculture production but capture fisheries and unfed
68 mariculture require little to no freshwater²⁴. Although blue foods are produced in water,
69 consumptive freshwater use is largely limited to feed production and on-farm evaporative losses
70 for freshwater production¹⁶, with feeds accounting for >80% for all groups apart from carps and
71 catfish (Fig S6). High evaporative losses cause carps to have the highest total water use, while
72 milkfish and miscellaneous marine and diadromous fishes have the highest feed-associated water
73 use. Among fed aquaculture, trout and salmon have the lowest water use, in part attributable to
74 lower crop utilization, highlighting a trade-off with fishmeal and fish oil.

75 Nitrogen and phosphorus emissions are responsible for marine and freshwater
76 eutrophication and are highly correlated due to natural biomass N:P ratios (Table S4). For fed
77 systems, the majority of N (>87%) and P (>94%) emissions occur on-farm. The highest total N
78 and P emissions result from miscellaneous farmed marine and diadromous fishes, milkfish, and
79 fed carp. Non-fed groups such as seaweeds and bivalves, as well as unfed and unfertilized finfish
80 systems (e.g., silver/ bighead carp), represent extractive systems that remove more N and P than
81 is emitted during production, resulting in negative emissions (Fig 1).

82 Across all blue foods, farmed seaweeds and bivalves generate the lowest stressors.
83 However, these groups also highlight several assumptions and nuances. First, bivalve estimates
84 change by nearly five-fold when expressed in terms of edible portion (Fig 1) compared to live
85 weight (Fig S10) due to the shell weight. Second, some processes falling outside our system
86 boundaries represent a potentially large fraction of life cycle emissions for these groups, even if
87 still small in absolute value in some cases. For seaweeds, a large proportion of GHG emissions

88 can occur at the drying stage²⁵ while for bivalves, CO₂ emissions during shell formation²⁶ and
89 high emissions associated with live product from transport²⁷ can be important. Third, impacts on
90 biogeochemical cycling and habitats are highly context dependent. For example, the systems
91 represented here extract nitrogen and phosphorus, which could be problematic in nutrient poor
92 environments. Additionally, ozone effects from volatile short-lived substances depend on the
93 location and varies widely across species^{28,29}. Fourth, sustainable diet recommendations based on
94 these or similar results must account for differences in nutrition content and bioavailability, a
95 particularly important consideration for seaweeds³⁰. Finally, these systems are underrepresented
96 in the literature, particularly for edible seaweeds (Fig S3). As recommendations point toward the
97 potential of these groups, it is important to increase data on these systems, deepen understanding
98 of the above nuances, and be mindful of the total impacts associated with large-scale production
99 on coastal habitats.

100 Capture fisheries, with negligible land, water, N and P values also compare favourably,
101 though groups fall at both the bottom and top of GHG rankings. Among farmed finfish and
102 crustaceans, silver and bighead carps result in the lowest GHG, N, and P emissions, while
103 salmon and trout use the least land and water. To compare with terrestrial foods, we estimated
104 stressors for industrial chicken produced in the US and Europe and find it falls in the middle of
105 farmed blue foods, with similar stressors as tilapia (Fig 1, S14). Because chicken typically has
106 lower stressors than other livestock³, it follows that many blue foods groups compare favourably
107 to other animal-sourced foods. Notably, groups generating among the lowest stressors (e.g.,
108 bivalves and small pelagic fishes) also provide the greatest nutritional quality across all forms of
109 aquatic foods^{2,10}.

110 Our results represent the most comprehensive and standardized blue food stressor
111 estimates to date. Overall, data availability is correlated with global aquaculture production
112 across these taxa groups, but there are still notable taxonomic and geographic gaps (Fig S3, S4).
113 Critically, there are substantial data gaps for silver/bighead carp and aquatic plants given their
114 level of production (Fig S3). Further, our capture fishery data primarily represents commercial
115 marine fisheries³¹. However, subsistence marine and inland catches often utilize non-motorized
116 or no vessels which likely generate few emissions, but there is insufficient data on fuel use
117 across the diversity of small-scale fishing methods to reliably estimate emissions. These systems
118 should be prioritized for additional research. Our estimates represent a snapshot of the
119 knowledge of current production, but future work on emerging production technologies, feed
120 innovations and growing sub-sectors is important for tracking changes against these benchmarks.

121 *From stressors to ecosystem impacts*

122 Emission and resource use stressors are valuable for comparing environmental
123 performance across foods but cannot fully capture final ecosystem and biodiversity
124 consequences (i.e., impacts). Estimating impacts stemming from blue food production requires
125 considering additional stressors and accounting for local context.

126 While GHG, N, P, land, and water are important stressors commonly used to compare
127 foods, other less studied stressors can be critical drivers of ecosystem impacts (Fig 2). Both
128 aquaculture and fisheries may impose other stressors through toxic substance applications (e.g.,
129 antifouling, pesticides in agriculture) and physical disturbance (e.g., bottom trawling, on-bottom
130 culture). Additional stressors include genetic pollution, invasive species introductions³²,

131 application of antibiotics³³, and disease spread³⁴. While capture fisheries have negligible N, P,
132 water and land stressors, other stressors can dramatically alter ecosystems. Fisheries often shift
133 size structure and abundance of targeted species (e.g.,^{35,36}), alter the structure of food webs (e.g.,
134 ³⁷) and impact non-targeted fauna through bycatch³⁸.

135 Local context, such as ecosystem function, carrying/assimilating capacity, and species
136 composition influence how stressors translate into environmental impacts^{39,40}. Notably, land use
137 impacts on biodiversity depend on the land use history and ecological context⁴¹. While all land
138 used for food cultivation represents habitats converted at one point, avoiding additional
139 agricultural expansion is important for preventing further habitat loss⁴². This is also true for on-
140 farm land use by aquaculture, where conversion of ecologically valuable ecosystems, such as
141 mangrove forests²³ that serve as critical carbon sinks⁴³ and nursery habitats, can generate severe
142 impacts. Local species composition and management contexts are also important, including risks
143 associated with marine mammal bycatch (Box 1). Individual stressors may also have nonlinear
144 relationships with impacts or act interactively^{44,45}, such as climate change impacts compounding
145 land use patterns that limit climate refuges or migration options⁴⁶ or resulting in more frequent
146 disease outbreaks, that increase antibiotic use and risk of antibiotic resistance.

147 Capturing the full suite of environmental impacts will require more systematic data
148 collection and methodological advancements. This is crucial for informing policy decisions and
149 realizing the potential contributions of blue foods to sustainable diets while avoiding undesirable
150 trade-offs. Combining local ecological risk and stressor estimates can reveal these important
151 trade-offs, as well as potential synergies (Box 1). Instances of trade-offs complicates
152 sustainability messaging. To this end, while there are no impact-free foods, highlighting
153 synergies simplifies sustainability messaging and helps identify priority interventions.

154 **Box 1 | Emissions and biodiversity risk:** Stressors from life cycle assessments quantify fishery
155 emissions but fail to capture local ecological risks. Combining stressors and impact assessments
156 can illuminate potential sustainability trade-offs. Ecological risk assessments have been
157 developed for capture fisheries to promote holistic assessment of local ecological risks.
158 Integrating GHG emissions with marine mammal risk assessments reveals that some low-GHG
159 emission gears are associated with higher marine mammal risks (e.g., gillnets and entangling
160 nets; Fig 3), while bottom trawls show the opposite. Acknowledging ecological context is critical
161 because risk from similar gears varies across regions. For example, traps and lift nets generally
162 pose low risk to marine mammals (Fig. 3). However, North Atlantic right whales (*Eubalaena*
163 *glacialis*) in the northwest Atlantic are at high risk from entanglements in American lobster
164 (*Homarus americanus*) traps⁴⁷.

165 **Levers for reducing environmental impacts**

166 Variance in stressors indicates diversity across fishing/farming systems (Fig S7-S9) as
167 well as potential “performance gaps.” High variability in milkfish and miscellaneous marine and
168 diadromous fish stressors points to large potential performance gains per unit. This is promising
169 given the interest in marine finfish expansion⁴⁸. Meanwhile, smaller performance gains per unit
170 for high production groups like carps likely generates larger total gains. While some variability
171 within a taxa group is due to differences in on-farm practices, production technology is an

172 important factor across stressors⁴⁹ as variability in stressors for a given species reared in different
173 farming systems can be considerable (e.g.,⁵⁰).

174 We find feed conversion ratios (FCRs) represent the strongest lever, wherein a 10%
175 reduction results in a 1–24% decrease in all stressors (Fig 4a). To evaluate potential shifts under
176 current technology, we estimate the effect of moving each species to the 20th percentile FCR and
177 find the largest reductions for silver and bighead carps (Fig 4b). However, lower FCRs generally
178 come at the cost of larger pond area³³, suggesting a potential trade-off with land and water use.

179 Holding all else constant, a 10% fish production yield improvement (t ha^{-1}) reduces land
180 and water use for freshwater pond systems by 1–9% (Fig 4a). Increasing yields to the 80th
181 percentile reduces land use by 1–49% and water use by 13–51% (Fig 4b). Intensifying
182 production, however, can require more energy for aeration and water pumping as well as
183 increased disease risks with higher animal densities.

184 Feed composition represents another potential lever. Overall, shifting relative proportions
185 of crop- and fish-derived inputs to feeds results in negligible changes in stressors (Fig 4a).
186 Comparing changes in feed sourcing, we found switching to deforestation-free soy and crops
187 reduced GHG emissions by 6–54% (Fig 4b). This could create a co-benefit of also reducing
188 biodiversity impacts. However, as part of integrated global commodity markets, reductions by
189 aquaculture producers will only help meet emissions targets if broader food sector commitments
190 are made. Replacing fish meal and fish oil with fishery by-products has a relatively small effect
191 (Fig 4b), but increased by-product utilization can improve system-wide performance when it
192 directs potential wastes toward more favourable applications⁵¹. Finally, novel aquaculture feeds,
193 including algal, microbe and insect meals, are increasingly available but currently account for a
194 small fraction of feeds. While they likely hold potential to improve feed quality and reduce
195 forage fish demand⁵², their impacts at scale remain uncertain⁵³ and therefore could not be
196 modeled here.

197 For capture fisheries, reducing fuel use represents the primary stressor improvement
198 opportunity. Increasing stock biomass could reduce fuel use per tonne of fish landed^{12,54}, where a
199 13% catch increase with 56% of the effort⁵⁵ corresponds to a 50% reduction in GHG emissions.
200 Alternatively, we find that prioritizing low fuel gears within each fishery can reduce GHG
201 emissions by 4–61%, depending on the species (Fig S16). In some cases, this could create co-
202 benefits for biodiversity impacts (Box 1). Another strategy is to transition fishing fleets to low
203 emission technologies⁸. While some fleets have transitioned to electric, hydrogen fuel and sail-
204 assisted vessels, general adoption necessitates transformations beyond traditional fishery
205 management.

206 **Realizing blue food's environmental potential**

207 Blue foods already have great potential for reducing food system environmental stressors.
208 Unfed aquaculture results in negligible values for most considered stressors, and many fed
209 aquaculture groups outperform industrial chicken, the most efficient major terrestrial animal-
210 source food. Capture fisheries vary widely in their GHG emissions but are low impact with
211 respect to the other stressors considered. This underscores the value of sustainably managing
212 wild fisheries to avoid the environmental replacement cost that would be incurred under fish
213 catch declines²⁴.

214 Our standardized estimates enhance the resolution of the potential role of blue foods
215 within sustainable diets, highlighting opportunities to shift demand from relatively high- to low-
216 stressor blue foods and from terrestrial animal-source foods to comparatively low-stressor blue
217 foods. Shifting to non-animal alternatives remains an efficient lever but low-stressor blue foods
218 may represent an appealing alternative for some consumers. Further, blue foods provide the
219 highest nutrient richness across multiple micronutrients (e.g., iron, zinc), vitamins (e.g., B12),
220 and long-chain polyunsaturated fatty acids (e.g., EPA and DHA) relative to terrestrial animal-
221 source foods¹⁰, which may provide greater incentive to shift demand since consumers generally
222 prioritize seafood freshness, food safety, health, and taste over sustainability⁵⁶.

223 Major challenges remain for shifting demand, as well as meeting increased demand.
224 While improved management offers potential opportunities for expanding some production from
225 low stressor capture fisheries, uncertainty remains around the extent and feasibility of rebuilding
226 many fisheries⁴⁸. Additional research is needed to understand the total environmental impacts of
227 large-scale expansion of low per unit stressor foods, especially for system-specific impacts (Box
228 1). Increasing production also requires creating appropriate incentives and reducing barriers for
229 producers. Historical food system transitions required public investment technologies that could
230 be scaled-up by the private sector and public policy leadership⁵⁷. Overly strict regulations or lack
231 of capital can prevent expansion of low stressor blue foods like offshore mussel farms (e.g., ⁵⁸).
232 Facilitating low-stressor blue food expansion and novel production methods may require new
233 and more adaptive policies and distribution of grants or other forms of start-up capital. Finally,
234 policies can steer production and consumption through taxes and subsidies⁵⁹ as well as softer
235 policies, like dietary advice considering environmental impacts⁶⁰.

236 Within the diversity of blue food production there are numerous opportunities to reduce
237 environmental stressors. As a young and rapidly growing sector, there are many promising
238 technological innovations in aquaculture (e.g., recirculating aquaculture systems, offshore
239 farming and novel feeds). However, less charismatic interventions may represent greater
240 potential for rapid and substantial impact reductions. These include policy or technological
241 interventions that improve husbandry measures (especially reducing disease and mortality) and
242 lower FCRs. Improved management in salmon aquaculture demonstrated considerable
243 sustainability benefits through disease and area management plans⁶¹ and improved stock
244 management with precision aquaculture and automation⁶². Further, selective breeding, genetic
245 improvements and high-quality feeds can all reduce FCRs (Table S8). While we looked at
246 individual interventions, improvements will likely occur through a suit of interventions and the
247 synergistic or antagonistic interactions of interventions represents an important area for future
248 work. Unfortunately, many innovations are often beyond the reach of smallholder producers of
249 low-value species. This highlights a need for public research and development as well as
250 technology transfer to enable all farmers to adopt practices that reduce environmental stressors.
251 For capture fisheries, continued management reforms together with incentives to utilize low fuel
252 gears could substantially improve the performance of capture fisheries^{11,48}. A range of actors will
253 be important for stimulating a shift to more sustainable production methods and, for instance,
254 nation states, civil society and the private sector all have important roles to play. Private sector
255 pre-competitive collaborations, e.g. SeaBOS⁶³ and the Global Salmon Initiative can help
256 stimulate production improvements at scale. Likewise, government-led initiatives helping small-
257 holders improve their farming practices through e.g., access to high quality feeds, seed and
258 broodstock, are crucial for closing the aquaculture performance gap⁶⁴⁻⁶⁶. Certification and
259 improvement projects can help reduce ecosystem impacts⁶⁷, but have been criticised for passive

260 exclusion of small-scale producers. Moving towards best practices like state-led, national
261 certification schemes and area-based approaches will therefore be key⁶⁸. Finally, the finance
262 sector can help steer the sector towards sustainability through strategic investments⁶⁹.

263 The above findings do not suggest unlimited blue food growth is possible nor that
264 expansion comes without environmental trade-offs. Further, without careful consideration for
265 local contexts and inclusion of relevant stakeholders, environmentally focused interventions can
266 generate social and economic trade-offs that undermine broader sustainability goals.
267 Nevertheless, farmed blue food is among the fastest growing food sectors and the global
268 community now faces a unique window of opportunity to steer expansion toward sustainability⁷⁰.
269 Our model and results provide blue food stressor benchmarks and enable data poor
270 environmental stressor assessments. This serves as a critical foundation for understanding blue
271 food environmental performance and to ensuring sustainable and healthy blue foods are available
272 now and into the future.

273 **Main references**

- 274 1. Gephart, J. A. *et al.* The environmental cost of subsistence: Optimizing diets to minimize
275 footprints. *Sci. Total Environ.* **553**, 120–127 (2016).
- 276 2. Hallström, E. *et al.* Combined climate and nutritional performance of seafoods. *J. Clean.*
277 *Prod.* **230**, 402–411 (2019).
- 278 3. Poore, J. & Nemecek, T. Reducing food’s environmental impacts through producers and
279 consumers. *Science* **360**, 987–992 (2018).
- 280 4. Halpern, B. S. *et al.* Opinion: Putting all foods on the same table: Achieving sustainable food
281 systems requires full accounting. *Proc. Natl. Acad. Sci.* **116**, 18152–18156 (2019).
- 282 5. FAO. *The State of World Fisheries and Aquaculture (SOFIA)*. (2020).
- 283 6. FAO. *The state of the world’s land and water resources for food and agriculture (SOLAW) –*
284 *Managing systems at risk*. (Earthscan, 2011).
- 285 7. HLPE. *Food security and nutrition: building a global narrative towards 2030*. 112
286 <http://www.fao.org/3/ca9731en/ca9731en.pdf> (2020).
- 287 8. Willett, W. *et al.* Food in the Anthropocene: the EAT–Lancet Commission on healthy diets
288 from sustainable food systems. *The Lancet* **393**, 447–492 (2019).
- 289 9. Micha, R. *et al.* 2020 Global nutrition report: action on equity to end malnutrition.
290 <https://globalnutritionreport.org/reports/2020-global-nutrition-report/> (2020).
- 291 10. Golden, C. D. Aquatic Foods to Nourish Nations. *Nature* (In Revision).
- 292 11. Parker, R. W. R. *et al.* Fuel use and greenhouse gas emissions of world fisheries. *Nat. Clim.*
293 *Change* **8**, 333–337 (2018).
- 294 12. Hoegh-Guldberg, O. *et al.* *The ocean as a solution to climate change: Five opportunities for*
295 *action*. 116 <http://www.oceanpanel.org/climate> (2019).
- 296 13. Farmery, A. K., Gardner, C., Jennings, S., Green, B. S. & Watson, R. A. Assessing the
297 inclusion of seafood in the sustainable diet literature. *Fish Fish.* **18**, 607–618 (2017).
- 298 14. MacLeod, M. J., Hasan, M. R., Robb, D. H. F. & Mamun-Ur-Rashid, M. Quantifying
299 greenhouse gas emissions from global aquaculture. *Sci. Rep.* **10**, 11679 (2020).
- 300 15. Hilborn, R., Banobi, J., Hall, S. J., Pucylowski, T. & Walsworth, T. E. The environmental
301 cost of animal source foods. *Front. Ecol. Environ.* **16**, 329–335 (2018).
- 302 16. Gephart, J. A. *et al.* The ‘seafood gap’ in the food-water nexus literature—issues surrounding
303 freshwater use in seafood production chains. *Adv. Water Resour.* **110**, 505–514 (2017).
- 304 17. Tilman, D. & Clark, M. Global diets link environmental sustainability and human health.
305 *Nature* **515**, 518–522 (2014).
- 306 18. Springmann, M. *et al.* Options for keeping the food system within environmental limits.
307 *Nature* **562**, 519–525 (2018).
- 308 19. Reap, J., Roman, F., Duncan, S. & Bras, B. A survey of unresolved problems in life cycle
309 assessment: Part 2: impact assessment and interpretation. *Int. J. Life Cycle Assess.* **13**, 374–
310 388 (2008).
- 311 20. Henriksson, P. J. G. *et al.* A rapid review of meta-analyses and systematic reviews of
312 environmental footprints of food commodities and diets. *Glob. Food Secur.* **28**, 100508
313 (2021).
- 314 21. Naylor, R. L. *et al.* Blue Food Demand Across Geographic and Temporal Scales. *Nature* (In
315 Revision).
- 316 22. Henriksson, P. J. G., Pelletier, N. L., Troell, M. & Tyedmers, P. Life cycle assessment and its
317 application to aquaculture production systems. in *Encyclopedia of sustainability science and*
318 *technology* (ed. Meyers, R.) (Springer-Verlag, 2012).

- 319 23. Richards, D. R., Thompson, B. S. & Wijedasa, L. Quantifying net loss of global mangrove
320 carbon stocks from 20 years of land cover change. *Nat. Commun.* **11**, 4260 (2020).
- 321 24. Gephart, J. A., Pace, M. L. & D’Odorico, P. Freshwater savings from marine protein
322 consumption. *Environ. Res. Lett.* **9**, 014005 (2014).
- 323 25. van Oirschot, R. *et al.* Explorative environmental life cycle assessment for system design of
324 seaweed cultivation and drying. *Algal Res.* **27**, 43–54 (2017).
- 325 26. Ray, N. E., O’Meara, T., Williamson, T., Izursa, J.-L. & Kangas, P. C. Consideration of
326 carbon dioxide release during shell production in LCA of bivalves. *Int. J. Life Cycle Assess.*
327 **23**, 1042–1048 (2018).
- 328 27. Iribarren, D., Moreira, M. T. & Feijoo, G. Revisiting the Life Cycle Assessment of mussels
329 from a sectorial perspective. *J. Clean. Prod.* **18**, 101–111 (2010).
- 330 28. Tegtmeier, S. *et al.* Emission and transport of bromocarbons: from the West Pacific ocean
331 into the stratosphere. *Atmospheric Chem. Phys.* **12**, 10633–10648 (2012).
- 332 29. King, G. M. Aspects of carbon monoxide production and oxidation by marine macroalgae.
333 *Mar. Ecol. Prog. Ser.* **224**, 69–75 (2001).
- 334 30. Flores, S. R. L., Dobbs, J. & Dunn, M. A. Mineral nutrient content and iron bioavailability in
335 common and Hawaiian seaweeds assessed by an in vitro digestion/Caco-2 cell model. *J.*
336 *Food Compos. Anal.* **43**, 185–193 (2015).
- 337 31. Parker, R. W. R. & Tyedmers, P. H. Fuel consumption of global fishing fleets: current
338 understanding and knowledge gaps. *Fish Fish.* **16**, 684–696 (2015).
- 339 32. Molnar, J. L., Gamboa, R. L., Revenga, C. & Spalding, M. D. Assessing the global threat of
340 invasive species to marine biodiversity. *Front. Ecol. Environ.* **6**, 485–492 (2008).
- 341 33. Henriksson, P. J. G. *et al.* Unpacking factors influencing antimicrobial use in global
342 aquaculture and their implication for management: a review from a systems perspective.
343 *Sustain. Sci.* **13**, 1105–1120 (2018).
- 344 34. Murray, A. G. Epidemiology of the spread of viral diseases under aquaculture. *Curr. Opin.*
345 *Virol.* **3**, 74–78 (2013).
- 346 35. Myers, R. A. & Worm, B. Rapid worldwide depletion of predatory fish communities. *Nature*
347 **423**, 280–283 (2003).
- 348 36. Svedäng, H. & Hornborg, S. Selective fishing induces density-dependent growth. *Nat.*
349 *Commun.* **5**, 4152 (2014).
- 350 37. Howarth, L. M., Roberts, C. M., Thurstan, R. H. & Stewart, B. D. The unintended
351 consequences of simplifying the sea: making the case for complexity. *Fish Fish.* **15**, 690–711
352 (2014).
- 353 38. Roda, M. A. P. *et al.* *A third assessment of global marine fisheries discards.* (Food and
354 Agriculture Organization of the United Nations, 2019).
- 355 39. Halpern, B. S., Selkoe, K. A., Micheli, F. & Kappel, C. V. Evaluating and Ranking the
356 Vulnerability of Global Marine Ecosystems to Anthropogenic Threats. *Conserv. Biol.* **21**,
357 1301–1315 (2007).
- 358 40. Weitzman, J. & Filgueira, R. The evolution and application of carrying capacity in
359 aquaculture: towards a research agenda. *Rev. Aquac.* raq.12383 (2019)
360 doi:10.1111/raq.12383.
- 361 41. Martin, D. A. *et al.* Land-use history determines ecosystem services and conservation value
362 in tropical agroforestry. *Conserv. Lett.* **13**, e12740 (2020).
- 363 42. Williams, D. R. *et al.* Proactive conservation to prevent habitat losses to agricultural
364 expansion. *Nat. Sustain.* **4**, 314–322 (2021).

- 365 43. Mcleod, E. *et al.* A blueprint for blue carbon: toward an improved understanding of the role
366 of vegetated coastal habitats in sequestering CO₂. *Front. Ecol. Environ.* **9**, 552–560 (2011).
- 367 44. Selkoe, K. A. *et al.* Principles for managing marine ecosystems prone to tipping points.
368 *Ecosyst. Health Sustain.* **1**, 1–18 (2015).
- 369 45. Crain, C. M., Kroeker, K. & Halpern, B. S. Interactive and cumulative effects of multiple
370 human stressors in marine systems. *Ecol. Lett.* **11**, 1304–1315 (2008).
- 371 46. Guo, F., Lenoir, J. & Bonebrake, T. C. Land-use change interacts with climate to determine
372 elevational species redistribution. *Nat. Commun.* **9**, 1315 (2018).
- 373 47. Myers, H. J. & Moore, M. J. Reducing effort in the U.S. American lobster (*Homarus*
374 *americanus*) fishery to prevent North Atlantic right whale (*Eubalaena glacialis*)
375 entanglements may support higher profits and long-term sustainability. *Mar. Policy* **118**,
376 104017 (2020).
- 377 48. Costello, C., Cao, L. & Gelcich, S. The future of food from the Sea. *World Resour. Inst.*
378 *Wash. DC USA 2019–11* (2019).
- 379 49. Bohnes, F. A., Hauschild, M. Z., Schlundt, J. & Laurent, A. Life cycle assessments of
380 aquaculture systems: a critical review of reported findings with recommendations for policy
381 and system development. *Rev. Aquac.* **11**, 1061–1079 (2019).
- 382 50. Bergman, K. *et al.* Recirculating Aquaculture Is Possible without Major Energy Tradeoff:
383 Life Cycle Assessment of Warmwater Fish Farming in Sweden. *Environ. Sci. Technol.* **54**,
384 16062–16070 (2020).
- 385 51. Stevens, J. R., Newton, R. W., Tlusty, M. & Little, D. C. The rise of aquaculture by-
386 products: Increasing food production, value, and sustainability through strategic utilisation.
387 *Mar. Policy* **90**, 115–124 (2018).
- 388 52. Cottrell, R. S., Blanchard, J. L., Halpern, B. S., Metian, M. & Froehlich, H. E. Global
389 adoption of novel aquaculture feeds could substantially reduce forage fish demand by 2030.
390 *Nat. Food* **1**, 301–308 (2020).
- 391 53. Pelletier, N., Klinger, D. H., Sims, N. A., Yoshioka, J.-R. & Kittinger, J. N. Nutritional
392 Attributes, Substitutability, Scalability, and Environmental Intensity of an Illustrative Subset
393 of Current and Future Protein Sources for Aquaculture Feeds: Joint Consideration of
394 Potential Synergies and Trade-offs. *Environ. Sci. Technol.* **52**, 5532–5544 (2018).
- 395 54. Hornborg, S. & Smith, A. D. M. Fisheries for the future: greenhouse gas emission
396 consequences of different fishery reference points. *ICES J. Mar. Sci.* **77**, 1666–1671 (2020).
- 397 55. World Bank. *The Sunken Billions Revisited: Progress and Challenges in Global Marine*
398 *Fisheries*. (Washington, DC: World Bank, 2017). doi:10.1596/978-1-4648-0919-4.
- 399 56. MSC. Understanding seafood consumers.
400 [https://urldefense.com/v3/_https://www.msc.org/understanding-seafood-](https://urldefense.com/v3/_https://www.msc.org/understanding-seafood-consumers_)
401 [consumers_](https://urldefense.com/v3/_https://www.msc.org/understanding-seafood-consumers_);!!IaT_gp1N!m86f1pPqlvfbRn05iW2ohGlRcul1TGj5Njbc-
402 [lQu7mWijd1_DY6oESnGLhUbB8yqn6k\\$](https://urldefense.com/v3/_https://www.msc.org/understanding-seafood-consumers_).
- 403 57. Moberg, E. *et al.* Combined innovations in public policy, the private sector and culture can
404 drive sustainability transitions in food systems. *Nat. Food* **2**, 282–290 (2021).
- 405 58. Fairbanks, L. Moving mussels offshore? Perceptions of offshore aquaculture policy and
406 expansion in New England. *Ocean Coast. Manag.* **130**, 1–12 (2016).
- 407 59. Säll, S. & Gren, I.-M. Effects of an environmental tax on meat and dairy consumption in
408 Sweden. *Food Policy* **55**, 41–53 (2015).

- 409 60. Fischer, C. G. & Garnett, T. *Plates, Pyramids, Planet - Developments in national healthy*
410 *and sustainable dietary guidelines: a state of play assessment*. (Food and Agriculture
411 Organization of the United Nations, 2016).
- 412 61. Jones, S., Bruno, D., Madsen, L. & Peeler, E. Disease management mitigates risk of
413 pathogen transmission from maricultured salmonids. *Aquac. Environ. Interact.* **6**, 119–134
414 (2015).
- 415 62. Antonucci, F. & Costa, C. Precision aquaculture: a short review on engineering innovations.
416 *Aquac. Int.* **28**, 41–57 (2020).
- 417 63. Österblom, H., Jouffray, J.-B., Folke, C. & Rockström, J. Emergence of a global science–
418 business initiative for ocean stewardship. *Proc. Natl. Acad. Sci.* **114**, 9038–9043 (2017).
- 419 64. Watson, J. R., Armerin, F., Klinger, D. H. & Belton, B. Resilience through risk management:
420 cooperative insurance in small-holder aquaculture systems. *Heliyon* **4**, e00799 (2018).
- 421 65. Hasan, M. R. On-farm feeding and feed management in aquaculture. *FAO Aquac. Newsl.* 48
422 (2010).
- 423 66. Bondad-Reantaso, M. G. *Assessment of freshwater fish seed resources for sustainable*
424 *aquaculture*. (Food & Agriculture Org., 2007).
- 425 67. Gutiérrez, N. L. *et al.* Eco-Label Conveys Reliable Information on Fish Stock Health to
426 Seafood Consumers. *PLoS ONE* **7**, e43765 (2012).
- 427 68. Bush, S. R. *et al.* Inclusive environmental performance through ‘beyond-farm’ aquaculture
428 governance. *Curr. Opin. Environ. Sustain.* **41**, 49–55 (2019).
- 429 69. Jouffray, J.-B., Crona, B., Wassénius, E., Bebbington, J. & Scholtens, B. Leverage points in
430 the financial sector for seafood sustainability. *Sci. Adv.* **5**, eaax3324 (2019).
- 431 70. Gephart, J. A. *et al.* Scenarios for Global Aquaculture and Its Role in Human Nutrition. *Rev.*
432 *Fish. Sci. Aquac.* **29**, 122–138 (2021).

433

434 **Fig. 1 | Stressor posterior distributions.** Panels represent a) Aquaculture GHG emissions (kg
435 CO₂-eq t⁻¹); b) Aquaculture N (kg N-eq t⁻¹); c) Aquaculture P (kg P-eq t⁻¹); d) Capture GHG
436 emissions (kg CO₂-eq t⁻¹); e) Aquaculture Water use (m³ t⁻¹); f) Aquaculture Land use (m²a t⁻¹).
437 Values represent tonnes of edible weight and use mass allocation. Dot indicates the median,
438 colored regions show credible intervals (i.e., range of values that have a 95% (light), 80%, and
439 50% (dark) probability of containing the true parameter value). Taxa group names are
440 abbreviated ISSCAAP names (See Table S3 for definitions). Beige bands represent estimated
441 chicken minimum to maximum range. See Fig S10 for estimates expressed in terms of live
442 weight.

443 **Fig. 2 | Major stressors stemming from aquaculture and capture fisheries.** Icons with
444 magenta border are quantified in this study while the others are discussed qualitatively.

445 **Fig. 3 | GHG emissions compared to marine mammal risk.** Data represent fisheries in Europe
446 (NE Atl) and Central America (C Am SSF) by gear type. Dot indicates the median estimate of
447 the mean kg CO₂-eq t⁻¹ and intervals show 95% (light), 80%, and 50% (dark) credible intervals.
448 Risk index is the sum of the number of marine mammals at risk times 3, 2 or 1 for high, medium,
449 and low risk, respectively.

450 **Fig. 4 | Aquaculture stressor intervention opportunities** a) Change (%) in each stressor
451 associate with a 10% reduction in the parameter value (black cell indicates stressor change
452 >20%); b) Change (%) in each stressor under four scenarios (defined in Table S8) relative to the

453 current estimate. Arrows indicate changes greater than 50%. Additional aquaculture scenario
454 results displayed in Fig S15 and capture scenario results in Fig S16.

455

456 **Data and Code Availability**

457 All data and code used to produce the results of our analysis are available in the supplementary
458 information and on GitHub (<https://github.com/jagephart/FishPrint>), with the published version
459 archived (DOI: 10.5281/zenodo.4768324).

460

461 **Acknowledgements**

462 This paper is part of the Blue Food Assessment (<https://www.bluefood.earth/>), a comprehensive
463 examination of the role of aquatic foods in building healthy, sustainable, and equitable food
464 systems. The assessment was supported by the Builders Initiative, the MAVA Foundation, the
465 Oak Foundation, and the Walton Family Foundation, and has benefitted from the intellectual
466 input of the wider group of scientists leading other components of the BFA work. JAG, KDG,
467 and CDG were supported by funding under NSF 1826668. AS was supported by a grant from
468 The Nature Conservancy. Funding for participation of SH, KB; MT, PH and FZ came from
469 Swedish Research Council Formas (Grants 2016-00227 and 2017-00842). This work was
470 financially supported, in part, by the Harvard Data Science Initiative.

471 **Author contributions**

472 JAG, PJGH, RP and AS contributed equally to the study. JAG and CG organized the initial
473 workshop. JAG, PJGH, RP, AS, GE, CDG, PT and MT conceived of the idea and designed the
474 overall study. JAG, PJGH, RP, AS, KDG, SH, KM, MM, RN, WZ, and FZ compiled the data.
475 JAG and KDG developed the model and analysed the data. All authors reviewed the results and
476 contributed to and approved the final manuscript.

477

478 **Additional information**

479 Supplementary Information is available for this paper.

480

481 The authors declare the following competing interests: RWRP became employed by the
482 Aquaculture Stewardship Council while this manuscript was under consideration.

483

484 Correspondence and requests for materials should be addressed to Jessica Gephart.

485

486 Reprints and permissions information is available at www.nature.com/reprints

487